

development over the 10 wks before term equivalent age in pre-term infants.

Methods 35 infants (GA: 27.1 ± 0.7 ; BW: 937 ± 172) without morphine, were monitored with EEG/aEEG. Three periods were selected at 20–24 h, 32–36 h, 44–48 h. Minimum amplitude,% of time

Results Increased SATrate was positively associated with deltaGMv, inner and outer surface (resp β :7.4, p :0.001; β :46.6, p :0.002; β :57.5, p :0.001). Consistent with these findings, ISI was negatively associated with changes in GMv, inner and outer surface (β :-3.4, p : 0.007; β :-17.8, p : 0.034; β :-27.7, p : 0.006). Min aEEG and% of time <5 μ V were associated with inner and outer surface at 40 wks (respectively: β :46.2, p :0.043; β :53.0, p :0.041; and β :-2.9, p :0.025; β :-3.5, p :0.019). No effect on thickness and gyrification was found.

Conclusions Early brain activity seems to be associated with cortical development suggesting that adequate brain activity in the early neuronal networks is necessary to lead to growth and development of neonatal cerebral cortical brain, measured by structural MRI.

0-063 DATA QUALITY IN DIFFUSION TENSOR IMAGING STUDIES OF THE PRETERM BRAIN

¹K Pieterman, ¹A Plaisier, ¹P Govaert, ²A Leemans, ³M Lequin, ¹J Dudink. ¹Neonatology, Erasmus University Medical Center - Sophia Children's Hospital, Rotterdam, Netherlands; ²Image Sciences Institute, University Medical Center Utrecht, Utrecht, Netherlands; ³Radiology, ErasmusMC - Sophia Children's Hospital, Rotterdam, Netherlands

10.1136/archdischild-2014-307384.131

Background and aims Quantitative measurement of brain maturation is increasingly performed in preterm infants using diffusion tensor imaging (DTI). To study white matter properly, reliability of underlying DTI data is of paramount importance, as acquisition and processing steps can substantially affect DTI analyses. We systematically reviewed literature to raise awareness regarding these matters.

Methods We systematically reviewed studies published between 1991 and September 2013, in which DTI scanning of preterm infants was performed within 28 days after term-equivalent age. Based on our inclusion criteria, 75 preterm DTI studies were considered relevant and further analysed. We primarily focused on use of dedicated neonatal equipment, DTI acquisition parameters and processing methodology.

Results There was wide variation among different studies in acquisition and processing methodology, and frequently incomplete reporting of these settings. 25.3% reported the use of dedicated neonatal equipment. Data quality assessment was not reported in 34.7%. Correction for artefacts and exclusion of datasets was not reported in 45.3% respectively 30.7%. Only 54.7% of the studies reported specific correction methods. Tensor estimation methodology was reported in 82.7%. Fast but less accurate tensor calculation algorithms were applied more frequently than advanced algorithms.

Conclusion

DTI acquisition and processing settings are described incompletely in current literature, and vary considerably among different neonatal DTI research groups. In addition, described settings do frequently not meet the highest standards possible. Hence the premature population should be regarded as one of the most

challenging groups to image using DTI, maximal awareness regarding these matters is a prerequisite.

Neonatal Immunity

0-064 DEVELOPMENT AND MATURATION OF MAIT CELLS IN HUMAN NEONATES: RELATIONS WITH GESTATIONAL AGE AND MICROBIAL INFECTION

¹V Biran, ²G Ben Youssef, ²M Tourret, ³V Houdoin, ⁴O Baud, ²S Caillat-Zucman. ¹Neonatal Intensive Care Unit, Robert Debre Hospital, Paris, France; ²Immunology, Robert Debre Hospital, Paris, France; ³Pneumology, Robert Debre Hospital, Paris, France; ⁴Neonatal Intensive Care Unit, Robert Debre Hospital, Paris, France

10.1136/archdischild-2014-307384.132

Newborns, in particular preterm neonates, suffer a high frequency of microbial infections. MAIT cells are innate-like T cells expressing a semi-invariant V α 7.2-J α 33 TCR which recognises MR1-restricted, microbial-derived riboflavin (vitamin B2) metabolites unique to bacteria and yeast.

We studied 151 newborns admitted in the Neonatology Department at Robert Debré Hospital divided into four groups according to gestational age (group 1: 24–27 wks; group 2: 28–31 wks; group 3: 32–36 wks; group 4: >37 wks). The rate and kinetics of MAIT cell expansion and maturation were determined longitudinally at birth (day 0), day 3, day 30 and day 60. We performed multiparametric 10-colour flow cytometry analyses using combinations of antibodies to CD45, CD3, CD4, CD8, TCR V α 7.2, CD161, CD45RA, TCR V α 24 and TCR $\gamma\delta$ on 100 ml residual whole blood (left over of blood count), allowing characterisation of MAIT cells in parallel with other non-conventional and conventional T cells.

Our results show that the frequency of MAIT at birth is low and significantly differs according to gestational age (median at D0 group 1: 0.21%; group 2: 0.14%; group 3: 0.12%; group 4: 0.06%).

Of note, this frequency remains relatively stable over the first 2 months of life. However, the phenotype of MAIT cell changes after birth with rapid maturation and increased proportion of CD8aa cells. Significant difference was observed between high preterm neonates with and without maternofetal infection. Analysis of MAIT cell frequency in 20 twin pairs showed it was very similar, suggesting that it might be controlled by a genetic and/or early environmental factor.

In conclusion, the frequency of MAIT cells at birth is inversely correlated with gestational age, and is correlated with the presence of maternofetal microbial infection in preterm neonates. Whether it may reflect the presence of microbial products in amniotic liquid, and/or differences in the gut microbiota immediately after birth is under investigation.

0-065 MACROPHAGE MIGRATION INHIBITORY FACTOR BALANCES NEONATAL INNATE IMMUNE RESPONSES

¹T Roger, ²M Weier, ²A Schneider, ³Fcgj Sweep, ⁴J Bernhagen, ¹T Calandra, ²E Giannoni. ¹Infectious Diseases Service, Centre Hospitalier Universitaire Vaudois and University of Lausanne, Lausanne, Switzerland; ²Service of Neonatology and Infectious Diseases Service, Centre Hospitalier Universitaire Vaudois and University of Lausanne, Lausanne, Switzerland; ³Department of Laboratory Medicine, Radboud University, Nijmegen, Netherlands; ⁴Institute of Biochemistry and Molecular Cell Biology, Aachen University, Aachen, Germany

10.1136/archdischild-2014-307384.133