Dysoxia and lactate

Trevor Duke

Disturbances of oxygen supply or cellular oxygen metabolism are common in critically ill patients. An understanding of dysoxia, or oxygen limited energy depletion, requires an understanding of the normal physiology of cell oxygen metabolism, and the compensatory mechanisms that supply high energy molecules under conditions of hypoxia. Understanding disorders associated with hyperlactataemia requires consideration of the cellular response in dysoxia, and pathology specific derangements in lactate metabolism. Much has recently been discovered about the causes of lactate acidosis in sepsis, and about the role of lactate in monitoring critically ill children.

This review discusses how cells produce energy for metabolism under normal and hypoxic conditions; what happens to lactate produced during these processes; the clinical situations in which lactic acidosis has been described; the reasons why excess lactate may occur in sepsis; the evidence that a high lactate concentration is not simply a surrogate for tissue dysoxia; the relevance of lactate in the management of critically ill children; and suggested strategies to manage high blood lactate.

Normal cellular oxygen metabolism

Cells require oxygen for the production of ATP, the principal energy source. ATP is hydrolysed to ADP and high energy phosphate by adenosine triphosphatasases in the cytosol. Energy released is used for the maintenance of membrane integrity, ionic pumps, and other specialised functions, such as contractility of muscle cells, and impulse transmission in neurons. The body's stores of ATP will last no more than a few minutes, so it must be synthesised continuously as it is being used. Under physiological conditions, most ATP is generated from the metabolism of glucose, by the process of oxidative phosphorylation. The first stage of oxidative phosphorylation is the conversion of glucose to pyruvic acid; this occurs in the cytoplasm. The second stage, the oxidation of pyruvic acid, can only occur in the mitochondria as part of the Krebs (citric acid) cycle. Oxidative phosphorylation produces a net 36 molecules of ATP (or 1270 kJ of available energy) for every glucose molecule oxidised.

Under normal conditions some tissues, such as myocytes, preferentially use free fatty acids, rather than glucose, as the substrate for ATP generation. In myocytes, palmitate forms about 60% of the total substrate metabolised, glucose 10%, and lactate 30%. During cellular hypoxia the consumption of lactate ceases, and that of glucose increases to 90% of the total substrate consumed. The brain, on the other hand, under any conditions can use only glucose or ketone bodies for ATP production.

Thresholds of hypoxia and cell (dys)function

Oxidative phosphorylation can only occur when the partial pressure of oxygen (PO₂) within the mitochondrion is above a critical level, thought to be in the order of 1 mm Hg (0.13 kPa). Although the PO₂ of dry air at sea level is 159 mm Hg, and the alveolar PO₂ is about 100 mm Hg, the mean capillary blood PO₂ is 50 mm Hg. From the capillaries oxygen diffuses down a further concentration gradient from one cell to another, and within cells, such that normal mitochondrial PO₂ is in the order of 4–20 mm Hg (0.5–2.7 kPa). Some cells, such as the centrilobular cells in the liver, are particularly susceptible to hypoxia because they are further away from capillaries. These undergo necrosis early under hypoxic conditions.

Connett et al defined three theoretical thresholds of cell hypoxia. The first threshold is crossed when cell oxygen decreases but ATP production is maintained at a level sufficient to match ATP demand by metabolic adaptation. Adaptation involves recruitment of the redox component of electron transport, changes in the phosphorylation states of mitochondria, and increased glycolysis. The critical level of mitochondrial PO₂ for oxidative phosphorylation depends on the cells ability to adapt the phosphorylation process metabolically, and the level of ATP demand.

The second threshold occurs when steady state ATP turnover can only be maintained by supplementary production of ATP from anaerobic glycolysis, by the Embden-Meyerhof pathway. This energy inefficient mechanism for producing high energy molecules generates only two molecules of ATP (67 kJ of available energy) for every molecule of glucose metabolised. The pathway must therefore either consume relatively larger quantities of glucose or, alternatively, yield much less ATP. In high energy consuming organs, such as the brain,
kidney, and liver, the rapid transfer of such quantities of glucose across cell membranes is not possible. Therefore, these organs develop ATP depletion rapidly under hypoxic conditions. Dysoxia can be defined below the second threshold, where ATP production becomes oxygen limited. The third threshold is crossed when glycolysis becomes insufficient to produce enough ATP to maintain cell function and structural integrity.

Most data on cell dysfunction in hypoxia are from in vitro studies; mechanisms are complex and poorly understood, and application to clinical pathophysiology is speculative. Technology, such as nuclear magnetic resonance spectroscopy, which will non-invasively measure cell biochemical changes, is as yet impractical in acute clinical settings.

The effects of ATP depletion on cell function can be divided into those occurring as a result of the initial hypoxic insult, and those that result from reperfusion or reoxygenation. Depletion of ATP results in loss of function of sodium/potassium (Na+/K+)-channels, leading to accumulation of intracellular Na+. Because cell membrane synthesis is a continuous energy requiring process, ATP depletion leads to loss of cell membrane integrity. This, and the accumulation of intracellular Na+, results in cell swelling. Similarly, there is disruption of lysosomal membranes and release of enzymes into the cytosol, causing autolysis. Massive influx of Na+ inhibits or reverses the sodium/calcium (Na+/Ca++) ion channel, and is one mechanism leading to the intracellular sequestration of Ca++. Intracellular Ca++ increases during the first 30 minutes of hypoxia, but can decrease to normal when hypoxia is of longer duration. However, during reperfusion there is a pronounced increase in cytosol Ca++. Ca++ activated proteases can destroy the sarcolemma and the cytoskeleton; Ca++ activated phospholipases further degrade membrane phospholipids. Concurrent with the depletion of intracellular ATP there is an accumulation of AMP. AMP is released from the cell into the interstitium, where it is dephosphorylated rapidly to adenosine. Adenosine acts as a vasodilator in most vascular beds, improving blood flow in capillaries, but acts as a vasoconstrictor in the renal vascular bed. During reoxygenation, adenosine is a source of oxygen radicals through the formation of inosine, hypoxanthine, and xanthine. Oxygen radicals cause cell damage by lipid peroxidation, release of excitatory amino acids, and inhibition of enzymes.

**Lactate production and elimination**

Anaerobic metabolism is not only energy inefficient and unsustainable, but it produces two molecules of lactic acid for every molecule of glucose metabolised. Although lactic acid has energy potential, it is only in the presence of oxygen that it can be reconverted into pyruvic acid, which in turn can be metabolised in the citric acid cycle. Lactic acid can be converted into glucose, by the process of gluconeogenesis (Cori cycle, requiring six molecules of ATP), and stored in the liver as glycogen. This is also an ATP requiring process.

Under normal conditions, the liver has a large capacity for lactate removal, and other organs including the kidneys, gastrointestinal tract, and muscle also remove lactate. Lactate production has to be substantially increased before the metabolic threshold of the liver and other organs is exceeded, and increased blood concentrations occur. Lactate accumulation depends on the rate of glycolysis, the exchange of lactate across cell membranes, washout by the circulation, and consumption and clearance by tissues. Lactate clearance by the liver and other organs may be reduced in sepsis and in respiratory alkalosis.

Other conditions interfere with lactate production, so that despite tissue hypoxia, blood lactate will be normal. In severe malnutrition—for example, glucose stores are insufficient to sustain glycolysis. Figure 1 outlines the mechanisms of lactate production and elimination.

Most plasma cell membranes contain a symport for lactate and hydrogen ions, and lactic acid produced in cells is released into the circulation. In the brain, however, excess lactate accumulates in neurons, because the blood–brain barrier is relatively impermeable to charged ions. In severe cerebral hypoxia, some of the brain tissue injury might be caused by intracellular acidosis, in addition to the depletion of high energy compounds.

**The clinical relevance of high blood lactate**

High lactate concentrations are seen in the setting of circulatory shock as a result of haemorrhage, myocardial failure, burns, and sepsis; after cardiopulmonary bypass; and after liver transplantation. Lactic acidosis might exist not only in the presence of hypotension...
Dysoxia and lactate

and low cardiac output, but also in normoten-
sive patients with a normal or high cardiac
output.\(^{21,22}\) Situations where the rate of glycol-
sis exceeds the rate of pyruvate utilisation by
the Krebs cycle include alkalosis, and excess
catecholamine release, and are also associated
with hyperlactataemia.\(^{23}\) Drugs and chemicals
that interfere with gluconeogenesis, such as
ethanol, methanol, ethylene glycol, salicylates,
and phenformin, can also raise lactate
concentrations.\(^{24}\) Congenital defects of mito-
chondrial function also present with hyperlac-
tataemia.

The association of lactic acidosis with
increased mortality in critically ill patients is
well recognised.\(^{16,24,25}\) In a heterogeneous

and 126 severely ill adults with metabolic
acidosis, defined as a lactate $\geq 5$ mmol/l, and
either an arterial pH $\leq 7.35$ or a base deficit
$> 6$ mmol/l, Stacpoole and colleagues\(^{11}\) found
higher lactate concentrations in non-survivors
(mean, 12.2; SD, 5.9 mmol/l) than survivors
(mean, 9.2; SD, 4.9 mmol/l; \(p = 0.004\)). Less
than 17% of the patients in this series survived
to hospital discharge.\(^{13}\)

Mild hyperlactataemia with respiratory alka-
losis might not have the same adverse prognos-
tic value as hyperlactataemia with acidosis.
Some authors have proposed that mild rises in
lactate are not indicative of serious pathology,
depending on the mechanism involved.\(^{9,20}\)

In critically ill patients, there is some value in
the trend of change in lactate as a means of
assessing response to treatment, and
prognosis.\(^{15,18,27,29}\) Vincent \textit{et al} described the
time course of blood lactate in adults who
responded to rapid volume resuscitation for
circulatory shock.\(^{39}\) In all cases, during the first
hour there was at least a 10% reduction in
blood lactate. This contrasted with patients
who died during circulatory shock, in whom
lactate concentrations did not change with
resuscitation. In a study of adults with sepsis,
Tuchschild and colleagues\(^{10}\) observed that
despite the similarity in oxygen consumption
(V\(_{O_2}\)) after the initial resuscitation period
among survivors and non-survivors, lactate
concentrations decreased in the survivors, but
not in those who died.

Cause of hyperlactataemia in sepsis

The traditional distinction between type A
(where there is clinical evidence of tissue
hypoxia) and type B lactic acidosis (where
there is no clinical evidence of tissue hypoxia)
proposed by Cohen and Woods\(^{31}\) is simplistic in
sepsis. Several mechanisms are involved. Clin-
ical application of this distinction also implies
that there is a valid gold standard for the
measurement of tissue hypoxia, which is argu-
able.

Tissue hypoxia can occur because of a
decrease in oxygen delivery in hypodynamic
sepsis. Sepsis with low cardiac output and
peripheral vasoconstriction might be more
common in young children, and is a well
described feature of severe meningococcal
sepsis.\(^{32,33}\) Even a normal or raised cardiac
output might not supply sufficient oxygen to
tissues that have an increased metabolic rate.

Hypoxia induced hyperlactataemia might also
occur because of the formation of peripheral
arterio-venous shunts, or the regional redistrib-
ution of blood away from the hepatosplanch-
nic vascular bed. The true role of impaired cel-
lar oxygen delivery (DO\(_2\)) in sepsis is
uncertain, however, because covert tissue
hypoxia might not be a common antecedent for
lactic acidosis or adverse sequelae in sepsis.\(^{34}\)

Although there is some evidence for a
relation between delivery dependent oxygen
consumption and lactate, this has not been
shown in all studies, and cannot be generalised
to all septic patients. In support of lactate
reflecting delivery dependent oxygen
consumption, Gilbert and colleagues\(^{35}\) showed that
patients with a high lactate had significant
increases in V\(_O_2\), in response to colloid fluid
volume expansion and packed red cell transfu-
sion, whereas patients with normal lactate had
no such increase in V\(_O_2\). Against these findings
are those of Silverman,\(^{14}\) who used intravenous
fluids, packed red cell transfusion, and do-
butamine infusion to improve DO\(_2\) in 17 adults
with sepsis. Of 52 interventions that improved
DO\(_2\) and V\(_O_2\) in the 17 patients, about one
third were associated with an unchanged, or
increased, blood lactate. There was no predict-
able relation between improvements in V\(_O_2\)
and reductions in blood lactate.

Further evidence has been used to argue
against tissue hypoxia as the major cause of
lactic acidosis, namely: (1) the effect of sodium
dichloroacetate (DCA) on lactate concentra-
tions in sepsis\(^{36}\); (2) the finding that in patients
with mild rises of blood lactate, clearance is
reduced rather than lactate production
increased; (3) in clinical studies, efforts to
drive DO\(_2\) to supranormal levels do not
consistently improve outcome\(^{37}\) or lower blood
lactate; (4) in some models of sepsis, organ
failure occurs despite levels of V\(_O_2\) and DO\(_2\)
within the physiological range\(^{38}\); and (5) in
some studies, tissue oxygen tension has been
measured in septic patients and found not to be
low.

DCA, in the presence of oxygen, stimulates
pyruvate dehydrogenase activity and increases
oxidation of pyruvate to acetyl coenzyme A and
carbon dioxide, thereby increasing pyruvate
and lactate metabolism. DCA reduces blood
lactate in septic patients. Some have argued,
therefore, that deficits in cellular oxygen
cannot be the rate limiting step in lactate
accumulation,\(^{39}\) but instead there is increased
pyruvate production or decreased lactate
clearance.\(^9\) Lactate reductions in septic pa-
tients treated with DCA are modest, however.
In one study, with of a pretreatment mean
value of 11.6 mmol/l, the reduction was
between 1 and 3 mmol/l, and one third of
patients had less than a 20% decrease in blood
lactate concentration.\(^{35}\) There was still an
excess of lactate that was not eliminated by this
DCA induced aerobic process, which might be
attributable to cell hypoxia.

The absence of an apparent delivery depend-
ent relation between DO\(_2\), V\(_O_2\), and lactate
does not exclude dysoxia as the cause of high
lactate in sepsis. Supply dependent oxygen
consumption at a tissue level might be poorly detected by the global values of VO$_2$ and DO$_2$, even if independent and valid methods are used to measure each variable. It is untested whether regional measures of dysxia, such as the hepatic venous β-hydroxybutyrate to acetacetoate ratio, would better determine whether supply dependent oxygen consumption exists in sepsis, and how sensitive high blood lactate is at reflecting this.

If tissue hypoxia is not the major cause of lactate accumulation in sepsis, then what other mechanisms are involved? Levraut et al showed that haemodynamically stable adults with sepsis and mildly raised blood lactate had more than a 50% reduced lactate clearance compared with septic adults who had normal lactate, whereas lactate production was similar in the two groups. Reduced activation of the pyruvate dehydrogenase complex in sepsis might cause the accumulation of pyruvate and formation of lactate.

Although mitochondrial PO$_2$ might be normal in sepsis, theoretically, ATP production would be reduced by impaired delivery of pyruvate into the Krebs cycle, inhibition of mitochondrial enzymes in the Krebs cycle or electron transport chain, or other disruption of mitochondrial function. What has been termed “cytopathic hypoxia” might really be a cytopathic inability to use available oxygen. This is well defined—for example, in cyanide poisoning, where cytochromes are inhibited and aerobic metabolism ceases. Whether a similar mechanism operates in clinical sepsis, what causes it, and whether it contributes to hyperlactataemia are uncertain.

**Lactate and prediction of outcome in critically ill children**

Whatever the causes of hyperlactataemia, high or rising blood lactate might help identify children at high risk of mortality.

**SEPSIS**

We prospectively compared the predictive value of several variables for mortality and major sequelae in 31 children admitted to the intensive care unit (ICU) with severe sepsis. Those variables were: mean arterial pressure, heart rate, arterial pH, base deficit, gastric intramucosal pH, and blood lactate. These were measured at the time of admission and 12, 24, and 48 hours later. There were 10 deaths and 21 survivors. Blood lactate was the earliest predictor of mortality, and identified survivors from those who subsequently died, when measured as early as 12 hours after admission. Twelve hours after admission a blood lactate > 3 mmol/l had a positive predictive value for death of 56%, and a lactate of 3 mmol/l or less had a positive predictive value for survival of 84%. At 24 hours, a lactate > 3 mmol/l had a positive predictive value for death of 71%, and a lactate of 3 mmol/l or less had a positive predictive value for survival of 86%. No other variable identified deaths from survivors as early as 12 hours.

**CARDIAC SURGERY**

Siegel and colleagues measured arterial blood lactate in 41 children after cardiopulmonary bypass. The mean (SD) lactate concentration for survivors was significantly lower than for the seven who died (2.38 (0.13) mmol/l v 6.86 (0.79) mmol/l). Higher lactate concentrations on admission to the ICU were significantly associated with an increased number of extracardiac organ systems failing in the postoperative period, and were positively correlated with total bypass and circulatory arrest times.

We compared the predictive power of markers of cardiovascular function for clinically important major postoperative adverse events in a prospective cohort study of 90 children after cardiopulmonary bypass. Heart rate, blood pressure, cardiac output by thermodilution, oxygen delivery, base deficit, mixed venous oxygen saturation, blood lactate, gastric intramucosal pH, and toe core temperature gradient were measured at the time of admission to the ICU, and every four hours thereafter. We also recorded the duration of cardiopulmonary bypass, aortic cross clamping, and circulatory arrest. The major postoperative adverse events of interest were cardiac arrest, need for emergency chest opening, need for commencement of extracorporeal life support, development of multiple organ failure, and death. The subjects were selected for their high risk for major postoperative adverse events, and included 23 neonates having the arterial switch procedure, and 23 infants undergoing repair of multiple ventricular septal defects. In the series, 12 children had major postoperative adverse events and three died. The variables that were the earliest predictors of major postoperative adverse events were the duration of cardiopulmonary bypass, the blood lactate at the time of admission to the ICU, and admission hypotension. Blood lactate was the most consistent predictor of major postoperative adverse events, identifying subsequent major postoperative adverse events when measured also at four and eight hours after admission. Gastric intramucosal pH was also predictive of major postoperative adverse events, but was a later predictor than lactate. At no time in the first 24 hours after ICU admission were cardiac output, oxygen delivery, mixed venous oxygen saturation, or the toe core temperature gradient predictive of major postoperative adverse events. Lactate might be a useful indicator of children at risk of major morbidity and mortality after cardiac surgery. Of note, a high lactate did not predict more than one third of all major postoperative adverse events so, although a specific marker of risk of such events, it had poor sensitivity.

**NEONATAL INTENSIVE CARE**

Hyperlactataemia has been described as an early marker of sepsis and necrotising enterocolitis in preterm neonates. Deshpande et al measured lactate, arterial pH, and base excess in 75 mechanically ventilated newborns, most of whom were premature. They found significantly higher blood lactate in those who died compared with survivors. After a...
Dysoxia and lactate

347

...and might further depress myocardial function. However, the evidence that bicarbonate administration causes paradoxical intracellular acidosis is contentious. In cell preparations, the effect on intracellular pH of adding bicarbonate to extracellular fluid depends on the starting pH of the system, the size of the bicarbonate boluses added to the system, and the type of buffering solutions used as the extracellular suspension. In vitro studies that have reproduced metabolic acidosis, which used physiological solutions buffering the extracellular fluid, and added bicarbonate as a series of small boluses, have shown only small and transient reductions in intracellular pH.

What of the effect of bicarbonate on myocardial function? Two canine studies found transient increases in coronary venous CO₂ and decreases in myocardial contractility at the beginning of an infusion of sodium bicarbonate. This was followed by improved contractile function beyond baseline levels. One study documented a reduction in intracellular pH, whereas the other study showed no change in intracellular pH, but a decrease in serum free Ca²⁺, coinciding with myocardial depression. These investigators proposed that myocardial depression after bicarbonate infusion might be caused by reduced availability of free Ca²⁺, rather than intracellular acidosis.

The effect of bicarbonate on the intracellular pH in other organs is similarly contentious. In a rabbit model of hypoxic lactic acidosis, brain intracellular acidosis was partially corrected by bicarbonate administration; however, in a study of five normal adult humans, intravenous sodium bicarbonate resulted in a reduction in brain intracellular pH, measured by magnetic resonance imaging spectroscopy. Despite all this, it is not known whether the therapeutic correction of intracellular acidosis is without adverse effects, and one experimental study shows that intracellular acidosis might protect again reperfusion injury.

It has been proposed that the effect of bicarbonate in shifting the haemoglobin-oxygen dissociation curve to the left will result in delayed tissue reoxygenation. In a porcine model of haemorrhagic shock, bicarbonate delayed increases in tissue PO₂ and delayed falls in plasma lactate. This study used small patient numbers, and repeated measures over time, but analysed differences in the data at time intervals that were not predetermined. Therefore, the conclusions are not strongly supported.

Clinical trials of bicarbonate in children are few. Fanconi et al., in an open non-randomised, “before and after” trial in neonates with metabolic acidosis, examined the cardiovascular effects of bicarbonate infusion. Sodium bicarbonate induced a transient increase in cardiac output and a fall in systemic vascular resistance. Although this appears at odds with the animal models described above, from this clinical study there is no way of telling whether the effect was because of bicarbonate correction of acidosis, or because...
ICU admission lactate > 5 mmol/l has a 30% risk of major adverse events. Lactate at four hours

**Table 2 Protocol to manage high blood lactate after cardiopulmonary bypass**

<table>
<thead>
<tr>
<th>Step</th>
<th>Action</th>
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<tbody>
<tr>
<td>1.</td>
<td>Give intravenous fluid volume bolus of 10–20 ml/kg of colloid or normal saline</td>
</tr>
<tr>
<td>2.</td>
<td>Continuously monitor arterial blood pressure, central venous pressure (CVP), heart rate, and cutaneous oxygen saturation (SpO₂)</td>
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<tr>
<td>3.</td>
<td>Maintain CVP at 8–12 mm Hg with fluid volume up to 40–60 ml/kg might be required in the first six hours</td>
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<tr>
<td>4.</td>
<td>Optimize SpO₂ with oxygen and mechanical ventilation</td>
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<td>5.</td>
<td>Transfuse with packed cells if haemoglobin &lt; 100 g/l</td>
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<tr>
<td>6.</td>
<td>Give clotting products as volume if there is a coagulopathy, but beware of hypotension caused by fresh frozen plasma</td>
</tr>
<tr>
<td>7.</td>
<td>Repeat lactate; if still high or blood pressure low, add dopamine at 5 µg/min</td>
</tr>
<tr>
<td>8.</td>
<td>Apply mechanical ventilation to reduce respiratory muscle oxygen consumption and allow redistribution of cardiac output to vital organs</td>
</tr>
<tr>
<td>9.</td>
<td>Transfuse with packed cells if haemoglobin &lt; 100 g/l</td>
</tr>
<tr>
<td>10.</td>
<td>Give paracetamol for fever and use other cooling methods as necessary (to normothermia)</td>
</tr>
<tr>
<td>11.</td>
<td>Increase inotropic support if lactate still rising</td>
</tr>
<tr>
<td>12.</td>
<td>Consider that catecholamines, especially adrenaline (epinephrine), might also cause high lactate concentrations</td>
</tr>
<tr>
<td>13.</td>
<td>Apply mechanical ventilation to reduce respiratory muscle oxygen consumption and allow redistribution of cardiac output to vital organs</td>
</tr>
<tr>
<td>14.</td>
<td>Do not stop dopamine on first postoperative night</td>
</tr>
<tr>
<td>15.</td>
<td>Maintain haemoglobin at 100–110 g/l</td>
</tr>
<tr>
<td>16.</td>
<td>Maintain sodium bicarbonate and avoid large volumes of fluid</td>
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<tr>
<td>17.</td>
<td>Give antihypertensive agents and diuretics as necessary to control blood pressure</td>
</tr>
<tr>
<td>18.</td>
<td>Increase oxygen supply if there is any respiratory abnormality</td>
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<tr>
<td>19.</td>
<td>Give inotropic agents as necessary to support cardiac output</td>
</tr>
<tr>
<td>20.</td>
<td>Close the aorta if necessary</td>
</tr>
<tr>
<td>21.</td>
<td>Continue monitoring until the child is fully supported</td>
</tr>
</tbody>
</table>

*Assumes appropriate antibiotics have been given.

**Table 2 Protocol to manage high blood lactate after cardiopulmonary bypass**

Correct deficits in oxygen delivery

- Maintain adequate circulating volume
- Maintain haemoglobin at 120–140 g/l
- Do not stop dopamine on first postoperative night
- Optimize arterial oxygen saturation
- Give paracetamol for fever and use other cooling methods as necessary (to normothermia)
- Do not stop neuromuscular blocking drugs or wean off ventilation on first postoperative night if there is a specific reason (for example, Fontan, Glenn), and then only with frequent reassessment of perfusion
- Start peritoneal dialysis early if the child has generalised oedema, oliguria, or large volumes of fluid are required to maintain circulation
- Repeat lactate at four and eight hours after intensive care unit (ICU) admission

If lactate is rising, or > 4 mmol/l

- Notify the ICU consultant and cardiac surgeon
- Check cardiac rhythm and perform echocardiography to examine cardiac function and exclude pericardial effusion
- Maintain intravascular volume

If the child is receiving adrenaline, and has an acceptable blood pressure but a rising lactate, reducing the adrenaline infusion rate might lower lactate

- Titrate the inotrope to the lowest infusion rate that will maintain a blood pressure in the normal range for age, a urine output > 1 ml/kg/hour, a reduction in lactate

If lactate continues to rise despite these measures, consider the use of extracorporeal life support

Continuous monitoring of fluid volume, or infusion of a hyperosmolar solution.

Therefore, the theoretical disadvantages of administered bicarbonate are: (1) complications of hyperosmolar solutions, particularly an increased risk of intraventricular haemorrhages in neonates; (2) ventilatory failure and arterial hypercarbia in patients with compromised pulmonary function; (3) worsening of intracellular acidosis; and (4) delayed tissue reoxygenation.

The current role of bicarbonate that is supported by an arguable balance of evidence; it should prompt consideration of associated conditions that might impair lactate production, such as depletion of glucose stores in malnutrition. It remains untested whether there is any metabolic abnormality common to children with fatal septic shock who never have hyperlactatemia.

**Future directions in the monitoring of dysoxia**

The earliest detection of dysoxia might be found by specific measures of the redox state of cells. The β-hydroxybutyrate to acetocetate (ketone body) ratio estimates liver mitochondrial NADH/NAD, and has been shown to be a marker of oxygen limited ATP flux in liver, and a predictor of adverse events after liver transplantation. What role it may have in other pathologies is unclear. Near infrared spectroscopy might be useful to monitor cytochrome redox states. The lactate to pyruvate ratio, reflecting the redox potential of the cytosol, might be a more specific marker of dysoxia than lactate alone. Before any new technique is adopted for practice, careful clinical studies in critically ill children, which compare the
predictive value for major adverse events, and the cost effectiveness, with those of established techniques, will be needed.

Conclusions

In dysoxia, oxygen limited ATP production produces lactic acid, but this is not the only mechanism for excess lactate in critically ill patients. There is not a direct relation between lactate concentrations and supply dependent oxygen consumption in all patients, although current techniques to test this are imprecise. One of the causes of mild hyperlactataemia in sepsis is impaired lactate removal. The role of lactate in the management of critically ill children is in alerting clinicians to a higher risk of mortality or adverse events, and it is but one useful tool to monitor response to treatment. The immediate management depends on consideration of the aetiology of hyperlactataemia. High blood lactate does not suggest a universal therapeutic response, but indicates severe physiological derangement associated with an increased risk of mortality. Lactate concentrations are not well predicted by other acid-base variables, and lactate is a better predictor of adverse outcomes, including mortality. Hyperlactataemia might be a specific predictor of morbidity risk, but in a population of critically ill children, has low sensitivity.

In children with hyperlactataemia it is sensible to consider whether there are correctable deficits in oxygen delivery. Current treatment aimed at lowering the lactate concentration by specifically altering a biochemical pathway does not improve survival, and neither does driving DO2 to supranormal levels with inotropes. Bicarbonate infusion has a limited role, and is only effective in normalising blood pH when generated CO2 is eliminated from the body. In many cases there will be identifiable and avoidable causes of insufficient tissue oxygen delivery, such as hypovolaemia or inadequate cardiac output.

Further research is required to define more sensitive, and equally inexpensive and non-invasive, markers of tissue dysoxia.