Campylobacter 0:41 isolation in Guillain-Barré syndrome

E A Goddard, A J Lastovica, A C Argent

Abstract
Over a period of 15 months, 17 children were admitted to the Red Cross War Memorial Children’s Hospital (RCWMCH) in Cape Town with Guillain-Barré syndrome. Stool specimens were collected from 14 children and campylobacter was isolated from nine. Six of the nine isolates of Campylobacter jejuni were further identified as C jejuni biotype 2, serotype 0:41. This biotype 2, serotype 0:41 has been identified in only 12 of the 7119 campylobacter isolates at the RCWMCH over a 19 year period. Eight of the nine patients with campylobacter isolates and one of five with negative stool cultures required ventilation. Patients with C jejuni biotype 2, serotype 0:41 were ventilated for a mean (SD) of 33.5 (19.4) days, whereas patients with other campylobacter isolates were ventilated for 17.3 (3.8) days. This is the first report of campylobacters of serotype 0:41 in Guillain-Barré syndrome and could reflect a geographical isolation of this strain.

(Keywords: Guillain-Barré syndrome; Campylobacter jejuni)

The Guillain-Barré syndrome (GBS) is the most common cause of acute neuromuscular paralysis. Although GBS has been associated with a wide variety of preceding infections, campylobacter infections appear to be the most common preceding event, with evidence of infection in up to 38% of adults with GBS. 

There have been a few reports of an association of childhood GBS and campylobacter infection.  

At the Red Cross War Memorial Children’s Hospital (RCWMCH) in Cape Town, an average of 10 cases of GBS have been seen each year since 1975. During the period June 1994 to August 1995 a total of 23 children with GBS were seen at the hospital, of whom 17 were admitted to the intensive care unit. This paper reports the associations between campylobacter infection and GBS in these patients.

Patients and methods
Stool specimens were collected from patients admitted to the intensive care unit at RCWMCH between June 1994 and August 1995 with clinical features of GBS. Patients were included in the study if they fulfilled the diagnostic criteria for acute GBS as published by Asbury and Cornblath. Clinical data were obtained from a retrospective analysis of the patients’ charts.

Isolation and growth
All stool specimens were cultured by established methods. Gram negative motile, spiral, or curved rods were cultured on tryptose blood agar plates (CM233, Oxoid Ltd) containing 10% v/v unlysed horse blood. Culture plates were incubated at 37°C under microaerophilic (12% carbon dioxide, 88% air, and 95% humidity) growth conditions.

Biotyping and serotyping
Identification was accomplished by established tests and all isolates were biotyped by the scheme of Skirrow and Benjamin. Serotyping on the basis of thermostable somatic (O) lipopolysaccharide antigens was performed with the 66 antisera of the Penner scheme. Isolates were also serotyped against an additional 30 antisera to new serotypes not included in this scheme.

Results
Over a period of 15 months, 17 children aged 14–108 (median 47) months were admitted to the intensive care unit of the RCWMCH in Cape Town with GBS (table 1). Campylobacter species were isolated from nine of the 14 patients from whom stool specimens were obtained. Six of these isolates were identified as C jejuni biotype 2, serotype 0:41. The other three campylobacter isolates were C concisus, C upsaliensis, and C jejuni biotype 1.

Before presentation with GBS only three of the nine patients with a positive campylobacter culture had a history of a preceding diarrhoeal illness, four had had a preceding upper respiratory tract infection, and two had no prior infection.

The GBS in patients with campylobacter isolates was more severe than in patients with negative stool cultures. Eight of the nine patients with campylobacter isolates and one of five patients with negative stool cultures required ventilation (p< 0.05) with a mean duration of ventilation of 28.1 days. The duration of admission to hospital was also prolonged in patients with campylobacter (mean 43 ± 9.8 days, p< 0.01). Only one patient who was ventilated for more than 20 days had a negative stool culture for campylobacter isolates. One patient from whom no stool was collected was ventilated for more than 20 days.

Among patients with campylobacter isolates, those with C jejuni biotype 2, serotype 0:41 had more severe disease. Overall, patients with C jejuni biotype 2, serotype 0:41 were ventilated
40% of whom are under the age of 14 years. Serving a population of three million people, the RCWMCH is one of two referral hospitals between the groups of patients. There was no difference in age (mean 28.1 days) ventilation and admission to hospital for a mean of 33.5 and 50.8 days, respectively. It is possible that the two patients (one with no stool isolate of campylobacter and one with no stool culture) with severe disease and no evidence of campylobacter infection had had recent infection as the organisms are not always found in stools at the time of presentation. Of the seven patients with CAGBS (four of these had serotype 0:19) reported by Kuroki et al,^1^ none required ventilation. One patient with CAGBS required ventilation for six weeks, but this patient had the complicating issue of severe short bowel syndrome.

_C jejuni_ biotype 2, serotype 0:41 is an extremely rare biotype of campylobacter in this region. Only 35 _C jejuni_ biotype 2 isolates have been identified from the 7119 campylobacter isolates characterised at the RCWMCH in the last 19 years. Of these 35 strains, 12 (including the _GBS_ related isolates) were serotyped as 0:41. The serotype 0:41 isolates not associated with _GBS_ were from six children with severe chronic diarrhoea. Previous reports of CAGBS have implicated bacterial strains of Penner serotype 0:19 and Lior biotype 11. The incidence of serotype 0:19 in campylobacter stool isolates seen at RCWMCH is <3% and _GBS_ was not recognised in patients with this serotype.

Previous studies have suggested that there is an association between the development of _GBS_ and the presence of antibodies to ganglioside GMM1. Several studies have suggested that there are similarities between the lipopolysaccharide structures of campylobacter and human gangliosides. Of particular interest is evidence that lipopolysaccharides extracted from _C jejuni_ of serotype 0:19 and 0:4 for 33.5 (95% confidence interval (CI) 13.1 to 54.0) days, whereas patients with other campylobacter isolates were ventilated for 17.3 (95% CI 7.9 to 26.7) days. However, our patients with _C jejuni_ biotype 2, serotype 0:41 were younger than those previously described, with the exception of the patients reported by De Bont et al (2 years) and Sugita et al (4 years). Patients reported by Kuroki et al were aged 7–14 years (median 11 years), whereas Yuki et al reported two patients aged 13 and 9 years with Miller-Fisher syndrome associated with campylobacter. The proportion of preceding gastrointestinal symptoms in our patients with CAGBS is similar to other studies of CAGBS where diarrhoea was noted in four of nine and 19 of patients.

Patients with CAGBS from whom _C jejuni_ biotype 2, serotype 0:41 was isolated had a particularly severe form of _GBS_ requiring ventilation and admission to hospital for a mean of 33.5 and 50.8 days, respectively. It is possible that the two patients (one with no stool isolate of campylobacter and one with no stool culture) with severe disease and no evidence of campylobacter infection had had recent infection as the organisms are not always found in stools at the time of presentation. Of the seven patients with CAGBS (four of these had serotype 0:19) reported by Kuroki et al, none required ventilation. One patient with CAGBS required ventilation for six weeks, but this patient had the complicating issue of severe short bowel syndrome.

_C jejuni_ biotype 2, serotype 0:41 is an extremely rare biotype of campylobacter in this region. Only 35 _C jejuni_ biotype 2 isolates have been identified from the 7119 campylobacter isolates characterised at the RCWMCH in the last 19 years. Of these 35 strains, 12 (including the _GBS_ related isolates) were serotyped as 0:41. The serotype 0:41 isolates not associated with _GBS_ were from six children with severe chronic diarrhoea. Previous reports of CAGBS have implicated bacterial strains of Penner serotype 0:19 and Lior biotype 11. The incidence of serotype 0:19 in campylobacter stool isolates seen at RCWMCH is <3% and _GBS_ was not recognised in patients with this serotype.

Previous studies have suggested that there is an association between the development of _GBS_ and the presence of antibodies to ganglioside GMM1. Several studies have suggested that there are similarities between the lipopolysaccharide structures of campylobacter and human gangliosides. Of particular interest is evidence that lipopolysaccharides extracted from _C jejuni_ of serotype 0:19 and 0:4 for 33.5 (95% confidence interval (CI) 13.1 to 54.0) days, whereas patients with other campylobacter isolates were ventilated for 17.3 (95% CI 7.9 to 26.7) days. However, our patients with _C jejuni_ biotype 2, serotype 0:41 were younger than those previously described, with the exception of the patients reported by De Bont et al (2 years) and Sugita et al (4 years). Patients reported by Kuroki et al were aged 7–14 years (median 11 years), whereas Yuki et al reported two patients aged 13 and 9 years with Miller-Fisher syndrome associated with campylobacter. The proportion of preceding gastrointestinal symptoms in our patients with CAGBS is similar to other studies of CAGBS where diarrhoea was noted in four of nine and 19 of patients.

Patients with CAGBS from whom _C jejuni_ biotype 2, serotype 0:41 was isolated had a particularly severe form of _GBS_ requiring ventilation and admission to hospital for a mean of 33.5 and 50.8 days, respectively. It is possible that the two patients (one with no stool isolate of campylobacter and one with no stool culture) with severe disease and no evidence of campylobacter infection had had recent infection as the organisms are not always found in stools at the time of presentation. Of the seven patients with CAGBS (four of these had serotype 0:19) reported by Kuroki et al, none required ventilation. One patient with CAGBS required ventilation for six weeks, but this patient had the complicating issue of severe short bowel syndrome.

_C jejuni_ biotype 2, serotype 0:41 is an extremely rare biotype of campylobacter in this region. Only 35 _C jejuni_ biotype 2 isolates have been identified from the 7119 campylobacter isolates characterised at the RCWMCH in the last 19 years. Of these 35 strains, 12 (including the _GBS_ related isolates) were serotyped as 0:41. The serotype 0:41 isolates not associated with _GBS_ were from six children with severe chronic diarrhoea. Previous reports of CAGBS have implicated bacterial strains of Penner serotype 0:19 and Lior biotype 11. The incidence of serotype 0:19 in campylobacter stool isolates seen at RCWMCH is <3% and _GBS_ was not recognised in patients with this serotype.

Previous studies have suggested that there is an association between the development of _GBS_ and the presence of antibodies to ganglioside GMM1. Several studies have suggested that there are similarities between the lipopolysaccharide structures of campylobacter and human gangliosides. Of particular interest is evidence that lipopolysaccharides extracted from _C jejuni_ of serotype 0:19 and 0:4

### Table 1: Patient details

<table>
<thead>
<tr>
<th>Patient No</th>
<th>Age (months)</th>
<th>Stool culture</th>
<th>Duration of ventilation (days)</th>
<th>Duration of stay in hospital (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>16</td>
<td>None cultured</td>
<td>24</td>
<td>34</td>
</tr>
<tr>
<td>2</td>
<td>73</td>
<td>None cultured</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>3</td>
<td>45</td>
<td>None cultured</td>
<td>0</td>
<td>6</td>
</tr>
<tr>
<td>4</td>
<td>108</td>
<td>None cultured</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>5</td>
<td>48</td>
<td>None cultured</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>Mean (SD)</td>
<td>58.0 (34.5)</td>
<td>None cultured</td>
<td>4.8 (10.7)</td>
<td>11.8 (13.0)</td>
</tr>
<tr>
<td>6</td>
<td>22</td>
<td>Biotype 2, serotype 0:41</td>
<td>47</td>
<td>51</td>
</tr>
<tr>
<td>7</td>
<td>108</td>
<td>Biotype 2, serotype 0:41</td>
<td>37</td>
<td>42</td>
</tr>
<tr>
<td>8</td>
<td>84</td>
<td>Biotype 2, serotype 0:41</td>
<td>54</td>
<td>74</td>
</tr>
<tr>
<td>9</td>
<td>36</td>
<td>Biotype 2, serotype 0:41</td>
<td>0</td>
<td>28</td>
</tr>
<tr>
<td>10</td>
<td>26</td>
<td>Biotype 2, serotype 0:41</td>
<td>29</td>
<td>51</td>
</tr>
<tr>
<td>11</td>
<td>22</td>
<td>Biotype 2, serotype 0:41</td>
<td>33</td>
<td>59</td>
</tr>
<tr>
<td>Mean (SD)</td>
<td>49.7 (37.0)</td>
<td>C concisus</td>
<td>33.5 (19.4)</td>
<td>50.8 (15.5)</td>
</tr>
<tr>
<td>12</td>
<td>48</td>
<td>C jejuni biotype 2</td>
<td>19</td>
<td>26</td>
</tr>
<tr>
<td>13</td>
<td>47</td>
<td>C jejuni biotype 1</td>
<td>13</td>
<td>23</td>
</tr>
<tr>
<td>14</td>
<td>14</td>
<td>Biotype 1</td>
<td>20</td>
<td>33</td>
</tr>
<tr>
<td>Mean (SD)</td>
<td>20.3 (37.0)</td>
<td>Not done</td>
<td>17.3 (3.8)</td>
<td>27.3 (5.13)</td>
</tr>
<tr>
<td>15</td>
<td>47</td>
<td>Not done</td>
<td>0</td>
<td>13</td>
</tr>
<tr>
<td>16</td>
<td>64</td>
<td>Not done</td>
<td>71</td>
<td>71</td>
</tr>
<tr>
<td>17</td>
<td>96</td>
<td>Not done</td>
<td>8</td>
<td>15</td>
</tr>
<tr>
<td>Mean (SD)</td>
<td>69 (24.9)</td>
<td>Not done</td>
<td>26.3 (38.9)</td>
<td>33.0 (33.0)</td>
</tr>
</tbody>
</table>
had core oligosaccharides with terminal structures resembling human gangliosides GM₁ and GD₁b. To date, there is no published data about the lipopolysaccharide structures of C. jejuni biotype 2, serotype 0:41 and this should be studied further.

We did not perform nerve conduction studies in these patients and thus do not know whether our patients had patterns of acute inflammatory demyelinating polyneuropathy or acute motor axonal neuropathy. It is impractical to treat patients with C. jejuni positive stools with antibiotics to prevent the rare complication of GBS. Previous studies have suggested that treatment of patients with campylobacter associated GBS does not prevent the development of GBS. 2, 3 26 It may be worth investigating the possibility that early treatment with erythromycin may shorten the duration of the illness, however. Although treatment with erythromycin does not affect the natural course of campylobacter enteritis, it does clear campylobacter from stools. In two studies of acute campylobacter enteritis erythromycin cleared campylobacter from stools of all 28 patients within two days, and in a study of 10 convalescent excretors erythromycin treatment cleared the stool within 48 hours. 29 Thus erythromycin does clear the organism and if there is a cross reaction with molecular mimicry between campylobacter and the central nervous system, clearance of the organism and reduction of the antigen load may make a difference to the clinical course of the patients.

Conclusions

We have documented the association of a severe form of GBS in children with concurrent infection with campylobacter species and particularly with C. jejuni biotype 2, serotype 0:41 (an extremely rare serotype in this community). This highlights the importance of campylobacter infection as a possible cause of GBS and raises further possible directions for the treatment and investigation of pathogenetic mechanisms of GBS.

We thank Mrs I Heuer for typing this paper. We also thank E le Roux in the Department of Microbiology at the Red Cross War Memorial Children’s Hospital for culturing of the stool specimens. This research was supported by grants from the South African Medical Research Council and the University of Cape Town to AJL.