Influence of infant feeding and gluten intake on coeliac disease

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Abstract

Objectives—To study the impact of infant feeding habits and actual gluten intake on gluten induced enteropathy.

Methods—A case-referent design, controlling for the HLA alleles conferring increased genetic risk, was used. All 164 siblings of 97 probands were investigated. Eighty five of the siblings, carrying the genes DQA1*0501-DQB1*02 conferring susceptibility for the disease, were investigated by interview, food recording, and taking a small intestinal biopsy sample. Eight cases of silent coeliac disease were found and these were compared with the 73 siblings in whom the diagnosis was excluded.

Results—No statistically significant differences were found between cases and referents in terms of duration of breast feeding, age at introduction of cows’ milk products, frequency of breast feeding after gluten introduction, and gluten consumption.

Conclusions—The studied factors may be of less importance for the development of gluten induced enteropathy.

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Keywords: coeliac disease; HLA alleles; infant feeding habits; gluten consumption.

There are at least two prerequisites for developing coeliac disease: a genetic predisposition and gluten intake. So far, two HLA alleles on chromosome 6 have been shown to confer a primary susceptibility.1 These alleles encode the heterodimer DQα1*0501,β1*02), found in 80–100% of patients with coeliac disease2 and corresponding to the serological HLA type DR3-DQ2 or DR5/7-DQ2, considered as the HLA haplotypes at risk for coeliac disease.3

Most people who carry these genes do not develop coeliac disease, however, despite gluten intake. It therefore seems probable that additional genes, as yet unknown, are involved or that other factors are of importance. The duration of breast feeding, the age at which gluten is introduced into the infant diet, the duration of the period when both breast milk and gluten are given, the amount of gluten consumed, and infectious agents are environmental factors suggested to be of importance.4

Sweden, in contrast with its neighbouring countries,5,6 has had an increasing incidence of coeliac disease during the last decades, particularly in children born after 1982.6,7 In the search for an explanation, interest has mainly been focused on infant feeding patterns for two reasons: the infant feeding habits in Sweden are different to those in neighbouring countries, and the increase in incidence has occurred during a period when infant feeding patterns have changed.8,9 Whether these changes have caused genetically less susceptible subjects to develop coeliac disease or if they have led to more obvious symptoms in children with existing gluten induced enteropathy is discussed.10–12

The aim of this study was to analyse the impact of early infant feeding habits and actual gluten intake on gluten induced enteropathy by controlling for the HLA alleles considered as an expression of the genetic risk.

Patients and methods

A case-referent design was applied in the study using the siblings of patients verified as having coeliac disease. Only siblings with DR3-DQ2 or DR5/DR7-DQ2 were included to match for genetic risk factors. The cases were defined as siblings with previously unknown coeliac disease and the referents as siblings without coeliac disease.

Patients

An overview of the study design is shown in fig 1. The selection was made in the following way. A letter of invitation was sent to the 268 families of 272 patients with coeliac disease living in the area, the coeliac disease having been diagnosed between 1970 and 1991 at the department of paediatrics, East University Hospital, Göteborg. The family was included only if (a) the proband was diagnosed according to the original ESPGAN criteria,13 (b) he or she had at least one full sibling older than 6 months without diagnosed coeliac disease, and (c) both parents were available for HLA testing. Of the 268 families, 150 families with 160 patients and 234 siblings fulfilled the inclusion criteria, whereas 84 families did not. Whether the remaining 25 families fulfilled the criteria is unclear as they did not answer the letter of invitation.

Ninety six families with 97 probands and 164 siblings, including three families with three probands and four siblings from the vicinity of Göteborg who approached us spontaneously and requested participation, were enrolled in the study (fig 1). All family members were tested for HLA class I and II antigens and serological coeliac disease markers (see later).

At the same time the families were instructed to ensure that the siblings were eating normal

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gluten containing foods. Eighty five siblings in 66 families carried the HLA genotypes DR3-DQ2/DR3-DQ2, DR3-DQ2/x (where x are configurations other than DR3-DQ2) or DR5/DR7-DQ2, whereas 79 did not and were excluded from further study. The mothers were interviewed and a small intestinal biopsy sample was taken from 82 siblings, preceded by a four day recording of food intake. Of the remaining three siblings, one had had a small intestinal biopsy sample taken previously showing normal mucosa, and two siblings dropped out from the study. Of the 79 siblings who did not carry any of the HLA risk alleles, 35 of 36 who shared other HLA haplotypes with their proband sibling had a small intestinal biopsy sample taken as they were expected to have an increased genetic risk. The intestinal mucosa was normal in all instances. All but one of the 79 siblings were negative in serological markers for coeliac disease and were considered to be healthy. The only boy with slightly increased antibodies to gliadin later had a small intestinal biopsy sample taken which showed a normal mucosa.

Eight cases were found with an intestinal mucosa compatible with coeliac disease (table 1). The referent group (73 subjects) consisted of 61 siblings with a normal intestinal histology, the one sibling who previously had a normal small intestinal biopsy sample taken, and the two siblings with normal serological markers for coeliac disease who dropped out of the study. Of 13 siblings with various degrees of mucosal inflammation but normal villus height and crypt depth, nine became normal histologically in spite of increased gluten intake. They were accordingly included in the referent group. The final diagnosis of the remaining four of the 13 siblings with slight histological changes remained unclear and they were excluded from further analysis.

### Table 1 Characteristics of the cases

<table>
<thead>
<tr>
<th>Patient No</th>
<th>Birth year</th>
<th>Age when small intestinal biopsy sample taken (years)</th>
<th>Sex</th>
<th>HLA class II</th>
<th>Duration of breast feeding (months)</th>
<th>Age at cows' milk introduction (months)</th>
<th>Age at gluten introduction (months)</th>
<th>Duration of mixed breast-milk-gluten feeding (months)</th>
<th>Wheat protein consumption at time of small intestinal biopsy sample (g/kg/day)</th>
<th>EMA</th>
<th>AGA IgG</th>
<th>AGA IgA</th>
<th>Symptoms given in parental interview</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1990</td>
<td>3.2</td>
<td>F</td>
<td>DR3-DQ2/DR4-DQ3</td>
<td>5</td>
<td>3</td>
<td>6</td>
<td>0</td>
<td>0.24</td>
<td>Positive</td>
<td>Positive</td>
<td>Negative</td>
<td>None</td>
</tr>
<tr>
<td>2</td>
<td>1988</td>
<td>5.7</td>
<td>F</td>
<td>DR2-DQ1/DR3-DQ2</td>
<td>9</td>
<td>6</td>
<td>6</td>
<td>3</td>
<td>0.14</td>
<td>Positive</td>
<td>Positive</td>
<td>Negative</td>
<td>Constipation, eczema</td>
</tr>
<tr>
<td>3</td>
<td>1987</td>
<td>6.4</td>
<td>F</td>
<td>DR3-DQ2/DR4-DQ3</td>
<td>7.5</td>
<td>4</td>
<td>6</td>
<td>1.5</td>
<td>0.36</td>
<td>Positive</td>
<td>Negative</td>
<td>Negative</td>
<td>Previous abdominal protrusion and suspected cows' milk intolerance, Constipation, DAMP, delayed development</td>
</tr>
<tr>
<td>4</td>
<td>1985</td>
<td>7.4</td>
<td>M</td>
<td>DR1-DQ1/DR3-DQ2</td>
<td>1.5</td>
<td>1.5</td>
<td>5</td>
<td>0</td>
<td>0.07</td>
<td>Positive</td>
<td>ND</td>
<td>ND</td>
<td>None</td>
</tr>
<tr>
<td>5</td>
<td>1986</td>
<td>7.8</td>
<td>M</td>
<td>DR2-DQ1/DR3-DQ2</td>
<td>8</td>
<td>3.5</td>
<td>6</td>
<td>2</td>
<td>0.27</td>
<td>Positive</td>
<td>ND</td>
<td>ND</td>
<td>Previous diarrhoea, headache and abdominal protrusion, Sporadic constipation and abdominal pain</td>
</tr>
<tr>
<td>6</td>
<td>1984</td>
<td>9.1</td>
<td>F</td>
<td>DR3-DQ2/DR6-DQ1</td>
<td>3.5</td>
<td>3</td>
<td>5</td>
<td>0</td>
<td>0.11</td>
<td>Positive</td>
<td>ND</td>
<td>ND</td>
<td>Diffuse abdominal complaints, Previous diarrhoea, vomiting, abdominal pain</td>
</tr>
<tr>
<td>7</td>
<td>1980</td>
<td>13.4</td>
<td>F</td>
<td>DR3-DQ2/DR7-DQ2</td>
<td>7</td>
<td>5</td>
<td>6</td>
<td>1</td>
<td>0.25</td>
<td>Positive</td>
<td>ND</td>
<td>ND</td>
<td>None</td>
</tr>
<tr>
<td>8</td>
<td>1976</td>
<td>17.7</td>
<td>F</td>
<td>DR1-DQ1/DR3-DQ2</td>
<td>6</td>
<td>4</td>
<td>5</td>
<td>1</td>
<td>0.14</td>
<td>Positive</td>
<td>ND</td>
<td>ND</td>
<td>None</td>
</tr>
</tbody>
</table>

ND, not done; DAMP, dysfunction in attention, motor control, and perception; AGA, antigliadin antibodies; EMA, antieldomysium antibodies.
infantfeedingandglutenintakeincoeliacdisease

115

dietician by the same method as in a previous study.9 barley) was calculated by one paediatric ten containing cereals (wheat, rye, oats, and gluten was not known either by the mother or the interviewer at the time of the interview. The questions aimed to assess the duration of breast feeding, the age at which cows’ milk and gluten containing foods were introduced, and, when relevant, the occurrence of symptoms for each child in the family. The answers were categorised and recorded by the interviewer.

INTERVIEW

After pilot testing one specially trained children’s nurse performed a semistructured interview with the mother of each family in their own home environment at a time selected by the mother. The result of the HLA typing was read by the same experienced pathologist.

SMALL INTESTINAL BIOPSY SAMPLE

A small intestinal biopsy sample was taken (preferably at the level of the Treitz ligament) with a Watson capsule (paediatric size) or a Storz capsule under fluoroscopic control by one of us (HA). The specimens were handled and classified as previously described16 and read by the same experienced pathologist.

SEROLOGICAL MARKERS

All parents and siblings were tested for total IgA and antibodies to endomysium of IgA class in the serum.17 Children less than 5 years of age were also tested for the presence of IgA and IgG class antibodies to gliadin.18

STATISTICAL METHODS

For the statistical evaluation of the differences between cases and referents, Fisher’s exact test in fourfold tables and, where appropriate, the \( \chi^2 \) test were used. The infant feeding pattern and gluten consumption were not normally distributed, median values and Wilcoxon’s two sample test were used instead.

Table 2 Early feeding pattern and actual gluten intake in cases and referents

<table>
<thead>
<tr>
<th></th>
<th>Cases Median</th>
<th>Range No</th>
<th>Referents Median</th>
<th>Range No</th>
</tr>
</thead>
<tbody>
<tr>
<td>Duration of breast feeding (months)</td>
<td>6.5</td>
<td>1.5–9.0</td>
<td>8</td>
<td>5.0</td>
</tr>
<tr>
<td>Age at introduction of foods containing cows’ milk (months)</td>
<td>4.0</td>
<td>1.5–6.0</td>
<td>7</td>
<td>3.0</td>
</tr>
<tr>
<td>Age at introduction of foods containing gluten (months)</td>
<td>6.0</td>
<td>3.5–6.0</td>
<td>8</td>
<td>6.0</td>
</tr>
<tr>
<td>Consumption of wheat protein at the time of biopsy sample (g/kg/day)</td>
<td>0.19</td>
<td>0.07–0.36</td>
<td>8</td>
<td>0.27</td>
</tr>
</tbody>
</table>

HLA TYPING

T and B lymphocytes were prepared from 10 ml of a mixture of EDTA and blood by immunomagnetic separation.14 HLA class I and II types were determined by serology using a microlymphocytotoxic technique. Each specificity was assigned by several well defined antisera. The same typing trays were used during the whole investigation.

FOOD RECORDING

After receiving instructions, food recordings were made during four days by the parents or the adolescent patients under study. All foods and beverages given, measured with household measures, were included.15 The amount of gluten containing cereals (wheat, rye, oats, and barley) was calculated by one paediatric dietician by the same method as in a previous study.9

ETHICS

The study was approved by the ethical committee of the Faculty of Medicine at Göteborg University. Several reasons justified the use of taking a small intestinal biopsy sample in the protocol. No serological marker in coeliac disease has a 100% sensitivity and antibodies to endomysium have a lower sensitivity in the youngest age groups.17 18 In addition, these tests have mainly been evaluated in patients with symptoms and the results could not simply be extrapolated to this study investigating subjects without symptoms. In spite of improved serological markers for coeliac disease, a small intestinal biopsy sample therefore remains the golden standard for the diagnosis.

As the long term health risks with untreated coeliac disease are mainly related to the state of the mucosa rather than the existence or severity of symptoms,21 the study offered benefits for the siblings under study in identifying undiagnosed coeliac disease.

RESULTS

The median age at which the small intestinal biopsy sample was taken was 7.9 years for the cases (range 3.2–17.9) and 7.4 years for the referents (0.92–25.0; NS). Seventy five per cent of the cases and 59% of the referents were girls. Fifty per cent of the cases were the first born child of the family, compared with 38% of the referents (NS) and 63% of the cases were born before their proband compared with 51% of the referents (NS).

Compared with the referents, the cases had a longer duration of breast feeding (NS), a later introduction of foods containing cows’ milk (table 2) (NS), as well as a higher frequency of breast feeding during the introduction of gluten containing follow up formulas (table 3) (NS) and a lower consumption of gluten at the time the small intestinal biopsy sample was taken (table 2) (p = 0.07; NS). Two of 26 siblings (8%) born before 1982, when the recommended age for gluten introduction changed from 4 to 6 months and the gluten content in the baby foods increased, had coeliac disease, compared with six of 51 (12%) of those born after that year (NS).

The risk for coeliac disease was 5.5% (9/165, the previously diagnosed case included) in all siblings and 10.6% (9/85) in siblings carrying the HLA risk allele.

DISCUSSION

Since the method of taking a small intestinal biopsy sample was introduced, a number of sibling studies of coeliac disease have been performed,22–33 but only a few have also investi-
Table 3 The relation of breast feeding and introduction of gluten containing follow up formulas

<table>
<thead>
<tr>
<th>Breast feeding continued during introduction</th>
<th>No (%) cases (n=8)</th>
<th>No (%) referents (n=73)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast feeding discontinued at time of introduction</td>
<td>5 (63)</td>
<td>25 (34)</td>
</tr>
<tr>
<td>Breast feeding discontinued before introduction</td>
<td>0</td>
<td>13 (18)</td>
</tr>
</tbody>
</table>

Breastfeeding discontinued before introduction 3 (37) 35 (48)

The present study is the first which has tried to study the influence of this factor and the actual gluten consumption on coeliac disease matching for a genetic risk.

The reason for using a family design was that we could expect to find both a sufficient number of cases and genetically matched referents within a study group of a reasonable size. A population study, which theoretically would be advantageous, would need a sample size practically and ethically difficult to justify.

Previous studies indicated an expected overall risk for coeliac disease in siblings to coeliac disease patients of 10%, increasing to 28–40% in HLA identical or DR3 positive siblings. We could only find eight new cases, however, which together with the already diagnosed sibling corresponded to an overall risk of 5.5% or 10.5% in HLA identical or DR3-DQ2 or DR5/DR7-DQ2 positive siblings. One possible explanation for these variances might be different selections.

A selection bias may have worked on two levels. Firstly, if families with symptomatic children are more prone to participate in studies, this would result in a falsely higher prevalence in studies with a low participation rate. Except for the studies by Rolles et al and Stenhammaren al, who studied the parents and families, respectively, of consecutive cases, it is difficult to identify the number invited and the selection procedures used in previous family studies. In our study, all patients with coeliac disease available at our department were invited and 61% of the patients fulfilling the inclusion criteria agreed to participate.

Secondly, in many studies the figures for the sibling frequency are calculated by dividing the number of new cases by the number of biopsy samples taken rather than by all the siblings included. This makes the result vulnerable to the selection procedure for taking the small intestinal biopsy sample. A higher rate of acceptance among relatives with decreased wellbeing has to be expected and a falsely high prevalence in studies with a low sample rate for the small intestinal biopsy sample could be the effect. An analysis of the nine studies, including this study, in which the necessary figures were available did show a strong negative correlation between the occurrence (number of new cases of coeliac disease per number of biopsy samples) and the frequency of small intestinal biopsy samples (number of biopsy samples per total number of siblings). When the frequency was calculated as a prevalence—that is, the number of new cases divided by the total number of siblings—no such association was found and the prevalence was 6.5% (50/769, already known cases included) with a corresponding 95% confidence interval of 4.8 to 8.2—that is, about half of that which has generally been believed. This figure is in accordance with the results of our study.

A factor that may be of importance for the development of gluten induced enteropathy is the duration of gluten exposure. As the time at risk is relatively short in our study, the prevalence may increase in the future. On the other hand, if this factor was important, a higher age in the cases than in the referents would be expected.

Early termination of breast feeding has been found to be a risk factor for diagnosed—that is, symptomatic—patients with coeliac disease in previous studies. The early introduction of cows’ milk or gluten containing foods, the termination of breast feeding before the introduction of gluten, and a high consumption of gluten have also been hypothesised as risk factors. In this study we found no support for these theories. The duration of breast feeding was long compared with patterns in other Western countries, both in the cases and referents. No significant differences in the feeding patterns were found between the cases and referents. The tendency was rather towards a longer duration of breast feeding, a later introduction of formulas containing cows’ milk, more mixed breast and gluten feeding, and a lower gluten consumption in the cases. The latter might be an effect of a spontaneous reduction in the gluten consumption to avoid symptoms. It might be suspected that both cases and referents normally consume low amounts of gluten as they have siblings on a gluten free diet. According to our data, however, they consumed 0.28 and 0.34 g of flour protein/kg body weight/day (wheat, rye, oats, and barley), which can be compared with the consumption of approximately 0.27 g/kg body weight/day in Swedish adults with newly diagnosed dermatitis herpetiformis before dietary treatment.

Various explanations for the lack of differences between the cases and referents found in this study can be suggested. Firstly, the studied populations are different. Most previous studies on early infant feeding and coeliac disease have been made on diagnosed—that is, symptomatic—cases whereas we investigated patients with ‘silent’—that is, more or less asymptomatic—coeliac disease. The factors under study may have an effect on the symptoms of coeliac disease rather than on the enteropathy itself. Even if most of the cases in our study had some symptoms (table 1), they were so discrete that the parents had not suspected them to be caused by coeliac disease in spite of the awareness of the disease in these families. Secondly, the unfortunately low number of cases in our study might result in biologically significant differences remaining statistically insignificant. For any study of this type obtaining a sufficient number of cases is a problem. Meta-analysis may be a solution to this problem.

On the other hand, the lack of statistical significance may indicate that the factors studied are actually of less importance for the develop-
ment of gluten induced enteropathy. In addition, environmental conditions were more important than genetic predisposition in determining the occurrence of coeliac disease, a higher prevalence of the disease among relatives would be expected in a high incidence area, such as Sweden, compared with areas with a lower incidence. The low prevalence of coeliac disease in siblings found in this study as well as in the whole group of patients with coeliac disease in Göteborg therefore suggests that genetic factors, most probably factors besides DQA1*0501 and DQB1*02 and as yet unknown, are of greater importance for determining the prevalence of gluten induced enteropathy than environmental factors, which may play a more important part in the clinical expression of the disease.

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