

REGULAR REVIEW

Recent advances in understanding muscular dystrophy

K M D Bushby

The isolation of the causative genes in the muscular dystrophies, and from these the identification of the defective protein products, is at last giving an insight into the molecular basis of these conditions. These advances have come about through the techniques of reverse genetics, or positional cloning.¹ All the muscular dystrophies have consistently eluded attempts to use standard biochemical techniques to identify the underlying defects in affected muscle. Most advances have been made in Duchenne and Becker muscular dystrophy, for which the gene and protein product have been known for some time.²⁻³ The gene for myotonic dystrophy has been isolated very recently,⁴⁻⁶ and linkage analysis has identified chromosomal localisations for facioscapulohumeral dystrophy on 4q,⁷ a form of recessive limb-girdle on 15q,⁸ a form of dominant limb-girdle on 5q,⁹ and Emery-Dreifuss muscular dystrophy at Xq28.¹⁰

The gene involved in Duchenne and Becker muscular dystrophy is the largest yet identified, comprising over 2.5 megabases of genomic DNA with at least 75 exons and some very large introns.²⁻¹¹ This encodes a 14 kilobases mRNA, from which a previously unknown muscle protein, dystrophin, is translated. The predicted molecular weight of dystrophin is approximately 427 kilodaltons and it comprises about 0.001% of total muscle protein.³ Dystrophin is also present in brain, where it uses a different promoter and exists as a different isoform.¹² It appears to be localised to neurons.¹³ A protein with significant homology to dystrophin encoded by a gene on chromosome 6 has also been described.¹⁴

Study of the sequence of dystrophin suggests the presence of four distinct domains.¹⁵ The N-terminal region has homology to α -actinin and appears to have a site of interaction with F-actin.¹⁶ The second domain is thought to contain 24 or 25 triple helical repeats,¹⁷⁻¹⁸ which may confer elasticity on the molecule.¹⁷ There follows a sequence relatively rich in cysteine residues and a C-terminus domain that is highly conserved between species.¹⁹ Dystrophin has been shown by electron microscopy to be located at the internal cytoplasmic face of the plasma membrane²⁰ where it is associated with integral membrane glycoproteins²¹ via its C-terminus domain. The structure of this glycoprotein complex has recently been described and it has been shown that dystrophin deficient fibres also show a reduction in some components

of this complex.²²⁻²³ The structure and functional characteristics of the glycoprotein complex associated with dystrophin suggests that the function of dystrophin may be to link the sarcolemma membrane skeleton through a transmembrane complex to an extracellular glycoprotein which binds laminin.²² It has been shown that dystrophin deficient fibres are more sensitive to hypo-osmotic shock than controls,²⁴ which supports the theory that dystrophin deficiency leads to mechanical weakening of the plasma membrane. An influx of calcium has been observed in dystrophin negative fibres, which may, directly or indirectly, stimulate protease activity,²⁵ and myotubes negative for dystrophin have been shown to have abnormal calcium ion channels.²⁶

It is now known that most mutations in the dystrophin gene are deletions, which can be identified in approximately 65% of patients with Duchenne muscular dystrophy and up to 85% in Becker muscular dystrophy.²⁷⁻²⁸ The high frequency of deletions and in particular the tendency for deletions to cluster in two particular regions of the gene has not yet fully been explained.²⁹⁻³⁰ In those families where a patient has been shown to have a deletion DNA analysis offers a specific diagnostic test and a means of reliable and quick prenatal diagnosis. The demonstration of DNA abnormalities in patients without deletions is much more difficult in such a large gene, and only a handful of such mutations (point mutations and mutations in the promoter region) have been described.³¹⁻³² Techniques such as the use of dosage analysis for deletion detection in females, based on both Southern blotting and polymerase chain reaction (PCR) analysis, pulsed field gel electrophoresis, and most recently lymphocyte RNA PCR³³ have led to direct carrier detection being possible in some cases. However, in many cases carrier risk still has to be assessed as a probability calculation based on the results of creatine kinase testing and intragenic polymorphic markers. The high mutation rate in Duchenne muscular dystrophy has long been recognised, with approximately one third of cases estimated to result from a new mutation. The use of techniques that can directly detect a mutation in a boy and his mother has led to the recognition that germline mosaicism exists at a significant level in the Duchenne muscular dystrophy gene, with the risk of another affected son to a mother of an isolated case of Duchenne muscular dystrophy being around 20% with the at-risk

Department of
Human Genetics,
19/20 Claremont Place,
Newcastle upon Tyne
NE2 4AA

Correspondence to:
Dr Bushby.

haplotype even when a deletion has been identified in the boy and shown to be somatically absent in the mother.³⁴

Dystrophin analysis, using antibodies to different parts of the dystrophin molecule, can be used both on muscle sections and western blots in diagnosis. Abnormalities of dystrophin seem to be specific to the Xp21 dystrophies, and so the finding of abnormal dystrophin can confirm the diagnosis even in the absence of a cDNA deletion.^{35 36} The use of a combination of genetic and dystrophin analyses to distinguish Becker muscular dystrophy accurately from the often clinically similar conditions of spinal muscular atrophy and limb-girdle muscular dystrophy has allowed the prevalence of this condition to be reassessed, and it has been shown to be much commoner than had previously been thought,³⁷ the prevalence being similar to that of Duchenne muscular dystrophy and the cumulative birth incidence approximately one third. Some isolated cases of females with limb-girdle muscle weakness have been shown by dystrophin analysis to be manifesting carriers of dystrophin mutations³⁸ a finding of great significance in counselling these women. Dystrophin abnormalities do not seem to be so easily detectable in female carriers without symptoms.³⁹

Studies of the gene and protein abnormalities in large numbers of patients are beginning to unravel the complexity of the relationship between genotype and phenotype in the Xp21 dystrophies. A range of clinical severity, from the classical Duchenne muscular dystrophy patients at one end to the relatively mild Becker muscular dystrophy patients at the other, with patients of 'intermediate' severity between, is now known to exist as a result of different mutations in this gene.⁴⁰ The ability to look directly for the molecular defect has also led to the recognition that severe muscle cramps may, at least for many years, be the only manifestation of Xp21 dystrophy.⁴¹ Occasional asymptomatic cases have been recognised with dystrophin gene deletions.⁴² The clinical range of severity is now known to be a reflection of a similar range of gene and protein defects. Deletions that disrupt the reading frame of the gene⁴³ (with the expected result that no recognisable protein could be produced beyond the position of the mutation), are often associated with undetectable dystrophin, and these patients are generally very severely affected. However, dystrophin is detected in some patients with out of frame deletions, and the size of this dystrophin implies that the deletion behaves as if 'in frame', that is recognisable dystrophin is produced right through to the C-terminus of the protein,^{44 45} and the size of the protein is as predicted from the loss of the deleted exons.

Preliminary results suggest that the presence in muscle fibres of even a small amount of dystrophin may confer some functional advantage on these boys (LVB Nicholson, personal communication). At the other end of the clinical spectrum, patients with Becker muscular dystrophy are most commonly found to have deletions which do not disrupt the translational reading frame of the gene, so that a dystrophin

molecule is produced which is internally deleted, but with both ends present and intact. Even very large deletions, providing they are in frame can be associated with a mild phenotype,⁴⁶ but it appears that deletions 45-47 and 45-48 of the dystrophin gene do tend to be associated with a relatively consistently mild clinical course.²⁸ Across the range of Xp21 dystrophy, increasing abundance of dystrophin does seem to be associated with a milder clinical course, but there is no one value for abundance which reliably predicts a particular phenotype.

It is likely that alternative splicing mechanisms around deletions at the RNA level may be more widespread in the dystrophin gene than has yet been described,⁴⁷ and this may account for apparent anomalies in phenotype-genotype correlations. Interactions with the dystrophin-associated glycoproteins or other genes may yet account for some of the unexplained variability in the dystrophin gene.

With an understanding of the molecular basis of the Xp21 dystrophies, work on possible therapeutic measures can now be more rationally planned and assessed. Animal models with dystrophin abnormalities have been identified.^{48 49} Myoblast transfer and gene therapy using a 'mini-gene' construct have both shown some promising results in animal work,^{50 51} and dystrophin transcripts have been identified in patients after myoblast transfer,⁵² but widespread use of either technique in patients is still generally held to be a long way off.

The mutation responsible for myotonic dystrophy has recently been described.⁴⁻⁶ There is variable amplification of a trinucleotide (CTG) repeat at the 3' end of a gene, with an increase in the number of repeats associated with the disease phenotype. The number of repeats increases with the transmission of the mutation to successive generations. This mechanism explains the phenomenon of anticipation seen in families with myotonic dystrophy, where the severity of the disease is often seen to increase through successive generations. However, as amplification of the repeat may occur whether the mutation is passed through the male or female line, this mechanism is not enough alone to explain why congenital myotonic dystrophy is almost always maternally inherited. Women with myotonic dystrophy have around a 10% chance of having a child with congenital myotonic dystrophy, a risk which increases to 40% after the birth of one congenitally affected child, and which also rises with the severity of the disease in the mother.⁵³

The gene has homology with genes encoding cyclic-AMP-dependent protein kinase, and its protein product has been designated myotonin protein kinase.⁵⁴ A 3 kilobase transcript is expressed at high levels in heart and lower levels in skeletal muscle and brain.⁵⁵

Linkage analysis in the other muscular dystrophies should also ultimately lead to the identification of the genes and proteins involved, and no doubt to new ideas of how normal and abnormal genes and proteins interact in muscle. In the meantime, the work so far achieved has resulted in significant improvements in diagnosis and counselling for patients and their families.

Many thanks to David Gardner-Medwin for helpful comments. KMDB is an MRC Training Fellow.

- 1 Orkin SH. Reverse genetics and human diseases. *Cell* 1986; 47:845-50.
- 2 Koenig M, Hoffman EP, Bertelson CJ, Monaco AP, Feener C, Kunkel LM. Complete cloning of the Duchenne muscular dystrophy (DMD) cDNA and preliminary genomic organisation of the DMD gene in normal and affected individuals. *Cell* 1987;50:509-17.
- 3 Hoffman EP, Brown RH Jr, Kunkel LM. Dystrophin: the protein product of the Duchenne muscular dystrophy locus. *Cell* 1987;51:919-28.
- 4 Harley HG, Brook JD, Rundle SA, et al. Expansion of an unstable DNA region and phenotypic variation in myotonic dystrophy. *Nature* 1991;355:545-7.
- 5 Buxton J, Shelbourne P, Davies J, et al. Detection of an unstable fragment of DNA specific to individuals with myotonic dystrophy. *Nature* 1992;355:547-8.
- 6 Aslandis C, Jansen G, Amemiya C, et al. Cloning of the essential myotonic dystrophy region and mapping of the putative defect. *Nature* 1992;355:548-51.
- 7 Wijmenga C, Frantz R, Brouwer OF, Moerer P, Weber JL, Padberg GW. Location of facioscapulo humeral muscular dystrophy gene on chromosome 4. *Lancet* 1990;336:651-3.
- 8 Beckmann J, Richard I, Hillaire D, et al. A gene for limb-girdle muscular dystrophy maps to chromosome 15 by linkage. *Comptes Rendus de l'Academie des Sciences (Paris)* 1991;312 series III:141-8.
- 9 Speer MC, Yamaoka LH, Gilchrist JM, et al. Localisation of an autosomal dominant form of limb-girdle muscular dystrophy to chromosome 5q. *Human genome mapping 11*, 1991: abstract 26929.
- 10 Romeo G, Roncuzzi L, Sangiorgi S, et al. Mapping of the Emery-Dreifuss gene through reconstruction of crossover points in two Italian pedigrees. *Hum Genet* 1988;80:59-62.
- 11 den Dunnen JT, Grootsholten PM, Bakker E, et al. Topography of the Duchenne muscular dystrophy (DMD) gene: FIGE and cDNA analysis of 194 cases reveals 115 deletion and 13 duplications. *Am J Hum Genet* 1989;45: 835-47.
- 12 Chelly J, Hamard G, Koulakoff A, Kaplan JC, Kahn A, Berwald-Netter Y. Dystrophin gene transcribed from different promoters in neuronal and glial cells. *Nature* 1990; 344:64-5.
- 13 Lidov HGW, Byers TJ, Watkins SC, Kunkel LM. Localisation of dystrophin to post-synaptic regions of central nervous system cortical neurons. *Nature* 1990;348:725-8.
- 14 Love DR, Hill DF, Dickson G, et al. An autosomal transcript in skeletal muscle with homology to dystrophin. *Nature* 1989;339:55-7.
- 15 Koenig M, Monaco AP, Kunkel LM. The complete sequence of dystrophin predicts a rod-shaped cytoskeletal protein. *Cell* 1988;53:219-28.
- 16 Levine BA, Moir AJG, Patchell VB, Perry SV. The interaction of actin with dystrophin. *FEBS Lett* 1990;263:159-62.
- 17 Koenig M, Kunkel LM. Detailed analysis of the repeat domains of dystrophin reveals four potential hinge segments that may confer flexibility. *J Biol Chem* 1990;265:4560-6.
- 18 Cross RA, Stewart M, Kendrick-Jones J. Structural predictions for the central domain of dystrophin. *FEBS Lett* 1990;262:87-92.
- 19 Lemaire C, Heilig R, Mandel JL. The chicken dystrophin cDNA: striking conservation of the C-terminal coding and 3' untranslated regions between man and chicken. *EMBO J* 1988;7:4157-62.
- 20 Cullen MJ, Walsh J, Nicholson LVB, Harris JB. Ultrastructural localisation of dystrophin in human muscle by using gold immunolabelling. *Proc R Soc Lond* 1990;240: 197-210.
- 21 Campbell KP, Kahl SD. Association of dystrophin and an integral membrane glycoprotein. *Nature* 1989;338:259-62.
- 22 Ibrahimov-Beskrovanaya O, Ervasti JM, Leveille CJ, Slaughter CA, Sernett SW, Campbell KP. Primary structure of dystrophin-associated glycoprotein linking dystrophin to the extracellular matrix. *Nature* 1992;355:696-702.
- 23 Ervasti JM, Campbell KP. Membrane organisation of the dystrophin-glycoprotein complex. *Cell* 1991;66:1121-31.
- 24 Menke A, Jockuses H. Decreased osmotic stability of dystrophin less muscle cells from mdx mouse. *Nature* 1991; 349:334-6.
- 25 Turner PR, Westwood T, Regan CM, Steinhardt RA. Increased protein degradation results from elevated free calcium levels found in muscle from mdx mice. *Nature* 1988; 335:735-8.
- 26 Franco A, Lansman JB. Calcium entry through stretch-inactivated ion channels in mdx myotubes. *Nature* 1990; 344:670-3.
- 27 Koenig M, Beggs AH, Mayer M, et al. The molecular basis for Duchenne versus Becker dystrophy: correlation of severity with type of deletion. *Am J Hum Genet* 1989;45: 498-506.
- 28 Bushby KMD, Gardner-Medwin D, Nicholson LVB, et al. The clinical, genetic and dystrophin characteristics of Becker muscular dystrophy. 2. Correlation of phenotype with genetic and protein abnormalities. *J Neurol* (in press).
- 29 Blonden LAJ, Den Dunnen JT, Van Passen HMB, et al. High resolution deletion breakpoint mapping in the DMD gene by whole cosmid hybridisation. *Nucleic Acids Res* 1989;17:5611-21.
- 30 Love DR, England SB, Speer A, et al. Sequences of junction fragments in the deletion-prone region of the dystrophin gene. *Genomics* 1991;10:57-67.
- 31 Boyce FM, Beggs AH, Feener C, Kunkel LM. Dystrophin is transcribed in brain from a distant upstream promoter. *Proc Natl Acad Sci USA* 1991;88:1276-80.
- 32 Bulman DE, Gangopadhyay SB, Bechuck KG, Worton RG, Ray PN. Point mutation in the human dystrophin gene: identification through western blot analysis. *Genomics* 1991;10:457-60.
- 33 Roberts RG, Bentley DR, Barby TFM, Manners E, Bobrow M. Direct diagnosis of carriers of Duchenne and Becker muscular dystrophy by amplification of lymphocyte RNA. *Lancet* 1990;336:1523-6.
- 34 Van Essen AJ, Abbs S, Bouget M, et al. Parental origin and germline mosaicism of deletions and duplications of the dystrophin gene. A European study. *Hum Genet* 1992;88: 249-57.
- 35 Hoffman EP, Fischbeck KH, Brown RH Jr, Johnson M, Medori R, Loike JD. Characterisation of dystrophin in muscle biopsy specimens from patients with Duchenne's or Becker's muscular dystrophy. *N Engl J Med* 1988;318: 1368-8.
- 36 Nicholson LVB, Johnson MA, Gardner-Medwin D, Bhattacharya SS, Harris JB. Heterogeneity of dystrophin expression in patients with Duchenne and Becker muscular dystrophy. *Acta Neuropathol (Berl)* 1990;80:239-50.
- 37 Bushby KMD, Thambayah M, Gardner-Medwin D. Prevalence and incidence of Becker muscular dystrophy. *Lancet* 1991;337:1022-4.
- 38 Arahata K, Ishihara T, Kamabura K, et al. Mosaic expressions of dystrophin in symptomatic cases of Duchenne's muscular dystrophy. *N Engl J Med* 1989;320:138-42.
- 39 Vainzof M, Pavanello RCM, Pavanello I, Tsanaclis AM, Levy JA, Passos-Bueno MR. Dystrophin immunofluorescence pattern in manifesting and asymptomatic carriers of Duchenne's and Becker muscular dystrophies of different ages. *Neuromuscular Disorders* 1991;1:177-83.
- 40 Bushby KMD. Genetic and clinical correlations of Xp21 dystrophy. *J Inherited Metab Dis* (in press).
- 41 Gospe SM, Lozano RP, Lava NS, Grootsholten BS, Scott MD, Fischbeck KH. Familial X-linked myalgia and cramps: a non progressive myopathy associated with a deletion in the dystrophin gene. *Neurology* 1989;39:1277-80.
- 42 Nordenskjold M, Nicholson L, Edstrom L, et al. A normal male with an inherited deletion of one exon within the DMD gene. *Hum Genet* 1990;84:207-9.
- 43 Monaco AP, Bertelson CJ, Liechti-Gallati S, Moser H, Kunkel LM. An explanation for the phenotypic differences between patients bearing partial deletions of the DMD locus. *Genomics* 1988;2:90-5.
- 44 Nicholson LVB, Johnson MA, Davison K, et al. Dystrophin or a 'related protein' in Duchenne muscular dystrophy? *Acta Neurol Scand* 1991 (in press).
- 45 Nicholson LVB, Bushby KMD, Johnson MA, Den Dunnen JT, Ginjaar IB, Van Ommen GJB. Predicted and observed sizes of dystrophin in some patients with deletions that disrupt the open reading frame. *J Med Genet* (in press).
- 46 Norman AM, Thomas NJT, Kingston HM, Harper PS. Becker muscular dystrophy: correlation of deletion type with clinical severity. *J Med Genet* 1990;27:236-9.
- 47 Chelly J, Gilgenterantz H, Lambert M, Hamard G, Chafey P, Recan D. The dystrophin transcripts in DMD and BMD patients with gene deletion. In: Angelini C, Danielli GA, Fontanari D, eds. *Muscular dystrophy research: from molecular diagnosis toward therapy*. Amsterdam: Excerpta Medical International Congress Series 934, 1991:147-56.
- 48 Carpenter JL, Hoffman EP, Romanul FCA, et al. Feline muscular dystrophy with dystrophin deficiency. *Am J Pathol* 1989;135:909-19.
- 49 Sicinski P, Geng Y, Ryder-Cook AS, Bernard EA, Darlison MG, Barnard PJ. The molecular basis of muscular dystrophy in the MDX mouse—a point mutation. *Science* 1989;244:1578-80.
- 50 Ascadi G, Dickson G, Love DR, et al. Human dystrophin expression in MDX mice after intra muscular injection of DNA constructs. *Nature* 1991;352:815-8.
- 51 Partridge TA. Invited review—myoblast transfer: a possible therapy for inherited myopathies? *Muscle Nerve* 1991;14: 197-212.
- 52 Gussoni E, Pavlath GK, Lanctot AM, et al. Normal dystrophin transcripts detected in Duchenne muscular dystrophy patients after myoblast transplantation. *Nature* 1992;356:435-8.
- 53 Koch MC, Grimm T, Harley HG, Harper PS. Genetic risks for children of women with myotonic dystrophy. *Am J Hum Genet* 1991;48:1084-91.
- 54 Mahadeven M, Tsilifis C, Sabohrin L, et al. Myotonic dystrophy mutation: an unstable CTG repeat in the 3' untranslated region of the gene. *Science* 1992;255:1253-5.
- 55 Fuy-H, Pizzutti A, Fenwick RG Jr, et al. An unstable triplet repeat in a gene related to myotonic muscular dystrophy. *Science* 1992;255:1256-8.