Age related susceptibility to *Campylobacter jejuni* infection in a high prevalence population

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SUMMARY In a year long prospective study of diarrhoea in children under 2 years of age in Soweto, South Africa, *Campylobacter jejuni* was isolated in 18 of 60 children under 9 months of age with diarrhoea, compared with 4 of 60 age matched controls. In the older children, 16 of 51 children with diarrhoea and 17 of 51 control children excreted this organism in their faeces. These results indicate a change in susceptibility to *C jejuni* in children over 9 months of age. Campylobacter enteritis in the young children was usually mild, without macroscopic blood in the faeces, and prolonged excretion of the organism after acute attacks was not infrequent. Breast feeding did not seem to protect against colonisation with *C jejuni*.

In developing countries intestinal *Campylobacter jejuni* infections are common in young children.1-4 In studies in Zaire,1 4 Ruanda,3 South Africa,3 Bangladesh,5 and the Gambia7 an important association between the presence of *C jejuni* in the stool and diarrhoea has been shown. In the South African study *C jejuni* was isolated in 32% of infants less than 9 months old with diarrhoea compared with 4% of controls. In contrast, in children 9–24 months old the isolation of *C jejuni* was similar in the faeces of children with diarrhoea and controls.3 These findings prompted us to re-investigate more critically the age at which children convert from a diarrhoea susceptible to a diarrhoea refractory response to *C jejuni* in the intestinal tract. We investigated concurrently the duration of excretion of *C jejuni* after acute attacks of diarrhoea and evaluated the protective effect of breast feeding on the outcome of infection. We also noted the main clinical features of *C jejuni* associated enteritis and investigated the aetiological role of other bacterial pathogens in diarrhoea in these children.

Materials and methods

Patients. Clinical data on 111 children with diarrhoea admitted to the Baragwanath Hospital near Johannesburg were recorded. Stool specimens were obtained for culture and processed within 4 hours of admission. Mothers of babies in whom *C jejuni* was isolated were asked to bring them back 3 weeks later for further stool examination. Children with positive stool cultures on this occasion were checked weekly until negative results were obtained.

Controls. Within 24 hours each patient with diarrhoea was matched for age and sex with a control baby. These apparently healthy babies came with their mothers to family planning or childcare clinics. We excluded children who had suffered any illness, including diarrhoea, within the previous 2 weeks. Information on feeding was recorded. Faeces were collected from the napkins, transported to the laboratory in Johannesburg, and processed within 4 hours.

Media and bacteriological techniques. Three media were used for the isolation of *C jejuni*. The composition per litre medium was:

- BCA—39 g Columbia agar base (Oxoid Ltd, Basingstoke, England) and 100 ml citrated horse blood;
- BCH—31 g cysteine heart agar (Difco Laboratories, Detroit, USA), 100 ml citrated horse blood, 5 mg novobiocin (Upjohn, Kalamazoo, Michigan, USA), and 1.0 ml 0.5% brilliant green in distilled water;
- Skirrow's medium—40 g oxoid blood agar base No 2, 50–70 ml lysed horse blood, 10 mg vancomycin, 2 500 IU polymyxin B sulfate, and 5 mg trimethoprim lactate (Oxoid Ltd).
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Skirrow's medium was streaked directly with a loopful of faeces and incubated at 42°C. In addition, all 3 media were seeded with 0.2 ml each of a filtrate prepared as described and incubated at 37°C. All media were incubated in an atmosphere of 10% CO2, 18% O2, and 72% N2 for 48 hours. Colonies morphologically resembling C. jejuni were picked, subcultured, and identified according to Holdeman et al. Thermoresistance, sensitivity to nalidixic acid, hippurate hydrolysis, and H2S production were used for identification and biotyping of isolates.

Conventional techniques were used for the isolation and identification (including serotyping) of salmonellas and shigellas. Five Escherichia coli colonies per sample of faeces were tested for antigens associated with enteropathogenicity (EPEC) by using commercially available EPEC antisera (Hoechst, Behring-Werke, Germany; Wellcome Research Reagents, England), for enteroinvasiveness by Sereny's test (EIEC), and for both heat stable and heat labile enterotoxin strains (ETEC) as described by Robins-Browne et al.

**Results**

**Recovery of bacterial pathogens from stool specimens.** C. jejuni was isolated from 31% of children with diarrhoea regardless of their age group (Table 1) and in 19% of the controls. It was the sole pathogen in 15% and 14% respectively in the diarrhoea and control groups. The 13 isolates available for biotyping with hydrolysed hippurate belonged, with 1 exception, to biotype 1.

The incidence of salmonellas and shigellas was 4% or less in all groups. EPEC strains were recovered from 43% of children with diarrhoea and from 23% of controls (P<0.01). ETEC strains of E. coli were detected in 12 patients with diarrhoea and 5 controls. No EIEC were found.

**Age specific faecal excretion of C. jejuni.** There was a statistically significant difference (P<0.01) between the isolation rate of C. jejuni from infants with diarrhoea and from controls up to 8 months (Table 2). This was also true when children excreting C. jejuni as the sole bacterial pathogen were considered (P<0.02). This trend changed conspicuously once children reached the age of 9 months, and between 1 and 2 years of age there was a steady decline in number of children with C. jejuni associated diarrhoea. In contrast, in the control group the incidence of C. jejuni in the faeces increased markedly after 9 months of age. The finding of fewer cases of C. jejuni associated diarrhoea after age 1 year coincides with a general decline in the number of patients in this age group admitted to hospital with diarrhoea.

**Table 1 Faecal excretion of Campylobacter jejuni and other bacterial pathogens in Sowetan children**

<table>
<thead>
<tr>
<th>Group</th>
<th>Age (Months)</th>
<th>No</th>
<th>C. jejuni as sole bacterial pathogens</th>
<th>C. jejuni plus other bacterial pathogens*</th>
<th>Bacterial pathogens other than C. jejuni*</th>
<th>No bacterial pathogens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Diarrhoea</td>
<td>0-8</td>
<td>60</td>
<td>10</td>
<td>8</td>
<td>21</td>
<td>21</td>
</tr>
<tr>
<td></td>
<td>9-24</td>
<td>51</td>
<td>7</td>
<td>9</td>
<td>11</td>
<td>24</td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>111</td>
<td>17</td>
<td>17</td>
<td>32</td>
<td>45</td>
</tr>
<tr>
<td>Controls</td>
<td>0-8</td>
<td>60</td>
<td>1</td>
<td>3</td>
<td>16</td>
<td>40</td>
</tr>
<tr>
<td></td>
<td>9-24</td>
<td>51</td>
<td>14</td>
<td>3</td>
<td>2</td>
<td>28</td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>111</td>
<td>15</td>
<td>6</td>
<td>18</td>
<td>68</td>
</tr>
</tbody>
</table>

*Mainly enteropathogenic Escherichia coli; some children harboured 2 or more pathogens.

**Table 2 Age specific faecal excretion of Campylobacter jejuni**

<table>
<thead>
<tr>
<th>Age group (months)</th>
<th>&lt;4</th>
<th>5-8</th>
<th>9-12</th>
<th>13-16</th>
<th>17-20</th>
<th>21-24</th>
</tr>
</thead>
<tbody>
<tr>
<td>No investigated*</td>
<td>2 × 24</td>
<td>2 × 36</td>
<td>2 × 20</td>
<td>2 × 12</td>
<td>2 × 14</td>
<td>2 × 5</td>
</tr>
<tr>
<td>Total C. jejuni isolations</td>
<td>9 (38)</td>
<td>9 (25)</td>
<td>8 (40)</td>
<td>2 (17)</td>
<td>5 (36)</td>
<td>1 (20)</td>
</tr>
<tr>
<td>Diarrhoeal patients, No (%)</td>
<td>3 (13)</td>
<td>1 (3)</td>
<td>7 (35)</td>
<td>7 (33)</td>
<td>4 (29)</td>
<td>2 (40)</td>
</tr>
<tr>
<td>Controls, No (%)</td>
<td>6 (25)</td>
<td>4 (11)</td>
<td>5 (25)</td>
<td>1 (8)</td>
<td>1 (7)</td>
<td>0 (0)</td>
</tr>
</tbody>
</table>

*The diarrhoea and control groups were equally matched and the numbers are expressed as 2 × the number of each of the respective matched groups.
Clinical picture. The clinical picture of *C. jejuni* diarrhoea in our investigation was relatively mild: bloody stools, shock, acidosis, and hyperpyrexia were rare. Moderate dehydration was, however, common and moderate malnutrition was frequent especially among weaned children. Few of the younger and most of the older children had suffered earlier attacks of gastroenteritis. Clinical recovery was generally fast and most children were discharged home in 7 days without specific antimicrobial treatment.

Breast feeding. Patients aged up to 8 months old in whom *C. jejuni* was the only bacterial pathogen isolated were investigated for the protective effect of partial or complete breast feeding. Table 3 shows that breast feeding did not prevent colonisation with *C. jejuni*. Considering infants aged up to 8 months old as a group, a similar analysis (not tabulated) revealed that 31 of 60 infants with diarrhoea and 46 of 60 control infants were breast fed, indicating that breast feeding prevents some cases of diarrhoea caused by agents other than *C. jejuni* (<P<0.01).

*C. jejuni* excretion after acute gastroenteritis. Eleven children with diarrhoea excreted *C. jejuni* from 3 to 12 weeks and 1 for at least 40 weeks. After the acute attack, faecal samples from this patient were examined weekly from week 3 to week 10 and then fortnightly for the next 10 weeks. The organism was isolated on all but 2 occasions and mixed infections with EPEC, salmonellas, or shigellas were observed 7 times. Diarrhoea occurred during week 15 and week 29 and at the last episode the baby was treated with erythromycin for 5 days. The diarrhoea subsided, faeces were negative for *C. jejuni* 5 weeks later, but the organism reappeared in week 40. Control babies were not available for follow up cultures. As the isolates from these children were not serotyped at the time we do not know to what extent reinfection played a role in the prolonged recovery of *C. jejuni* from their faeces.

Discussion

These findings strongly reinforce earlier observations that *C. jejuni* infection is widespread in black Sowetan children. They also confirm that faecal excretion of the organism in early infancy (<9 months of age) is almost always associated with diarrhoea (<P<0.01). The evidence for an aetiological role for *C. jejuni* in these infants’ illness is further substantiated when the children in whom *C. jejuni* was the sole observed pathogen are considered. Again the difference between the diarrhoea and control groups is statistically significant (<P<0.02).

The decline in *C. jejuni* associated diarrhoea observed with increasing age occurred despite the persistently high excretion rates of *C. jejuni* in the >9 months age group (Table 2). The same trend was observed in Bangladesh where a 2% random sample of patients at a ‘diarrhoea hospital’ showed a decline in the isolation rates of *C. jejuni* in children with diarrhoea from 21% in infants under 1 year of age to 9.5% in patients aged between 1 and 2 years.

Why many older children in Soweto and developing countries do not develop diarrhoea while harbouring *C. jejuni* is not clear. Earlier studies in the Sowetan community indicated a high exposure of families to *C. jejuni* through eating chicken, which is the cheapest meat available to them, and their custom of keeping fowls in their backyards. Bovine intestine is also a cheap and popular source of food while many families keep pet dogs that are commonly fed on poultry scraps. We isolated *C. jejuni* in 26 of 30 (86%) samples of fowl faeces, 4 of 30 (13%) samples of bovine intestine, and 12 of 33 (36%) of dog faeces collected in Soweto. In a controlled study in Sweden an association between the preparation and consumption of chicken and campylobacter enteritis was shown. In the Swedish study sick pets transmitted the disease to 2 patients and 3 definite instances of secondary cases were noted. We also found household spread of *C. jejuni* in Soweto and have recently observed repeated reinfections of different serotypes of *C. jejuni* in Sowetan children who excrete *C. jejuni* for prolonged periods (unpublished data). The extensive exposure of this population to *C. jejuni* may well hold the key to the development of resistance to clinically manifested infections in older children.

The episodes of diarrhoea in our infants excreting *C. jejuni* were only moderately severe and the
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absence of frank blood in their stools was unexpected, although variable results on the prevalence of this feature have been reported in the published reports.10–21 Data from Britain22 suggest that blood in the stools of infants aged from 1 month to 1 year may be appreciably less common than in older patients.19

The apparent lack of protection given by breast feeding against C jejuni colonisation in young infants needs further investigation. The numbers in this study were small and it would be premature to deny a role for specific IgA in the prevention of diarrhoea caused by this organism. Bactericidal antibodies actively produced after infection20 may, however, be more important than those passively acquired through mother’s milk. The nature of host resistance resulting in subclinical and asymptomatic infections needs elucidation and apart from immunological mechanisms competition for, or changes in receptor sites, and other factors affecting intestinal epithelial cell physiology may be responsible.

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References


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