Serum thyrotrophin determination on day 5 of life as screening procedure for congenital hypothyroidism

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SUMMARY In 327 newborns cord blood thyroxine (T4) was 11.8 ± 0.4 μg/100 ml (SEM) (151.9 ± 5.1 nmol/l), and serum thyrotrophin (TSH) 6.7 ± 1.0 μU/ml. Variability was marked for both T4 and TSH. Remeasured in the same patients on the fifth day of life, the TSH level was 3.7 ± 1.0 μU/ml, lower than at birth (P < 0.001), while scattering of TSH values was much smaller, with 99.4% of values <12 μU/ml. TSH level at day 5 was therefore subsequently used as a screening procedure, considering 12 μU/ml as the upper limit of normal.

In 14 of 1805 newborns (0.8%), the TSH level exceeded this limit; in 3 raised TSH levels were subsequently confirmed, and hypothyroidism established by conventional criteria. One was a case of lingual thyroid (TSH > 50 μU/ml) and the other 2 (TSH 27 and 50 μU/ml) iatrogenic caused by massive iodine impregnation of the mother (amniocentesis). High TSH levels (160, 500, and 234 μU/ml) were also found in 3 hypothyroid infants aged less than one month who did not belong to the present screening programme.

The striking difference between the TSH levels observed in normal and in hypothyroid infants during the first weeks of life suggests that serum TSH level at day 5 of life is the best parameter for systematic screening for congenital hypothyroidism.

Mental retardation due to congenital hypothyroidism can be avoided only if adequate replacement treatment is initiated very early, ideally during the first days of life (Smith et al., 1957; Man et al., 1963; Raiti and Newns, 1971; Klein et al., 1972; Rager et al., 1974; Dutau and Rochiccioli, 1975). Recent investigations have shown that the incidence of the disease ranges from 1 in 5000 to 1 in 10,000 (Klein et al., 1974; Dussault et al., 1975b; Fisher, 1975; Walfish et al., 1975b); systematic screening for congenital hypothyroidism in the newborn thus represents an important factor in the prevention of mental retardation.

A diagnosis of hypothyroidism on clinical evidence at birth is rarely possible (Lowrey et al., 1958; König, 1968; Raiti and Newns, 1971; Dutau and Rochiccioli, 1975), and it must depend therefore on biochemical evidence. The problem is that of defining the biochemical change which would be the earliest and most reliable sign of the disease in a screening programme. Dussault and Laberge (1973) developed a screening technique based on radioimmunoassay of thyroxine (T4) on the 5th day of life from an eluate of dried blood. This technique was later adapted and modified by several teams (Walfish et al., 1975; Rochiccioli et al., 1975; Larsen and Broskin, 1975). Others use measurements of thyrotrophin (TSH) in cord blood (Klein et al., 1974; Walfish et al., 1975). Raised TSH has the advantage of being a far more sensitive test of thyroid insufficiency which is capable of detecting the disease in various conditions where the level of circulating hormones has not yet changed (Job et al., 1972; Shenkman et al., 1973; Hayek et al., 1973; Gordin et al., 1974; Barnes, 1975), and in particular where there is ectopic thyroid (Hamilton and Hutchison, 1975).

The object of this study was to define the optimum conditions of screening for congenital hypothyroidism by means of TSH assays. In a preliminary phase the method was selected on the basis of a comparison of TSH levels in cord blood and on day 5 of life in 327 newborns. Cord blood T4 levels were also determined in 273 of them. In the light of the
results, we began systematic screening by determining the TSH level on day 5. The method was applied to all newborns in the maternity section of this hospital. In this report, which covers the first 1478 cases studied by this method, we have tried to define the critical threshold of TSH above which thyroid pathology may be suspected. Particular attention was paid to 14 cases showing values above this critical level, among which we detected 3 cases of congenital hypothyroidism.

Patients

A total of 1805 neonates born in this hospital over a period of 20 months were screened. The maternity section handles 1500–1700 births per year. 60% of the mothers are Belgian (Centre d’Informatique en Gynécologie et Obstétrique, Brussels, 1975, unpublished data) while the other nationalities most frequently encountered are Spanish, Italian, and Moroccan immigrants who have lived in Belgium for periods ranging from a few months to 10 years or more.

Methods

TSH level was measured by radioimmunological analysis. At the end of the reaction the fraction of TSH bound to antibody must be separated from the free fraction. In the first phase of the work this was done by the double antibody technique (Samols and Bilkus, 1964) using the CEA-IRE-SORIN (CIS) TSH Kit (method A). In a second phase separation was achieved by a technique binding the antibody to a solid phase according to Wide’s method (1969) using the Phadebas TSH Kit (Pharmacia AB, Uppsala, Sweden) (method B). The upper and lower limits of sensitivity of these methods were 50 μU/ml and 1·5 μU/ml, respectively. For values greater than 50 μU/ml, the samples must first be diluted with serum containing a known low quantity of TSH (1·5 μU/ml). This value is taken into account when calculating the result.

The T4 level was determined by the method of Murphy and Pattee (1964). The triiodothyronine (T3) level was determined by radioimmunoassay, using the method described by Hesch and Evered (1973).

Investigations

Preliminary phase. Serum TSH and T4 were measured in the cord blood of 327 randomly-selected newborns, and TSH was remeasured in the same patients on day 5 after birth. At that time, 0·4 cm³ of whole blood was collected by heel puncture. The following information was also collected for analysis: general thyroid history of the mother before the pregnancy, course of the pregnancy, duration of periods of dilatation and expulsion, condition of the baby at birth, gestational age, sex, weight, length, T4 and TSH levels in cord blood, TSH on day 5, name and nationality of parents. A correlation was sought between each of the three biochemical measurements and the duration of dilatation, expulsion, and gestational age respectively, and also between the TSH level in the cord blood and that measured on day 5. Any differences between these results in relation to the nationality of the parents were also sought.

Systematic screening. On the basis of the results obtained above, systematic screening was undertaken by TSH determination on day 5 of life in 1478 infants born between 1 July 1974 and 14 July 1975, i.e. 96·4% of the liveborn infants. Average age at the time of sampling was 5 days ±0·5(SD).

Method A was used during the preliminary phase (327 patients) and for the first 577 patients of the second phase. In the other 901 patients TSH was measured by method B.

For measurements in cord blood the critical level of TSH was set at 20 μU/ml in line with Fisher’s recommendations (1973). For measurements taken on day 5 (discussed below), the threshold was arbitrarily lowered to 12 μU/ml, both during the preliminary phase of the work and during systematic screening. Infants showing values greater than this critical level were given further tests, namely determination of serum TSH, T4, and in a few cases T3. These tests were conducted as soon as possible, together with careful clinical examinations and x-rays of the knee and foot to determine bone matura-

Hoerr (1955) were used for the knee and those of Hoerr et al. (1962) for the foot. In cases where these examinations indicated possible hypothyroidism, a thyroid scintigram was carried out immediately, using 99mTc pertechnetate according to the technique proposed by Hayek et al. (1971) to determine the exact location and volume of the thyroid. When clinical, biochemical, and x-ray examinations were normal, the infants were followed up. In cases where hypothyroidism was confirmed, the infants were treated immediately with maximum doses of thyroid extracts, according to the technique of Petricciani et al. (1971) (120–150 mg/m² per day).

As reported below, abnormal TSH values were found in 3 newborns whose mothers had undergone amniocentesis. These x-rays had been taken 24 hours after amniocentesis and intra-amniotic injection of 30 ml diatrizoic acid and 12 ml iodophthalein (Lipiodol) containing 6·2 g iodine. Simultaneous injection of these two contrast products allows
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![Comparison of frequency distributions of TSH values in cord blood and on day 5 of life in 327 newborns, and of T4 values in cord blood in 273. Conversion: Traditional units to SI—Thyroxine: 1 μg/100 ml = 12.87 nmol/l.](image)

Radiographic visualization of the digestive tract and fetal skin covering. The injection had been performed 24 to 48 hours before delivery.

Results

The Fig. shows the distribution of TSH levels in the cord blood and on day 5 in the 327 newborns, and of T4 levels in cord blood in 273 of them. The distribution of T4 values in the cord blood is symmetrical, mean 11.8 ± 0.4 μg/100 ml (151.9 ± 5.1 nmol/l) (SEM). TSH levels measured at the same time show a distinctly asymmetrical distribution, with a marked spread of results towards high values. The geometrical mean of TSH values in the cord blood is 6.7 ± 1.0 μU/ml (SEM). TSH values on day 5 are less variable, and the asymmetry towards high values is much less pronounced than for cord blood. The geometrical mean is 3.7 ± 1.0 μU/ml (SEM), significantly lower than that at birth (P < 0.001).

Distinctly abnormal values for cord T4 were shown in 2 infants. In the first, the value was 2.1 μg/100 ml (27 nmol/l), whereas T3 resin uptake exceeded 160% and the thyroxine-binding-globulin (TBG) saturation capacity was 17 μg thyroxine-iodine/100 ml (1340 nmol/l); TSH was 2.2 μU/ml. These results suggest a partial congenital deficiency of TBG. In the second infant the T4 level was 24.6 μg/100 ml (317 nmol/l), showing no clinical signs of hyperthyroidism. No further tests could be done.

For cord blood TSH, 9 infants showed values greater than 20 μU/ml. The results are given in Table 1. In 5 cases expulsion had lasted more than an hour and in 1 case had necessitated use of a vacuum extractor. In another pre-eclampsia occurred at the end of the pregnancy. The T4 levels measured in 8 out of 9 patients were normal and control TSH measurements on day 5 were all normal. For TSH levels on day 5, 99.4% of the results lay below 12 μU/ml, which is why this value was regarded as the critical threshold. Results greater than this were obtained in 4 patients (discussed below).

Table 2 gives the correlation coefficients for T4 and TSH levels in the cord blood and for TSH on day 5 and duration of dilatation, duration of expulsion, and gestational age. There was no significant

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Cord blood</th>
<th>Control day 5</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>TSH (μU/ml)</td>
<td>T4 μg/100 ml (nmol/l)</td>
</tr>
<tr>
<td>1</td>
<td>28.8</td>
<td>12.9 (166)</td>
</tr>
<tr>
<td>2</td>
<td>25.2</td>
<td>9.9 (127)</td>
</tr>
<tr>
<td>3</td>
<td>24.6</td>
<td>12.2 (157)</td>
</tr>
<tr>
<td>4</td>
<td>24.0</td>
<td>15.3 (197)</td>
</tr>
<tr>
<td>5</td>
<td>25.8</td>
<td>9.8 (126)</td>
</tr>
<tr>
<td>6</td>
<td>23.8</td>
<td>10.1 (130)</td>
</tr>
<tr>
<td>7</td>
<td>29.2</td>
<td>8.9 (115)</td>
</tr>
<tr>
<td>8</td>
<td>21.9</td>
<td>9.2 (118)</td>
</tr>
<tr>
<td>9</td>
<td>24.6</td>
<td>-</td>
</tr>
</tbody>
</table>
correlation between these parameters except between T4 in cord blood and gestational age \((r = 0.159, P < 0.01)\) and TSH in cord blood and day 5 \((r = 0.309, P < 0.001)\). No significant differences were observed in the parameters of Belgian, Spanish \((P > 0.06)\), Italian \((P > 0.06)\), and Moroccan \((P > 0.2)\) newborns.

**Systematic screening.** 1478 newborns were screened, representing 96.4% of births in the maternity section during the 20 months of the study. The average value of TSH in 577 patients using method A, as in the preliminary phase, was \(4.2 \pm 1.0\) \(\mu\)U/ml (SEM). The value obtained in 901 patients according to method B was \(1.7 \pm 1.0\) \(\mu\)U/ml (SEM), i.e. significantly lower \((P < 0.001)\). Despite this difference in the mean values, we kept the critical threshold at 12 \(\mu\)U/ml because values below this level were observed in 99.3% and 99.2% of the cases, respectively, for both methods used; the corresponding value obtained during the preliminary phase was 99.4%.

Fourteen out of the 1805 infants studied \((0.8%\) of the cases) showed TSH values on day 5 greater than 12 \(\mu\)U/ml and were tested further, as described above. The data are summarized in Table 3. TSH on day 5 ranged from 12.3 to \(>50\) \(\mu\)U/ml. The age of the infants at the time of the control tests ranged between 9 and 68 days, 11 of them being less than 28 days.

These tests gave normal results in 11 out of 14 infants. These infants were followed-up for 1 to 12 months. None exhibited any further thyroid problem. In one (Case 23) the additional tests showed a lingual thyroid: on day 5 of life this infant had a TSH level greater than 50 \(\mu\)U/ml. Subsequent analyses of cord blood showed TSH 15.5 \(\mu\)U/ml; T4 16.8 \(\mu\)g/100 ml (216 nmol/l); T3 40 ng/100 ml. At 21 days of age clinical and x-ray examinations were normal, as were T4 levels (8.7 \(\mu\)g/100 ml; 112 nmol/l) and T3 (210 ng/100 ml). However, TSH was still high (50 \(\mu\)U/ml). Thyroid scintigram showed an ectopic thyroid located at the base of the tongue. The infant was treated with thyroid extracts from the 32nd postnatal day. TSH reverted to normal in a few days and clinical and biochemical findings were still normal at the age of 8 months. It is obvious that in this particular case thyroid extracts should be given lifelong.

The last 2 patients (Cases 10 and 22) had TSH values of 27 and 50 \(\mu\)U/ml respectively on day 5.

### Table 2: Correlations between biochemical parameters measured at birth and on day 5, and the durations of cervical dilatation and expulsion from uterus, and gestational age in 273 infants

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Coefficient of correlation</th>
<th>Significance</th>
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<tbody>
<tr>
<td>Cord TSH-duration dilatation</td>
<td>-0.034</td>
<td>NS</td>
</tr>
<tr>
<td>Cord TSH-duration expulsion</td>
<td>0.033</td>
<td>NS</td>
</tr>
<tr>
<td>Cord T4-duration dilatation</td>
<td>-0.064</td>
<td>NS</td>
</tr>
<tr>
<td>Cord T4-duration expulsion</td>
<td>0.041</td>
<td>NS</td>
</tr>
<tr>
<td>Cord TSH-gestational age</td>
<td>-0.013</td>
<td>NS</td>
</tr>
<tr>
<td>5th day TSH-gestational age</td>
<td>-0.054</td>
<td>NS</td>
</tr>
<tr>
<td>Cord T4-gestational age</td>
<td>0.159</td>
<td>P &lt; 0.01</td>
</tr>
<tr>
<td>Cord TSH-TSH on day 5</td>
<td>0.309</td>
<td>P &lt; 0.001</td>
</tr>
</tbody>
</table>

### Table 3: Findings in 14 of 1805 patients having TSH levels \(>12\) \(\mu\)U/ml on day 5

<table>
<thead>
<tr>
<th>Case no.</th>
<th>TSH day 5 ((\mu)U/ml)</th>
<th>Age (d)</th>
<th>Clinical signs</th>
<th>Bone age</th>
<th>TSH ((\mu)U/ml)</th>
<th>T4 (\mu)g/100 ml (nmol/l)</th>
<th>Thyroid scintigram</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>27.0</td>
<td>9</td>
<td>Apathy, jaundice</td>
<td>N</td>
<td>14.7</td>
<td>14.8 (190)</td>
<td>N</td>
<td>Amniofetography; treatment from day 13</td>
</tr>
<tr>
<td>11</td>
<td>13.8</td>
<td>68</td>
<td>N</td>
<td>N</td>
<td>3.7</td>
<td>8.1 (104)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>12</td>
<td>21.6</td>
<td>9</td>
<td>N</td>
<td>N</td>
<td>4.1</td>
<td>11.2 (144)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>13</td>
<td>13.8</td>
<td>30</td>
<td>N</td>
<td>N</td>
<td>5.1</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>14</td>
<td>21.0</td>
<td>13</td>
<td>N</td>
<td>N</td>
<td>8.9</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>15</td>
<td>12.5</td>
<td>41</td>
<td>N</td>
<td>N</td>
<td>2.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>16</td>
<td>18.0</td>
<td>27</td>
<td>N</td>
<td>N</td>
<td>3.2</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>17</td>
<td>36.5</td>
<td>17</td>
<td>N</td>
<td>N</td>
<td>5.6</td>
<td>7.0 (90)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>18</td>
<td>27.0</td>
<td>18</td>
<td>N</td>
<td>N</td>
<td>1.9</td>
<td>9.0 (116)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>19</td>
<td>13.5</td>
<td>14</td>
<td>N</td>
<td>N</td>
<td>1.5</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>20</td>
<td>14.0</td>
<td>-</td>
<td>N</td>
<td>N</td>
<td>2.3</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>21</td>
<td>23.5</td>
<td>25</td>
<td>N</td>
<td>N</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>22</td>
<td>50.0</td>
<td>16</td>
<td>Goitre</td>
<td>N</td>
<td>&gt;250</td>
<td>3.2 (41)</td>
<td>Large goitre</td>
<td>Amniofetography; treatment from day 23</td>
</tr>
<tr>
<td>23</td>
<td>&gt;50.0</td>
<td>21</td>
<td>N</td>
<td>N</td>
<td>50</td>
<td>8.7 (112)</td>
<td>Lingual thyroid</td>
<td>Treatment from day 32</td>
</tr>
</tbody>
</table>

N = normal.
Case 10 presented apathy and extremely slow sucking on the 9th day. TSH was then still high (14·7 μU/ml); T4 and thyroid scintigram were normal. Case 22 at the age of 16 days presented clinically apparent thyroid hyperplasia, confirmed by thyroid scintigram. TSH at that point was greater than 250 μU/ml and T4 had fallen to 3·2 μg/100 ml (41 nmol/l). These 2 infants were also treated with thyroid extracts and their clinical and biochemical signs rapidly returned to normal. Treatment was withdrawn after 23 and 16 days respectively. Further controls for TSH and T4 in serum gave normal values. It is particularly significant that the mothers of these 2 infants had each undergone amniofetography on the day before their delivery. The same examination had been performed 2 days before delivery on the mother of Case 17 whose TSH had reached 36·5 μU/ml on day 5 but reverted to normal spontaneously on day 17.

During these investigations diagnosis of congenital hypothyroidism was confirmed in 3 other infants, aged 26, 27, and 3 days, who had been sent to the paediatric department with probable diagnosis of thyroid insufficiency based on clinical evidence. The first 2 presented lingual thyroids, the third a probable metabolic goitre with hypothyroidism and no signs of iodine impregnation. TSH levels were 160, 500, and 234 μU/ml respectively, and T4 2·4, 3·0, and 7·2 μg/100 ml (31, 39, 93 nmol/l) respectively.

Discussion

The results indicate that determination of serum TSH level on day 5 of life is an eminently suitable technique for detecting congenital hypothyroidism in the newborn. Contrary to what is observed at birth, the variability of individual results on the 5th day is small. The sensitivity of the method is confirmed by the substantial deviation observed between the normal mean, i.e. 1·7±1·0 μU/ml (method B), and the pathological cases for which the values obtained were 15–50 times higher.

The controversial question is whether raised TSH alone constitutes sufficient evidence on which to base a diagnosis of hypothyroidism and to justify treatment (Herschman and Pittman, 1971; Ikram et al., 1973; Evered et al., 1973; Jackson et al., 1975). A moderate, but sometimes substantial, increase in TSH has been observed in the initial stage of development of a goitre regarded as euthyroid, either idiopathic (Young et al., 1975) or due to iodine deficiency (see review in Delange, 1974). In the latter condition a state of thyroid compensation can be maintained by permanently increased thyrotrophic stimulation. Various, perhaps transient, conditions can cause increased TSH in the newborn (Fisher and Sack, 1975) and do not necessarily imply hypothyroidism leading to mental retardation. However, out of the 14 abnormal TSH values observed on day 5 in these infants, 3 infants were still abnormal at the time of subsequent control examinations and 2 of these presented definite thyroid pathologies, namely lingual thyroid and goitre with hypothyroidism. Furthermore, various authors have reported raised TSH values in a number of conditions leading to thyroid insufficiency before any perceptible drop in the level of thyroid hormones or any clinical signs have been noted (Job et al., 1972; Hayek et al., 1973; Gordin et al., 1974). In this respect, raised TSH is one of the important biochemical indications of a state of premynxoedema (Ikram et al., 1973) or compensated hypothyroidism (Hayek et al., 1973; Barnes, 1975) which may develop into definite hypothyroidism, therefore justifying treatment with thyroid extracts (Hayek et al., 1973; Ikram et al., 1973; Evered et al., 1973; Young et al., 1975; Barnes, 1975; Rallison et al., 1975; Herrmann et al., 1975). The rise in TSH is an indicator whose sensitivity and specificity are useful for systematic screening.

As several authors have suggested (Klein et al., 1974; Fisher, 1975; Walfish et al., 1975), the sensitivity of the TSH method seems to give it a decisive advantage over screening based on T4 determination (Dussault et al., 1975b): the measurement of T4 undoubtedly offers considerable advantage from the public health view, especially its low cost and the possibility of performing the assay in an eluate of dried blood. However, the cases of hypothyroidism reported here confirm the view (Fisher and Sack, 1975) that myxoedema can be accompanied by no more than slightly abnormal serum T4 levels in the newborn. This was so with the infant with the lingual thyroid, in whom TSH on day 5 was, in contrast, distinctly increased. This newborn would have escaped detection based on T4 measurements on day 5 of life, or a method based on cord TSH. As recently underlined by Fisher and Sack (1975), this point is particularly important in the detection of congenital hypothyroidism because the most frequent cause of this disease is thyroid dysgenesis (Neumann et al., 1958; Little et al., 1965; Dutau and Rochiccioli, 1975) in which the thyroid deficit is only partial. The greater sensitivity of the TSH method is also confirmed by the study of the 3 cases of clinically detectable hypothyroidism in which, as other authors have reported (Dutau and Rochiccioli, 1975; Dussault et al., 1975b; Jackson et al., 1975), deviations of TSH levels from the normal mean were much more pronounced than those of T4.
The main disadvantages of screening based on TSH measurement are its fairly high cost and the fact that it has hitherto required a whole blood sample. The cost may be affected substantially by a number of factors, but it is obvious that 5000–10 000 TSH analyses cost substantially less than society pays out for the care of a gravely mentally handicapped person over several decades. The problem of TSH elution from a whole blood sample collected on paper is currently being investigated in a number of laboratories (Dussault et al., 1975a; Foley et al., 1975), including our own. We cannot preclude the possibility that subjects with normal serum TSH, or T4 concentrations on day 5, would not develop hypothyroidism later on. Further studies are necessary to clarify this point.

Our results concerning the distribution of T4 and TSH levels in the cord blood confirm results reported by other authors (see review in Similä et al., 1975). Our findings do not confirm the relationship hypothesized by Walfish et al. (1975) between the level of TSH in the cord and the duration of the delivery. However, in view of the lack of any major obstetrical pathologies in our series of cases, this hypothesis is not ruled out. The correlation observed between the level of T4 in the cord blood and the gestational age of the baby ties in with the findings of others (Fisher et al., 1970; Greenberg et al., 1970) who have reported a progressive increase in the level of T4 during fetal growth.

The rise in TSH in the 3 newborns whose mothers had undergone amniofetography at the end of their pregnancies ties in with a similar observation reported by Bühler et al. (1973). The presence of a moderately increased (Case 10) or substantially increased (Case 22) TSH level with a normal or low T4 level associated with thyroid hyperplasia after the administration of large doses of iodine to the mother suggests a partial blockage of organification due to a Wolff-Chaikoff effect (Wolff and Chaikoff, 1948). But this does not account for the fact that, paradoxically, the thyroid glands of these infants trap sufficient technetium to allow visualization of the gland, whereas this is not the case in severe iodine contamination. The possibility of a pre-existent anomaly of thyroid metabolism cannot be ruled out in our 3 patients since the follow-up period is not yet long enough to allow a definite answer to this question. The strong thyrotrophic stimulation observed in newborns whose mothers had undergone amniofetography calls for thorough investigation. In any event, it seems that the thyroids of such newborns should be tested systematically shortly after birth in order to test whether this investigation is safe (Amy and Mandelman, 1973).

In conclusion, our investigations strongly suggest that the level of TSH measured on the 5th day of life is a suitable parameter for systematic screening for hypothyroidism in newborns. The prospect of combining this method with screening for amino-acidopathies in whole blood sampled on paper is of major practical interest. It is still too early to fix a critical level of TSH which would reliably indicate thyroid pathology in the newborn. On the basis of a critical threshold of 12 μU/ml, our investigations showed 14 abnormal values in a total of 1805 cases, of which only 3 were confirmed by subsequent examinations. Using the threshold value of 20 μU proposed by Fisher (1973) the number of abnormal values would be reduced to 8, or 0.4%, and would include the 3 cases of confirmed hypothyroidism. The threshold of 20 μU/ml therefore seems reasonable, but remains to be confirmed.

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References


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Addendum

While this paper was being prepared, Klein *et al.* (1976) confirmed the sensitivity of the TSH test, but Dussault *et al.* (1976) showed that TSH was normal in 3 out of 28 cases of neonatal hypothyroidism detected on the basis of decreased T4, i.e. in 10 to 15% of the cases. The possibility thus arises that screening based on TSH assay could result in 'false negatives', especially in pituitary or hypothalamic hypothyroidism. As the present report indicates that T4 screening could result in 'false negatives' in primary hypothyroidism, the systematic measurement of both T4 and TSH during the first days of life as proposed by Buist *et al.* (1975) should be seriously considered.

References

