Estimation of glomerular filtration rate from plasma creatinine concentration in children

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The relation between the true plasma creatinine concentration (Pc) and the glomerular filtration rate corrected for body surface area (GFR/SA) was investigated in 108 individuals, and the following formula was derived: GFR/SA (ml/min per 1·73m²SA) = 0·43 Ht (cm)/Pc (mg/100 ml). This formula was tested in a second group of 83 children, and its accuracy and precision was compared to the 24-hour creatinine clearance. It was found to be superior to the creatinine clearance overall, and was as good, even if all results involving suspect 24-hour-urine collections were eliminated from analysis. The formula in SI usage is: GFR/SA (ml/min per 1·73 m²SA) = 38 Ht (cm)/Pc (μmol/l).

The estimation of glomerular filtration rate (GFR) is of central importance in assessment of renal function. However, the most precise methods are too cumbersome for routine clinical use, and in practice a compromise between accuracy and simplicity has to be sought. The 24-hour creatinine clearance (Cc) is widely used to estimate GFR, but difficulties in the accurate collection of timed urine samples, particularly from children, limit its value, and even under research conditions its reproducibility is poor (Chantler and Barratt, 1972). Because creatinine excretion is approximately constant, an inverse relation exists between GFR and the plasma creatinine concentration (Pc), and formulae for the estimation of GFR from Pc in adults have been presented (Doolan, Alpen, and Theil, 1962; Jelliffe, 1971; Cockcroft and Gault, 1976). In healthy children, however, Pc rises with age (Donckerwolcke et al., 1970), whereas GFR, corrected in accordance with convention for body surface area (SA) does not do so after the age of 2 years. It is therefore apparent that the relation between Pc and GFR/SA in children must reflect some further aspect of body size. Taking into account the fact that creatinine excretion (UcV; Uc = urine creatinine concentration, V = urine flow rate) is proportional to body weight (Wt) (Graystone, 1968), we argued that GFR/SA should be proportional to body height (Ht) and inversely proportional to Pc:

(i) GFR ∝ Cc = UcV/Pc
(ii) UcV ∝ Wt
(iii) Wt ∝ cube of height (Ht³)
(iv) SA ∝ square of height (Ht²),

substituting the proportionalities (ii), (iii), and (iv) in (i):

(v) GFR/SA ∝ Ht/Pc = a Ht/Pc.

The same conclusion was reached empirically by Schwartz et al. (1976).

We analysed the data from a first group of 108 individuals to estimate the proportionality constant a, and then compared the accuracy of the estimation of GFR from equation (v) with the 24-hour Cc in a second group of 83 individuals; we found that the estimate of GFR/SA from Pc was as good as that obtained from Cc.

Patients

Group I consisted of 103 children with renal disease and 5 healthy adults; the children were between 2 months and 14 years of age, and had various renal diseases but were not oedematous. They were in a
stable state with GFR in the range 4–200 ml/min per 1·73 m²SA. For group II, 83 children with kidney disease consecutively admitted to the renal ward, who had both Cc and GFR determined within an 8-day period, were identified. In both groups only the first relevant result in each child was recorded for analysis.

**Methods**

GFR was estimated from the plasma clearance of 51-chromium edetic acid (⁵¹Cr-EDTA) after intravenous injection. Blood samples were collected at 2 and 4 hours and GFR calculated by a single exponential analysis incorporating the correction factor described by Chantler and Barratt (1972). Surface area was derived from height and weight using the formula of Du Bois and Du Bois (1916) and GFR was corrected to 1·73 m²SA.

In group I, Pc was determined manually in duplicate by the Jaffe reaction after adsorption onto an ion-exchange resin to remove noncreatinine chromogens (Stoten, 1968), and also by an automated method (Technicon, 1966). The latter method includes some noncreatinine chromogens, and was found to overestimate the true creatinine concentration by 0·14 mg/100 ml ± 0·13 (SD). In group II only the automated method was used, and the true value for Pc estimated by subtracting 0·14 mg/100 ml. Throughout this paper, Pc refers to the true plasma creatinine concentration.

Creatinine clearance was calculated from a single 24-hour urine collection and a single determination of the plasma creatinine concentration by the automated method without the correction for noncreatinine chromogens. Urine creatinine concentration was measured by the same automated method as for plasma with different dilution factors. Urine collections were judged to be accurate if the creatinine excretion fell within the following limits (Ghazali and Barratt, 1974):

\[ U_c = \frac{V}{Wt} \times (\text{mg/kg body weight per day}) = 15 + 0·5 \times \text{Age (years)} \pm 6 \] (2SD). 26 (31%) of the 83 24-hour urine collections failed to satisfy this criterion.

**Results**

**Relation of Pc, Ht, and GFR/SA (group I).**

The relation between Pc and GFR/SA in the first group of individuals is shown in Fig. 1. The data have been treated logarithmically to stabilize variances. The symbols indicate groups of patients of different height, and the effect of height on the relation between Pc and GFR/SA is apparent.

Theoretical considerations given above suggested the following relation:

\[ \text{GFR/SA} = a \times \frac{\text{Ht}}{P_c}, \]

which was also indicated by the plot of log GFR/SA against log Ht/Pc (Fig. 2). The more general form was first considered,

\[ \text{GFR/SA} = a \times P_c^c, \]

which is equivalent to

\[ \log \text{GFR/SA} = \log a + b \times \log \text{Ht} + c \log P_c. \]

A multiple linear regression was undertaken. The test of regression \( b = c = 0 \) was highly significant, as was the test for including \( b \) in the model given that \( c \) was included. However, the estimates of \( b \) and \(-c\) did not differ significantly from unity. It was therefore assumed that \( b = -c = 1 \), i.e. log GFR/SA = log a + log Ht - log Pc.

The least-squares estimate of log a was log a = -0·37, estimated standard error = 0·01.

The estimated standard error of the predicted value of log GFR/SA from the observed value of
log Ht and log P<sub>c</sub> is 0·14. This model accounted for 87% of the variation in log GFR/SA. Its validity was tested by calculating and examining the residuals, and there was good evidence that they were normally distributed. From this analysis the best estimate of \( a \) is 0·43. Therefore, to predict GFR/SA from Ht and P<sub>c</sub> the relation is GFR/SA = 0·43 Ht/P<sub>c</sub>.

The 95% confidence limits for the predicted value of GFR/SA are 0·43 Ht/P<sub>c</sub><sup>10±0·28</sup>, i.e. 52 and 190% of the estimated value.

**Prediction of GFR/SA from C<sub>c</sub> and P<sub>c</sub> (group II).**

**C<sub>c</sub> and GFR/SA.** The relation between the 24-hour C<sub>c</sub>/SA and the measured GFR/SA in all the 83 children in group II is shown in Fig. 3. The

[Diagram: Fig. 3.—Relation between the 24-hour C<sub>c</sub>/SA and GFR/SA (group II).]

The correlation is highly significant, but the scatter of the observations is wide (\( r = 0·89, \ P = <0·001 \)). When the 26 children with inadequate urine collections were excluded from the analysis, the relation of C<sub>c</sub>/SA and GFR/SA was better (\( r = 0·94, \ P = <0·001 \)), and the scatter of observations narrower (Fig. 4).

0·43 Ht/P<sub>c</sub> and GFR/SA. GFR/SA predicted by the formula derived from the analysis of group I correlated well with the measured GFR/SA (\( r = 0·95, \ n = 83, \ P = <0·001 \)) (Fig. 5).

**Comparison of 0·43 Ht/P<sub>c</sub> and C<sub>c</sub>/SA as estimate of GFR/SA.** In order to compare the two methods of estimating GFR, the mean and variances of their individual differences from the measured GFR were analysed. The data were again treated logarithmically, and for each individual a value for dA and dB calculated, where dA = log GFR/SA—log (0·43 Ht/P<sub>c</sub>), dB = log GFR/SA—log C<sub>c</sub>/SA.

The mean value for dA was not significantly different from zero (Table) whereas dB for the whole group of 83 children was significantly positive,
indicating an underestimate by Cc/SA. However, when the 26 estimates with inadequate urine collection were excluded, dB was not significantly different from zero.

The variances of dA and dB were compared as for a paired sample (see for example Armitage, 1971). When the 26 individuals were excluded as above, there was no significant difference between them (P > 0.2), but when all 83 individuals were considered the variance of dB was significantly greater than that of dA (P < 0.001). This analysis implies that 0.43 Ht/Pc as an estimate of GFR/SA is superior to Cc/SA overall, and is just as good even when criteria are applied to exclude estimates of Cc/SA based on 24-hour urine collections of dubious adequacy.

Discussion

This study shows that measurement of Pc provides a simple and reliable method of estimating GFR in children, provided that height is taken into consideration. The formula GFR/SA (ml/min per 1.73 m²SA) = 0.43 Ht (cm)/Pc (mg/100 ml) is based on the true plasma creatinine concentration; using the commonly available automated method, 0.14 mg/100 ml should be subtracted to allow for noncreatinine chromogens. If SI units are used, and plasma creatinine concentration expressed in μmol/l, the formula should be rewritten: GFR/SA (ml/min per 1.73 m²SA) = 38 Ht (cm)/Pc (μmol/l) and 12.4 μmol/l should be subtracted from the automated estimates of Pc.

Our statistical analysis has shown that this method of estimating GFR is as accurate and as precise as measuring the 24-hour Cc, even when urine is collected on a ward specializing in the care of children with renal disease, and when results are eliminated if the creatinine excretion does not fall within the normal range, implying erroneous urine collections.

Our conclusion, therefore, is that the 24-hour Cc should be abandoned as an estimate of GFR in children except in special circumstances such as emaciation where creatinine production may be low, in changing states where the actual Pc does not represent the equilibrium value, and in special tests where Cc can be used as an internal reference clearance. It is possible, however, that greater precision may be obtained with shorter urine collections for measurement of Cc, but we have not examined this point.

Schwartz et al. (1976) have undertaken a similar analysis to our own, and present the formula: GFR/SA (ml/min per 1.73 m²SA) = 0.55 length (cm)/Pc (mg/100 ml). The difference between their value of 0.55 and ours of 0.43 can be explained by the fact that they did not measure true plasma creatinine concentration, and used the inulin clearance as reference estimate of GFR.

It should be recognized, however, that the error of estimation of GFR/SA from Pc is large, the 95% confidence limits being 52–190%. Nevertheless, measurement of 24-hour Cc does not reduce this error, and if more precise estimates of GFR are necessary, other methods, such as the plasma clearance of 51Cr-EDTA should be used.

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References


Bois, D. J., and Do Bois, E. F. (1916). Clinical calorimetry. X. Formula to estimate the approximate surface area if height and weight are known. Archives of Internal Medicine, 17, 863.


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