Ascorbic acid in fetal human brain

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Adlard, B. P. F., De Souza, S. W., and Moon, S. (1974). Archives of Disease in Childhood, 49, 278. Ascorbic acid in fetal human brain. Ascorbic acid concentrations in fetal human forebrain in the period 11 to 19 weeks' gestational age were 4 to 11 times higher than those of adults. Levels fell progressively with increasing gestational age but, in term babies dying within 4 weeks of birth, were still at least 3 times those of adults. It was confirmed that, in women delivering at term, ascorbic acid concentrations are approximately 4 times higher in cord blood plasma than in maternal blood plasma. The possible importance of ascorbic acid for normal human brain development is discussed.

The adult brain of several mammals including man has a high concentration of ascorbic acid when compared with most other organs (Yavorsky, Almaden, and King, 1934). In the brain of the immature rat, a species which synthesizes ascorbic acid, levels are even higher than those of adult brain and fall progressively during the major period of brain growth (Allison and Stewart, 1973; Adlard, De Souza, and Moon, 1973). To determine whether a similar phenomenon occurs in man, ascorbic acid concentrations have been examined in developing human brain.

Early reports suggested a marked fetal/maternal plasma gradient for ascorbic acid at term (Teel, Burke, and Draper, 1938; Snelling and Jackson, 1939). We have confirmed this finding and have attempted to investigate the possibility that such a gradient might result solely from changes in maternal levels during labour. For comparison, plasma ascorbic acid has been examined in groups of women at different stages of pregnancy and in women who were not pregnant.

Methods

Gestational age was calculated from the first day of the last menstrual period. Brains were obtained from three sources. Group 1, induced abortions, 11 to 19 weeks' gestational age (no. = 16); group 2, 10 perinatal deaths and one 3-month-old baby; group 3, adults (17-83 years), dying in hospital of non-neurological disease (no. = 6). Post mortem loss of brain ascorbic acid under normal refrigerated conditions was found to be negligible when up to 96 hours elapsed between death and availability of the specimen. Most brains were obtained well within this time.

Changes in fetal tissue levels might reflect differences in maternal intake of vitamin C, which was not assessed. However, mothers of babies dying in the perinatal period (group 2) were not prescribed ascorbic acid during pregnancy. It seems unlikely that women contemplating termination of pregnancy (group 1) would take extra amounts of vitamin C. Finally, the hospital diet which the adults (group 3) received was considered to contain an adequate amount of vitamin C.

Ascorbic acid was determined in whole forebrain in group 1, in a representative slice of frontal cortex (including underlying white matter) in group 3, and by either of these procedures in group 2. In groups 2 and 3 ascorbic acid was also estimated in portions of brainstem (medulla) and cerebellum. Results using the dinitrophenylhydrazine procedure (Roe and Kuether, 1943) are presented. The method has been widely used in animal tissues and seems to be highly specific for ascorbic acid in brain since chromogenic material almost disappears from guinea-pig brain when they are fed a diet free of vitamin C (Hughes, Hurley, and Jones, 1971), and a specific gas-liquid chromatographic method gives almost identical results when applied to both adult and immature rat brain (Allison and Stewart, 1973). Nevertheless, some human forebrain samples were also analysed by the dichlorophenolindophenol procedure (Evelyn, Malloy, and Rosen, 1938). Despite the different assay principles involved, this method gave results very similar to those obtained by the Roe and Kuether procedure.

Water content was determined by drying to constant weight and protein by the method of Lowry et al. (1951). Blood was collected from 13 healthy nonpregnant women not taking contraceptive steroids and from 33 women at different stages of pregnancy. In a further 16 women, delivering at term (38-42 weeks) after an uncomplicated pregnancy, blood was collected at the

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Results

Fetal forebrain ascorbic acid concentrations were very high and fell substantially with age (Fig.). In the period from 11 to 19 weeks' gestational age, levels varied between 4 and 11 times the mature values. 7 fetal brains from this age group, having a mean ascorbic acid level of 0.85 mg/g (range 0.45–1.25), showed a mean water content of 909.1 mg/g and protein concentration of 50.4 mg/g. Thus the mean concentration of nonprotein solids was 40.5 mg/g (range 33.4–60.2) of which ascorbic acid formed the remarkably high proportion of 2.1% (range 1.1–3.4).

In a number of near-term babies (37–42 weeks' gestational age) who died within 4 weeks of birth (conceptual age at death 37–46 weeks) forebrain ascorbic acid levels remained high, at least 3 times those of adults (Table I). The difference was also detectable in the cerebellum, though less conspicuous, but was only barely significant in brainstem. When a comparison is made with fetal plasma (Table II) it seems that the brain/plasma gradient in the term baby lies approximately between 12 (brainstem) and 25 (forebrain and cerebellum).

Neither plasma nor cell levels of ascorbic acid were altered during labour (Table II). Cord blood levels were much higher than those of maternal blood, the difference being more marked in plasma than in cells. Plasma urea levels did not differ between mother and fetus (Table II), in agreement with the finding that urea is not actively transported across the placenta (Battaglia et al., 1968).

Maternal plasma ascorbic acid in pregnancy was not significantly correlated with gestational age in the limited number of women examined. Women who were not pregnant had plasma levels

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**TABLE I**

Ascorbic acid concentrations in perinatal and adult brain

<table>
<thead>
<tr>
<th>Perinatal deaths (37–46 wk conceptual age)</th>
<th>Forebrain</th>
<th>Cerebellum</th>
<th>Brainstem</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adult (17–83 yr)</td>
<td>0.414</td>
<td>0.385</td>
<td>0.201</td>
</tr>
<tr>
<td></td>
<td>±0.136 (5)</td>
<td>±0.083 (6)</td>
<td>±0.040 (5)</td>
</tr>
<tr>
<td></td>
<td>0.127</td>
<td>0.209</td>
<td>0.153</td>
</tr>
<tr>
<td></td>
<td>±0.018 (6)</td>
<td>±0.041 (6)</td>
<td>±0.029 (6)</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>Ratio</td>
<td>3.25</td>
<td>1.84</td>
<td>1.31</td>
</tr>
</tbody>
</table>

Note: Each value (mg ascorbic acid/g wet weight) is the mean ± SD of the number of brains indicated in parentheses. Differences between perinatal and adult values were compared using Student's 't' test.

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**TABLE II**

Ascorbic acid and urea concentrations in maternal and cord blood at term

<table>
<thead>
<tr>
<th></th>
<th>Ascorbic acid (mg/100 ml)</th>
<th>Urea (mg/100 ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Plasma</td>
<td>Cells</td>
</tr>
<tr>
<td>Maternal blood</td>
<td>0.45</td>
<td>0.36</td>
</tr>
<tr>
<td>(onset of labour)</td>
<td>±0.27*</td>
<td>±0.15*</td>
</tr>
<tr>
<td>Maternal blood</td>
<td>0.47</td>
<td>0.33</td>
</tr>
<tr>
<td>(labour established)</td>
<td>±0.38*</td>
<td>±0.14*</td>
</tr>
<tr>
<td>Cord blood</td>
<td>1.65</td>
<td>0.85</td>
</tr>
<tr>
<td></td>
<td>±1.07</td>
<td>±0.39</td>
</tr>
<tr>
<td>Fetal/maternal  ratio†</td>
<td>4.4</td>
<td>2.8</td>
</tr>
<tr>
<td></td>
<td>±1.9</td>
<td>±1.7</td>
</tr>
</tbody>
</table>

Note: Each value represents the mean ± SD of blood samples taken during 16 deliveries.
*Significantly different (P <0.001) from cord blood value according to Student's 't' test.
†Calculated using the mean maternal value for each delivery.
significant higher than those of women in the second and third trimesters (Table III) and also

<table>
<thead>
<tr>
<th>Group</th>
<th>No.</th>
<th>Plasma ascorbic acid (mg/100 ml)</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Not pregnant</td>
<td>13</td>
<td>0.78 ± 0.36</td>
<td>0.26–1.28</td>
</tr>
<tr>
<td>1st trimester</td>
<td>8</td>
<td>0.66 ± 0.42</td>
<td>0.22–1.54</td>
</tr>
<tr>
<td>2nd trimester</td>
<td>11</td>
<td>0.50 ± 0.27*</td>
<td>0.07–0.82</td>
</tr>
<tr>
<td>3rd trimester</td>
<td>14</td>
<td>0.51 ± 0.33*</td>
<td>0.22–1.09</td>
</tr>
</tbody>
</table>

*Significantly different (P < 0.05) from the group that was not pregnant according to Student's 't' test.

significantly (P < 0.02) greater than those of women at the onset of labour (Table II).

Discussion

This study has shown that in an unselected group of fetal and neonatal brains (1) ascorbic acid is a constituent of major quantitative importance, (2) levels of ascorbic acid are much higher than in adult brains, and (3) these levels fall with increasing length of gestation. For the reasons outlined in Methods, it seems unlikely that these general trends were affected by variations in vitamin C intake, though such variations may have contributed to the relatively larger scatter in forebrain concentrations at a given gestational age.

In large nutritional surveys it has been found that on a constant intake of vitamin C plasma concentrations may fall with increasing length of pregnancy (Teel et al., 1938; Martin et al., 1957). In this limited study, levels throughout pregnancy were generally lower than those of women who were not pregnant. Briggs and Briggs (1972) found that pregnancy was not associated with a fall in leucocyte vitamin C. However, women receiving oral contraceptives containing oestrogen had low ascorbic acid levels in both leucocytes and plasma (Briggs and Briggs, 1973). It has been suggested (Clemetson, 1968) that oestrogens increase ascorbic acid breakdown. Such an effect may be mediated, in blood at least, through oxidation of ascorbic acid by caeruloplasmin, levels of which rise during pregnancy and in women on oral contraceptives (Tovey and Lathe, 1968). It has been reported that the redox state of plasma ascorbic acid varies during the menstrual cycle (Kofod et al., 1965). In mid-cycle, when oestrogen secretion is at its peak, ascorbic acid is present in a high proportion as its oxidized form. In summary, oestrogens may increase ascorbic acid oxidation and catabolism causing a rise in minimum requirement in pregnancy in addition to the demands of the growing fetus.

The minimum daily requirement for vitamin C is a matter of continuing controversy (Krebs, 1953; Goldsmith, 1961; Yew, 1973), though it is agreed that the requirement is likely to rise during pregnancy. A mean plasma ascorbic acid level in late pregnancy of 0.4 to 0.5 mg/100 ml (Tables II and III) probably indicates a daily intake of 40 to 60 mg (Martin et al., 1957; Goldsmith, 1961) at which tissues may not be completely saturated (Burch, 1961; Knox and Goswami, 1961). However, during vitamin C deficiency plasma levels may fall conspicuously before depletion of leucocytes (Lowry, 1952), so that a possible tissue deficit needs to be investigated more directly by examination of leucocyte levels in a similar group of pregnant women.

The present study confirms (Teel et al., 1938; Snelling and Jackson, 1939) that at term a marked transplacental gradient of ascorbic acid exists in favour of the fetus. The results suggest that this gradient is not influenced by labour. It is possible that the placenta may actively transport ascorbic acid by a mechanism similar to that which exists for amino acids (Dancis et al., 1968; Longo, Yuen, and Gusseck, 1973). Further, the brain may take up ascorbic acid from the blood by an energy-dependent process (Sharma, Johnstone, and Quastel, 1963). Whatever the mechanisms involved, it seems that the fetal brain at term concentrates ascorbic acid up to 100-fold from maternal plasma. This agrees with autoradiographical studies on the pregnant mouse (Hammarstrom, 1967), and perhaps implies an important function for ascorbic acid in brain development.

The function of ascorbic acid in the nervous system is largely unknown. Scurvy in guinea-pigs is associated with conspicuous histological changes in sympathetic and sensory ganglion neurons (Weatherford, 1961; Sulkin, Sulkin, and Nushan, 1973) as well as alterations in brain amino acids (Enwonwu, 1973), but it is not clear to what extent these changes are a direct result of loss of neuronal vitamin C. Two major roles for ascorbic acid in adult brain metabolism have been suggested. First, it may exert control over catecholamine metabolism (Izquierdo, Jofré, and Acevedo, 1968; Stone and Townsley, 1973), particularly as a cofactor of dopamine-β-hydroxylase (Kaufman and Friedman,
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1965). Secondly, ascorbic acid at physiological concentrations inhibits brain Na⁺-K⁺-activated ATPase (Inagaki, 1970; Boxall and Phizackerley, 1973; Frey, Pitts, and Askari, 1973) and hence may have a regulatory action on the sodium pump. However, neither of these hypothetical functions would be expected to be as important in the immature as in the mature brain.

A further possibility, for which there is some indirect evidence (Rinaldi, 1960; Edgar, 1970), is that ascorbic acid and its oxidized form are regulators of cell division. Forebrain neuroblast multiplication is thought to occur mainly in the period 10 to 18 weeks' gestational age (Dobbing and Sands, 1970, 1973) at a time when ascorbic acid concentrations were found to be high. The possible involvement of ascorbic acid in neuronal division should be examined.

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REFERENCES


The following articles will appear in future issues of this journal:

Review article: Onset of respiration at birth. M. J. Purves.

Brain maturation and damage in infants dying from chronic pulmonary insufficiency in postneonatal period. J. F. Smith, E. O. R. Reynolds, and A. Taghizadeh.


Hydrocephalus related to pulsion diverticulum of lateral ventricle. M. Mott and B. Cummins.

Thyroxine levels in normal newborn infants. R. H. Davies, K. Lawton, and D. Waring.