Control of fetal lung development in the rabbit

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Chiswick, M. L., Ahmed, A., Jack, P. M. B., and Milner, R. D. G. (1973). Control of fetal lung development in the rabbit. In a series of experiments, one rabbit fetus of a litter was decapitated in utero on day 24 of gestation and allowed to develop for a further 5 days. One effect of fetal decapitation was a reduction in the concentration of osmiophilic inclusion bodies in the type II pneumocytes of the lung. However, certain physical properties of the lung which depend on the presence of a surface active alveolar lining were normal. When 50 μg tetracosactrin was given to the fetus subcutaneously at the time of decapitation, there was no reduction in the concentration of inclusion bodies. It is suggested that though the production of surface active material in the pneumocyte is controlled at least in part by fetal adrenocortical hormones, the extrusion of this material into the alveolar space may be subject to other control. This may have important implications for the prophylactic treatment of the respiratory distress syndrome in premature babies by antepartum maternal glucocorticoid therapy.

The presence of a surface active lining which stabilizes alveoli and prevents their collapse on expiration is a prerequisite for fetal adaptation to extraterine existence. It is not until the onset of ventilation and the appearance of a gas/liquid interface that the lining is fully formed. The principle component of the lining is dipalmitoyl lecithin, and there is evidence that the osmiophilic inclusion bodies present in the type II pneumocytes represent surface active material that has been produced and stored in the cell (Buckingham et al., 1966; Kikkawa, Motoyama, and Cook, 1965). Pulmonary surface activity is first detected in late fetal life (Buckingham and Avery, 1962; Gluck et al., 1967). There is evidence that the respiratory distress syndrome of premature babies is associated with the absence of a normal surface active alveolar lining (Avery and Mead, 1959).

Liggins (1969) showed that when premature parturition was induced by the infusion of glucocorticoids to fetal lambs the animals survived, and when sacrificed later their lungs remained partially expanded. Stimulated by this finding, other workers showed that the administration of glucocorticoids to fetal rabbits (Kotas and Avery, 1971) and fetal lambs (DeLemos et al., 1970) accelerated the development of pulmonary surface properties. These findings culminated in a study which suggested that the administration of betamethasone to women for at least 24 hours before delivery might have lowered the incidence of the respiratory distress syndrome in those infants born before 32 weeks' gestation (Liggins and Howie, 1972). Knowledge of those factors which control the development of the surface active alveolar lining may put on a rational basis therapeutic effort to accelerate pulmonary maturation. We have studied the implication that endogenous fetal glucocorticoids contribute to normal development of pulmonary surface properties in the rabbit. Previous experiments have shown that fetal adrenal hypoplasia occurs in the absence of the pituitary gland (Jost and Picon, 1970). In a series of experiments the effect of fetal decapitation on the development of rabbit lung was studied.

Materials and methods

Dutch rabbits reared in the departmental animal house were used. The gestational age of a fetus was calculated from the time of artificial insemination of the doe. At laparotomy, under general anaesthesia, the uterus of a 24-day pregnant doe was exposed. One fetus of the litter was decapitated in utero as described by Beam (1968). On day 29 the doe was killed by a blow on the back of the head and the uterus was exposed. A ligature was firmly tied around the neck of each fetus through the uterine wall to prevent ventilation. The fetuses were delivered by hysterotomy and immediately killed by a blow on the back of the head. The

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Control fetus was compared with control littermates in terms of lung histology and certain pulmonary physical properties. In a further series of experiments, 50 µg tetracosactrin (Synacthen Depot, Ciba) was administered subcutaneously into the scapular region of the experimental fetus at the time of decapitation. After harvesting on day 29 as described above, the lungs were examined histologically.

**Histology.** Lung tissue was obtained from the decapitated fetus and a control littermate selected at random. Sections 1 µm thick were prepared (Gandy, Jacobson, and Gairdner, 1970) and stained with 0.5% p-phenylene diamine (British Drug Houses, Poole) in 70% alcohol (G. Gandy, personal communication, 1972). The number of type II pneumocytes and osmiophilic inclusion bodies per 100 µm² were counted using light microscopy at a magnification ×400 and an eyepiece grid. The average number of inclusion bodies per type II pneumocyte was calculated. Four sections were examined from each block, and four areas were counted on each section.

**Pressure-volume studies.** With the chest wall intact, the deflation limb of the pressure-volume curve was determined for the lungs of the decapitated fetus and 5 littermate controls nearest in headless body weight. The apparatus used was described by Avery, Frank, and Gribetz (1959). The pressure-volume curve was described as the volume of air (ml) retained per g lung weight at a distending pressure of 35 cm H₂O (V_max/g); the volume of air retained by the lungs on deflation to a pressure of 10 cm H₂O expressed as a percentage of V_max (% V_max at 10 cm H₂O).

**Bubble stability.** The method described by Pattle (1958) was followed. The mean stability ratio of bubbles squeezed from the lungs of the decapitated fetus and 5 littermates nearest in headless body weight was calculated. 20 to 30 bubbles from each pair of lungs were examined.

**Results**

Light microscopy showed the presence of well-formed alveoli in the lungs of both experimental and control fetuses, but there was considerable variation in alveolar size in different sections and in adjacent parts of a single section. The amount of osmiophilic inclusion bodies in the alveolar spaces of experimental and control fetuses appeared similar. Intra-alveolar inclusions were not counted as there was considerable variation in their concentration both within and between sections. The number of inclusion bodies per 100 µm² and inclusion bodies per type II pneumocyte was significantly less in the lungs of the decapitated fetuses compared with control fetuses (P < 0.001) (Table I and Fig.). However, the concentration of inclusion bodies per 100 µm² and inclusion bodies per type II pneumocyte in the fetuses that had received tetracosactrin at the time of decapitation was similar to that of control fetuses (Table II).

There was no significant difference between decapitated and control fetuses in terms of the parameters derived from the pressure-volume curves and the mean stability ratio of bubbles squeezed from the lungs (Table III).

**Discussion**

The reduction in concentration of inclusion bodies in the lungs of decapitated fetal rabbits, and the absence of this effect when tetracosactrin was administered at the time of decapitation suggests that fetal adrenocortical hormones and/or fetal adrenocorticotrophic hormone normally control the production of surfactant in the rabbit. Evidence that administration of 9-fluoro-prednisolone to fetal rabbits increases the concentration of pulmonary phosphoryl-choline-glyceride transferase, an enzyme necessary for the synthesis of dipalmityl lecithin, supports the concept of an adrenocortical hormonal influence on dipalmityl lecithin production (Farrell and Zachman, 1972).

**TABLE I**

<table>
<thead>
<tr>
<th></th>
<th>Type II pneumocytes (per 100µm²)</th>
<th>Inclusion bodies (per 100µm²)</th>
<th>Inclusion bodies (per type II pneumocyte)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Decapitated</td>
<td>74 ± 5</td>
<td>59 ± 7</td>
<td>0.8 ± 0.07</td>
</tr>
<tr>
<td>Control</td>
<td>70 ± 3</td>
<td>128 ± 18</td>
<td>1.8 ± 0.25*</td>
</tr>
<tr>
<td>Decapitated</td>
<td>65 ± 3</td>
<td>53 ± 5</td>
<td>0.8 ± 0.06</td>
</tr>
<tr>
<td>Control</td>
<td>74 ± 2</td>
<td>150 ± 10</td>
<td>2.0 ± 0.10*</td>
</tr>
<tr>
<td>Decapitated</td>
<td>87 ± 4</td>
<td>45 ± 3</td>
<td>0.5 ± 0.04</td>
</tr>
<tr>
<td>Control</td>
<td>81 ± 4</td>
<td>101 ± 7</td>
<td>1.3 ± 0.22*</td>
</tr>
<tr>
<td>Decapitated</td>
<td>92 ± 6</td>
<td>28 ± 3</td>
<td>0.3 ± 0.04</td>
</tr>
<tr>
<td>Control</td>
<td>90 ± 7</td>
<td>63 ± 6</td>
<td>0.7 ± 0.04*</td>
</tr>
</tbody>
</table>

Mean (± SE) of 16 observations in each lung is shown.  
*P < 0.001.
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(a) Lung from rabbit fetus showing osmiophilic inclusion bodies (arrows) within type II pneumocyte. (b) Decapitated fetus showing reduced numbers of inclusion bodies compared with both (b) control and (c) decapitated fetus treated with tetracosactrin. (× 500.)
TABLE II
Quantitative estimation of type II pneumocytes and inclusion bodies in four decapitation plus tetracosactrin experiments

<table>
<thead>
<tr>
<th>Decapitated + tetracosactrin</th>
<th>Inclusion bodies (per 100μm²)</th>
<th>Inclusion bodies (per type II pneumocyte)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Decapitated + tetracosactrin</td>
<td>86±3</td>
<td>112±10</td>
</tr>
<tr>
<td>Control</td>
<td>82±4</td>
<td>123±17</td>
</tr>
<tr>
<td>Decapitated + tetracosactrin</td>
<td>62±3</td>
<td>124±15</td>
</tr>
<tr>
<td>Control</td>
<td>69±2</td>
<td>166±10</td>
</tr>
<tr>
<td>Decapitated + tetracosactrin</td>
<td>90±7</td>
<td>117±14</td>
</tr>
<tr>
<td>Control</td>
<td>94±9</td>
<td>85±6</td>
</tr>
<tr>
<td>Decapitated + tetracosactrin</td>
<td>82±4</td>
<td>115±8</td>
</tr>
<tr>
<td>Control</td>
<td>90±5</td>
<td>153±9</td>
</tr>
</tbody>
</table>

Mean (±SE) of 16 observations in each lung is shown.

*Not significant.

TABLE III
Physical properties (means±SE) of lungs from decapitated rabbit fetuses and littermate controls

<table>
<thead>
<tr>
<th></th>
<th>Decapitated (no. = 7)</th>
<th>Control (no. = 35)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vmax/g</td>
<td>3·1±0·3</td>
<td>2·8±0·2*</td>
</tr>
<tr>
<td>%Vmax at 10 cm H2O</td>
<td>71±2</td>
<td>72±1*</td>
</tr>
<tr>
<td>Bubble stability ratio</td>
<td>0·75±0·02</td>
<td>0·77±0·03*</td>
</tr>
</tbody>
</table>

* Not significant.

There is no conclusive evidence concerning the ability of maternal glucocorticoids to cross the placenta in the rabbit. There is good evidence that the sheep placenta is resistant to diffusion of cortisol (Beitins et al., 1970). That it is the fetus which controls pulmonary maturation is teleologically sound, because there is abundant evidence that, in some species at least, the fetal adrenal gland plays an important role in the initiation of labour (Cowie et al., 1964; Liggins and Kennedy, 1968). Naeye, Harcke, and Blanc (1971) showed that human anencephalic newborn infants with adrenal hypoplasia had less than half the mass of osmiophilic inclusion bodies in the type II pneumocytes compared with those who were not anencephalic.

Though this supports our experimental results, it is difficult to explain the findings of Naeye et al. (1971) on the basis of reduced plasma levels of adrenocortical hormones during fetal life. The human placenta is relatively permeable to corticosteroids (Migeon, Bertrand, and Gemzell, 1961), and indeed the plasma levels of 17-hydroxycorticoids in the cord blood of anencephalic infants have been shown to be normal (Nichols, Lescure, and Migeon, 1958).

The determination of the stability of bubbles squeezed from the lungs correlates well with other methods of evaluating pulmonary surface properties (Gandy et al., 1968). Pittle (1958) showed that the mean stability ratio of bubbles derived from the lungs of normal term fetal rabbits was 0·572. Humphreys and Strang (1967) found that the mean stability ratio of bubbles from the lungs of 29-day fetal rabbits was 0·6 to 0·9. Our results indicated the presence of surface active material in the lungs of both decapitated and control fetuses in amounts capable of forming a normal alveolar lining. These findings are supported by the normal pressure-volume results in both decapitated and control fetuses which are comparable with earlier reports in normal 29-day fetal rabbits (Humphreys and Strang, 1967; Kotas and Avery, 1971). Our results show that the relation of the osmiophilic inclusion body to the development of the surface active alveolar lining is not a simple one. A reduction in the concentration of inclusion bodies is consistent with the development of normal pulmonary surface properties; this suggests that a considerable reserve of surface active material is present in the lung parenchyma. At operation the neck was ligated before removal of the fetal head, and the communication between alveoli and the amniotic space was permanently lost. There is evidence that the phospholipids on the alveolar surface are rapidly metabolized (Tierney, Clements, and Trahan, 1967). Surface active material extruded into the alveolar space and remaining there is therefore unlikely to be responsible for the normal pulmonary surface properties of the decapitated fetuses. Furthermore, the intra-alveolar concentration of inclusion bodies in experimental and control fetuses appeared similar on light microscopy.

The presence of ample parenchymal lecithin and a normal concentration of intracellular osmiophilic inclusion bodies has been reported in hyaline membrane disease occurring in infants of diabetic mothers ( Boughton, Gandy, and Gairdner, 1970).
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This is consistent with the suggestion that the synthesis of lecithin, its extrusion into the alveolar space, and its subsequent metabolism may be controlled by different mechanisms.

Our results show that naturally occurring fetal adrenocortical hormones probably play a role in the development of pulmonary surface properties in the rabbit fetus, and support the concept that maternal glucocorticoid therapy may be beneficial in accelerating pulmonary maturation in the human fetus. However, it is probable that impairment of the mechanism by which surface active material is transported within the type II pneumocyte or discharged into the alveolus is as important as impaired synthesis of surfactant active material in certain types of respiratory distress syndrome of prematurity. This may prove a limiting factor in the role of maternal glucocorticoid therapy for the prevention of the respiratory distress syndrome in prematurely born offspring.

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REFERENCES


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