Assessment of mitochondrial toxicity in newborns and infants with congenital cytomegalovirus infection treated with valganciclovir

Alba Ortiz-Gracia,1,2 Maria Ríos,3 Ester Tobías,4,5,6,7 Antoni Noguerà-Julian,3,4,8,9 Francesc Josep García-García,4,5,6,7 Judith Cantó-Santos,4,5,6,7 Laura Valls-Roca,4,5,6,7 Glòria Garrabou,4,5,6,7 Josep Maria Grau,4,5,6,7 Francesc Cardellach,4,5,6,7 Emilia Sánchez,10 Constanza Morén,10 Clàudia Fortuny10

INTRODUCTION

Congenital cytomegalovirus (CMV)-infection is considered a rare disease (ORPHANET:294). It occurs via the placenta during pregnancy and represents the most common congenital infection in newborns.4 It is estimated that 1%–4% of pregnant women who are seronegative for CMV become infected during pregnancy and 30%–40% of these women will transmit the infection to their children.2,7 Although it is estimated that 90% of congenital CMV-infected infants remain free of symptoms,4,5 this infection is the main cause of non-hereditary sensorineural hearing loss in infants and is also associated with severe neurodevelopmental disorders (cerebral palsy, mental retardation, seizures and impaired vision) and retinitis.1,3,6,8

Ganciclovir and valganciclovir are the main anti-CMV therapeutic options of choice. The first clinical trial on the administration of intravenous ganciclovir during 6 weeks in newborns with symptomatic congenital infection showed a slight improvement in hearing and neurodevelopmental outcomes at 6 and 12 months.9 In contrast to ganciclovir, the prodrug valganciclovir can be administered orally. A more recent clinical trial with valganciclovir showed better long-term neurocognitive and audiological outcomes with a 6-month compared with a 6-week regimen.10 These results support the current indication to treat newborns diagnosed with symptomatic congenital CMV infection with a 6-month regimen of oral valganciclovir. To date, the most common side effect of ganciclovir/valganciclovir treatment is neutropenia.1,3,6,11

Ganciclovir/valganciclovir are guanosine analogues and, thus, their mechanism of action is similar to that of the antiretrovirals: nucleoside reverse transcriptase inhibitors (NRTI). Ganciclovir/valganciclovir and NRTI act as substrates and inhibitors of viral DNA polymerases required for viral replication. Mitochondrial DNA (mtDNA) polymerase gamma in human cells is responsible for the synthesis of the mitochondrial genome. Thus, secondary inhibition of this enzyme may cause point mutations, deletions and depletion (loss of an entire copy of the mitochondrial genome) in the host cell.4

ABSTRACT

Background Ganciclovir/valganciclovir is currently indicated during the first 6 months of life in symptomatic children with congenital cytomegalovirus (CMV) infection. However, this treatment may have the potential to induce mitochondrial toxicity due to off-target inhibition of DNA-polymerases. Similar anti-HIV drugs have been associated with mitochondrial toxicity but this has never been explored in CMV.

Objective To determine the potential mitochondrial toxicity profile at the genetic, functional and biogenesis level in peripheral blood mononuclear cells from a cohort of newborns and infants with symptomatic congenital CMV infection (treated with valganciclovir, untreated and uninfected controls).

Design Longitudinal, observational and controlled study.

Setting and patients Subjects were recruited at the tertiary referral Hospital Sant Joan de Déu and experiments were conducted at IDIBAPS-Hospital Clinic of Barcelona, Spain. CMV-infected newborns underwent comprehensive monthly clinical follow-up.

Methods Mitochondrial parameters, audiometry and neurological assessment were measured at baseline, 3–6 and 12 months after inclusion in the study. The Kruskal-Wallis test for k-independent samples and Friedman tests for repeated measurements were applied.

Results Complex IV, citrate synthase enzymatic activities and mtDNA remained preserved in congenital CMV-infected infants treated with valganclovir compared with controls (p>0.05 in all cases).

Conclusions No evidence of mitochondrial toxicity was found in infants treated with valganciclovir for congenital CMV.
the mtDNA molecules. These alterations in the mitochondrial genome, which codes for proteins of the mitochondrial respiratory chain (MRC), can lead to mitochondrial dysfunction, ultimately compromising cell function, tissue viability and even the organism exposed, as described in previous literature in the case of NRTIs in vivo and in vitro. In fact, the range of clinical signs and symptoms that have been associated with mitochondrial dysfunction, of either congenital or acquired origin, is extensive and highly heterogeneous, with hearing loss being among the most relevant.

We hypothesise that human cellular DNA polymerases (nuclear and mtDNA polymerases, including mtDNA polymerase gamma) may be inhibited by valganciclovir treatment, as previously reported in the case of other antiviral treatments with a similar mechanism of action, leading to mtDNA depletion and subsequent enzymatic dysfunction. To our knowledge, this is the first study on potential mitochondrial toxicity derived from the administration of valganciclovir.

**METHODS**

**Study design**

This was a controlled, observational, longitudinal, prospective study conducted in the Hospital Sant Joan de Déu, Barcelona from January 2018 to March 2021.

**Patients**

Parents or legal guardians provided informed consent. Sample recruitment was also approved at each centre and was performed with the collaboration of the Infectious Diseases Departments of both tertiary care hospitals: Hospital Sant Joan de Déu (Barcelona, Spain) and Hospital Clinic of Barcelona (Barcelona, Spain).

**Inclusion/exclusion criteria and groups of study**

This study included 34 CMV-infected children and 8 healthy infants classified as the control group. The exclusion criteria were a personal or family history of mitochondrial or neuromuscular disease or contact with drugs with potential toxicity for mitochondria (eg, aminoglycosides, linezolid or antipsychotics).

Symptomatic CMV-infected infants were treated with valganciclovir (n = 16) (32 mg/kg/day, two times a day) for 6 months according to their presentation of clinical, neuroimaging and/or laboratory disorders related to CMV-infection.

CMV-infected infants underwent repeated measurements of clinical and mitochondrial parameters at 3, 6 and 12 months of age. The CMV-infected infants were divided into different groups depending on age: (i) GROUP 0: (age, 0–2 months; n=15), (ii) GROUP 3: (age, 2–4 months; n=20), (iii) GROUP 6: (age, 5–8 months; n=18) and (iv) GROUP 12: (age 9–16 months; n=24). Eighteen infants were longitudinally followed at the different time points. The control group was made up of eight healthy infants. Clinical and epidemiological data of both patients and controls are shown in table 1.

**Laboratory assays for mitochondrial function assessment**

Experimental analyses of mitochondrial parameters were performed in the Mitochondrial Laboratory at the Cellex-Institut d’Investigacions Biomèdiques August Pi i Sunyer (IDIBAPS), at the Hospital Clinic of Barcelona, Spain.

**Sample collection**

Peripheral blood mononuclear cells (PBMC), lymphocytes and monocytes were obtained from 1 to 3 mL of venous blood. PBMCs were isolated by centrifugation, using the Ficoll gradient (Histopaque 1077, Sigma Diagnostics, St Louis, Missouri, USA). Final pellets were collected and cryopreserved at −80°C for further experimental procedures.

**Protein quantification**

The bicinchoninic acid assay was used to calculate the total protein cell content following the manufacturer’s instructions (PierceBCA-Protein Assay Kit #23225; Thermo-Scientific).

**mtDNA quantification**

A mitochondrial-DNA (mtDNA) depletion study was performed as described elsewhere.

**Mitochondrial function**

Spectrophotometry was performed to assess complex IV (CIV)/cytochrome c oxidase (COX) enzyme activity and mitochondrial content by measuring citrate synthase (CS) enzyme activity. MRC enzyme activities were expressed as relative units, normalised for mitochondrial content, estimated by CS activity (COX/CS). A sample of pork muscle homogenate with known values of normality was assessed as an internal quality control of the technique. Experimental conditions for CIV and CS enzymatic activities are provided (online supplemental material).

**Statistical analysis**

Kolmogorov–Smirnov tests were used to assess normality of the variables. Statistical analysis was performed using the SPSS v27. The non-parametric Kruskal Wallis test for k-independent samples was used for cross-sectional comparisons between groups. Samples obtained along the follow-up period were longitudinally compared (n = 18) using the nonparametric Friedman test for repeated measurements. Statistical significance was set at p=0.05.

**RESULTS**

MtDNA content remained conserved among the different groups (figure 1). The sample size and the epidemiological and clinical data of the subjects included in the study are summarised in table 1. The patients included in the study were classified according to different age groups.

Out of 34 infants, 29 were diagnosed with CMV-infection at birth or prenatally and 5 were diagnosed retrospectively.
Drug therapy

Symptoms and signs of congenital infection were identified in 19 infants: severe neurological affection (n=2), only sensorineural hearing loss (n=6), chorioretinitis (n=1), cholestatic hepatitis (n=1) and microcephaly, laboratory abnormalities (thrombocytopenia, CSF alterations or elevated ALT) and neuroimaging findings (n=9).

Sixteen infants (14 newborns) initiated treatment with valganciclovir based on the clinical disorders described. The remaining three symptomatic infants were not treated because they were diagnosed after 6 months of age. In three infants, the treatment was interrupted after 2–6 weeks due to haematological toxicity.

Quantitative data of all measured mitochondrial parameters are shown (table 2).

Mitochondrial enzymatic activities of CIV (figure 2) and CS (figure 3) did not show differences between groups.

MtDNA content (figure 4A) and CIV enzymatic activity (figure 4B) relative values to CS are shown.

Since mitochondrial genome quantification did not differ over time in the different groups (figure 5), alterations at a functional level were not expected to be different. Correlations between genetic, functional and biogenic parameters were analysed, with no statistically significant relationship between any of the parameters.

DISCUSSION

In this study, we explored the potential mitochondrial toxicity related to CMV-infection and 6 months of anti-CMV treatment with valganciclovir in infants at different timepoints during

![Figure 1](image-url)
viral DNA polymerases to avoid replication. NRTI-which present analogous mechanisms of action by blocking the mtDNA depletion and mitochondrial defects in the functional is generalised to the mitochondrial and functional levels. Thus, chondrial damage is not restricted to a specific level, but rather exposure. Since the mechanism of action of valganciclovir treatments, such as anti-

Depletion and mitochondrial dysfunction by other antiviral based on a previous wide body of evidence reporting mtDNA bition of the mtDNA polymerase gamma. This assumption is carried out at different levels including genetic, functional and biogenic processes to further decipher potential interac-

tions and/or compensatory mechanisms. The rationale of the primary hypothesis of this study is that anti-

CMV treatment is similar to that of NRTI, it is conceivable that mitochondrial toxicity derived from valganciclovir could be present either at a genetic or functional level in newborns. While our study explored potential valganciclovir-mitochondrial interferences, mitochondrial safety was confirmed among the participants along the different time periods of age and follow-up timepoints.

The consequence of mitochondrial dysfunction is an imbalance in oxidation-reduction that leads to an accumulation of pyruvate, lactic acid, ketone bodies and alanine, producing heterogeneous spectrum of symptoms in the whole body. The relevance of investigations looking for mitochondrial toxicity in infants treated for congenital CMV is important as the clinical manifestations of potential toxicity are not easily identifiable in symptomatic patients with hearing loss or neurological alterations. In addition, there is a growing trend to screen for CMV

Figure 2 Mitochondrial cytochrome c oxidase or complex IV enzymatic activity (nmol/min mg protein) suggestive of mitochondrial function at the level of cell respiration, estimated by nanomoles of consumed reduced cytochrome c per minute and milligram of protein. Mitochondrial enzymatic activity of complex IV of the mitochondrial respiratory chain remained unaltered among the different treatment and age groups. Black dots show untreated groups whereas clear dots show valganciclovir treated groups. From left to right, the groups are divided depending on the age-dependent time period. The grey area indicates the cytochrome c oxidase reference values in the control group. G0, group 0 including newborns from 0 to 2 months; G3, group 3 including newborns from 2 to 4 months; G6, group 6 including newborns from 5 to 8 months; G12, group 12 including newborns from 9 to 16 months; UT, untreated; VAL, valganciclovir. Figure created by CM coauthor.

Figure 3 Citrate synthase enzymatic activity (nmol/min mg protein) suggestive of mitochondrial mass, estimated by nanomoles of product per minute and milligram of protein. Mitochondrial mass was preserved among the different treatments and between age difference. Black dots show untreated groups whereas clear dots show valganciclovir treated groups. From left to right, the groups are divided depending on the age-dependent time period. The grey area indicates citrate synthase enzymatic reference values in the control group. G0, group 0 including newborns from 0 to 2 months; G3, group 3 including newborns from 2 to 4 months; G6, group 6 including newborns from 5 to 8 months; G12, group 12 including newborns from 9 to 16 months; UT, untreated; VAL, valganciclovir. Figure created by CM coauthor.

Figure 4 Mitochondrial DNA and cytochrome c oxidase relative values normalised by mitochondrial mass (citrate synthase). (A) The mitochondrial DNA normalised by mitochondrial mass remained unaltered among the different study groups (either treatment or time period). (B) The relative values of enzymatic activity of cytochrome c oxidase or complex IV remained unaltered among the different study groups (either considering the treatment or time period). Black dots show untreated groups whereas clear dots show valganciclovir treated groups. From left to right the groups are divided depending on the age-dependent time period. The grey area indicates mitochondrial DNA and enzymatic reference values, relative to mitochondrial mass, in the control group. G0, group 0 including newborns from 0 to 2 months; G3, group 3 including newborns from 2 to 4 months; G6, group 6 including newborns from 5 to 8 months; G12, group 12 including newborns from 9 to 16 months; UT, untreated; VAL, valganciclovir. Figure created by CM co-author.
DNA in urine samples from newborns. The number of CMV-infected children receiving ganciclovir/valganciclovir treatment might be expected to grow substantially in the following years. Moreover, the first months of life are a key stage in the development of the infants and any health issue at this point may have an impact on their future lives. We cannot rule out that the lack of differences may be due to the limited sample size or the source of the sample. However, longitudinal assessment with repeated measurements is expected to increase statistical power, and PBMC represents the main target tissues of CMV-tropism.

In some patients with CMV infection, high doses of ganciclovir/valganciclovir may be indicated, representing greater exposure of the infant to the drug.4,24 Also, the treatment of infected infants requires exposure to these drugs over a longer period than that used in other indications.6 infected infants requires exposure to these drugs over a longer period than that used in other indications.6

Contributors AO-G and CM conducted all the experimental procedures. FC supervised the clinical and experimental data. GG, FG-G and JC-S contributed to enzymatic activities. CV-R, JMG and ET contributed to mtDNA analyses. CF is the principal investigator and obtained financial support. AO-G, MR, AN-I and CF gathered the clinical data and recruited patients and controls. ES, AO-G and CM conducted statistical analyses. CM supervised the laboratory experiments. AO-G and CM wrote the manuscript. All coauthors approved the final version to be published. CM and CF authors share main last authorship as principal investigators. CM is the guarantor.

Funding This study received funding from the Fundación Privada Cellex: (CP042187) (Fundación Clinic per la Recerca Biomédica) and Instituto de Salud Carlos III (ISCIII, Spain): (PI17/00359 and PI13/01738) cofunded by European Regional Development Fund ‘A way to make Europe’. Antoni Noguera-Julian was supported by ‘Subvencions per a la Intensificació de Facultats d’Expertos’ (Departament de Salut de la Generalitat de Catalunya, Programa PERIS 2016-2020) (STI008/18/00193). Constanza Morén was supported by a postdoctoral CIBERER contract.

Competing interests None declared.

Patient consent for publication Not applicable.

Ethics approval This study involves human participants and was approved by Ethics Committee of Hospital Sant Joan de Déu, Barcelona, Spain (CEIN NUMBER: PIC-145-17 (PI17/00359). Date of approval: 23/11/2017. Participants gave informed consent to participate in the study before taking part.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement All data relevant to the study are included in the article or uploaded as supplementary information.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BML) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BML. BML disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BML does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.