Abnormalities of vascular prostaglandins in Henoch-Schönlein purpura

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SUMMARY The ability of plasma to support prostacyclin like activity from human umbilical arterial rings was studied in 17 patients with Henoch-Schönlein purpura and 17 controls matched for age and sex. Plasma from 13 of the 17 patients showed a diminished or absent ability to support prostacyclin like activity in vitro. Six patients whose plasma had a low or absent ability to support prostacyclin like activity showed evidence of inhibitory activity. Plasma from three of these patients also failed to preserve the effect of a stable prostacyclin like analogue (ZK36-374). The plasma concentration of prostacyclin metabolite and the serum concentration of thromboxane A₂ metabolite, thromboxane B₂, were measured simultaneously. The concentration of plasma prostacyclin metabolite in 10 of the 14 patients was decreased, and a positive correlation was found between the plasma prostacyclin metabolite values and the ability of the plasma to support prostacyclin like activity. There was no significant difference in the serum thromboxane A₂ metabolite concentrations between the patients and controls. These data suggest that abnormalities of vascular prostaglandin metabolism are involved in the pathophysiology of Henoch-Schönlein purpura.

Key for abbreviations used in text

PGI₂: prostacyclin.

PSA: estimation of ability of plasma to support vascular PGI₂ like activity.

PSAI: inhibition of vascular PGI₂ like activity.

PGI₂m: prostacyclin metabolite.

TxB₂: thromboxane A₂ metabolite.

Henoch-Schönlein purpura is the most commonly encountered type of vasculitis in childhood. The skin lesions are the most obvious sign, but visceral involvement carries a more serious prognosis. In two thirds of patients arthritis and gastrointestinal involvement occur, and in a minority of patients, in addition, the central nervous system is involved. Renal involvement, however, is potentially the most serious manifestation as chronic renal impairment may develop.^{1 2} The aetiology and pathogenesis are obscure, but the primary manifestations are due to inflammation of small non-muscular vessels.³ Henoch-Schönlein purpura may follow exposure to drugs or allergens⁴ and is in many cases preceded by upper respiratory tract infection occasionally caused by group A β -haemolytic streptococci.³ None of these factors, however, have been confirmed to be of pathogenetic importance.

Increased number of data are now available about the possible role of vascular prostaglandins prostacyclin (PGI₂), and thromboxane (TxA₂) in the pathogenesis of other vasculopathies. PGI₂ is an unstable substance with potent antiaggregatory and vasodilatory activity,⁵ synthesised from arachidonic acid by the vessel wall cyclooxygenase system.⁶ TxA₂ is also an unstable endoperoxide with vasoconstricting and platelet aggregatory activity,⁷ derived from arachidonic acid predominantly by the platelet cyclooxygenase system.⁸ Remuzzi *et al* first showed deficient PGI₂ production in the haemolytic uraemic syndrome and the related disorder of thrombotic thrombocytopenic purpura. Plasma taken from these patients had a low capacity to support PGI₂ production in vitro from rat aortic rings.⁹ Reduced umbilical and placental vascular PGI₂ has been observed in patients with severe pre-eclampsia.¹⁰ Saldeen *et al* have reported an increased concentration of TxA₂ and PGI₂ metabolites in patients with deep venous thrombosis,¹¹ and increased TxA₂ synthesis by platelets has been shown in Kawasaki disease.¹² We report our preliminary findings on various aspects of vascular prostaglandin metabolism in the acute phase of Henoch-Schönlein purpura.

Patients

Seventeen patients with Henoch-Schönlein purpura aged between 2 and 13 years (mean (SD) 5.7 (2.5) years) and 17 controls matched for age and sex were studied. The control children were free of renal, cardiovascular, pulmonary, and inflammatory disease and were admitted for minor surgical operations. All patients had the classical purpuric rash, and in 13 cases the skin lesions were associated with joint manifestations. One child had recurrent purpura, and none of the patients had hypertension. In eight cases gastrointestinal symptoms were observed, and this included one patient who required operative intervention for ileocolic intussusception. Three patients had a mild illness without abnormal urinalysis or abdominal or joint manifestations. In eight patients haematuria (>10 red blood cells/ μ l) and/or proteinuria (31 (SD 19) mg/hr/ m^2), was observed. There was no significant difference in the creatinine clearance between those patients with abnormal urinalysis (94 (SD 33) ml/min/ $1.73 m^2$) and those with normal urinalysis (100 (SD 21) ml/min/ $1.73 m^2$). In all patients plasma albumin concentrations, plasma electrolyte concentrations, and blood film showed no abnormalities, and the mean platelet count was $357 (SD 54) \times 10^9/l$.

Methods

After informed consent had been obtained 10 ml of venous blood was taken from the controls and from the patients during the acute phase of the disease, 1–5 days after the occurrence of the rash. Blood for estimation of the ability of the plasma to support PGI₂ like activity (PSA) was anticoagulated with 3.2% trisodium citrate in a ratio of 9:1. Samples for PGI₂ metabolite (PGI₂m) measurements were collected into ice cold plastic tubes anticoagulated with 1:9 volume 3.8% w/v trisodium citrate: $3 \times 10^{-5} M$ indomethacin: $10^{-4} M$ adenosine. Tubes were kept on ice and spun within one hour at $4^\circ C$ and 2500 g. Plasma was separated and stored at $-70^\circ C$. Samples of this pool were included in each assay both at the beginning and end to determine reproducibility and to act as a standard. A known amount of 6-keto PGF₁ α was added to an aliquot of this pooled plasma to estimate recovery.

Platelet poor plasma, obtained for PSA estimation, was prepared within 20 minutes of collection by centrifugation at 2000 g for 10 minutes at $4^\circ C$. Simultaneously, serum samples were separated for estimation of TxB₂ (stable metabolite of TxA₂). All plasma and serum samples were stored at $-70^\circ C$ until examination.

Platelet rich plasma for aggregation studies was

prepared from normal adults by centrifugation of citrated plasma at 800 g for 10 minutes at room temperature. The final platelet count was adjusted to $250\text{--}300 \times 10^9/l$ by dilution with autologous platelet poor plasma. No patient or control was taking any drug known to alter prostaglandin metabolism for at least two weeks before the study.

Estimation of ability of plasma to support vascular PGI₂ like activity (PSA). The ability of test plasma (patient and control) to support PGI₂ like activity was assessed by measurement of platelet anti-aggregatory activity by the method of Moncada *et al.*¹³ Human umbilical arterial rings were obtained from freshly delivered umbilical cords. The umbilical arteries were freed from all surrounding tissue, cut into rings 1 mm in length (30–50 mg wet weight) and kept in Ringer's buffer (pH 7.4) at $0^\circ C$ for not more than 60 minutes. The rings were then incubated in 1 ml tromethamine buffer (pH 8.6) for five minutes at $37^\circ C$, and PGI₂ like activity (platelet antiaggregatory activity) of 100 μ l of the supernatant was tested. This was added to 200 μ l platelet rich plasma in a Servogor 120 dual channel aggregometer. The mixture was incubated for one minute at $37^\circ C$, then collagen (Hormonochemie), at a final concentration of 2 μ g/ml, was added. The rate of aggregation was recorded using a Malin platelet aggregation recorder.¹⁴ The rings were then washed several times with Ringer's buffer at $37^\circ C$ until no antiaggregatory activity could be detected. These exhausted rings were then incubated with 1 ml of test platelet poor plasma at $37^\circ C$ for 20 minutes. PGI₂ like activity was then assessed as before. The ability of each test platelet poor plasma to support PGI₂ like activity was expressed by calculation of the percentage difference in inhibition of platelet aggregation obtained by the same exhausted ring before and after the addition of test platelet poor plasma.

Detection of inhibition against PGI₂ like activity (PSAI). To detect inhibitory activity in the test plasma a modified method of Levin *et al* was used.¹⁵ Fresh, unexhausted umbilical arterial rings were incubated in phosphate buffered saline (pH 7.4) at $37^\circ C$ for five minutes. The PGI₂ like activity of the supernatant was compared with that produced by the same ring after five minutes incubation with test platelet poor plasma.

Preservation of PGI₂ like effect of stable PGI₂ analogue. The preservation of PGI₂ like effect of a stable PGI₂ analogue, ZK36-374 (Schering chemicals AG), was assessed using test platelet poor plasma. ZK36-374, at a final concentration of 100 ng/l, was

incubated with 300 µl of test platelet poor plasma at 37°C for five minutes. The concentration of ZK36-374 corresponded to that concentration known to have a definite antiplatelet effect when the drug is given by intravenous infusion.¹⁶ Then 100 µl of the supernatant was removed and added to 200 µl platelet rich plasma. After one minute of incubation and stirring, platelet aggregation studies using collagen were carried out as before. After incubation of normal plasma with ZK36-374 total inhibition of platelet aggregation should be expected due to the protective effect of the PGI₂ analogue.¹⁶

Radioimmunoassay studies of plasma PGI₂m and serum TxB₂. Plasma concentrations of PGI₂m were assessed by radioimmunoassay using a modification of the method of Mitchell.¹⁷ As only 40% of PGI₂ is metabolised to 6-keto PGF₁α we used an assay that measures in addition the more labile metabolites.¹⁸ The measurement was carried out on unextracted plasma buffered with 0.1% bovine albumin, as many extraction techniques may add further impurities.¹⁹ Our method gives good reproducibility and recovery of added PGI₂m and has a sensitivity of 5 pg/ml.²⁰ Serum TxB₂ was measured using a radioimmunoassay method as described by Granstrom and Kindahl.²¹

All platelet aggregation and radioimmunoassay studies were carried out in duplicate. Statistical analysis was carried out using the Student's *t* test and Spearman rank correlation test.

Results

Ability of plasma to support PGI₂ like activity was reduced in 13 of the 17 patients with Henoch-Schönlein purpura studied. In five cases PSA was undetectable (Fig. 1). The difference between the results of the patient group (21.3 (SD 20.5)%) and the control group (57.2 (SD 12.2)%) was significant (*p*<0.001). In addition, the results of those patients with abnormal urinalysis and gastrointestinal involvement (7.5 (SD 18.1)%) was significantly different from the results of those patients with neither of these manifestations, (32.3 (SD 23.5)%) (*p*<0.01).

Detection of inhibition against PGI₂ like activity (PSAI). Plasma from six patients in which the PSA was low or absent was further investigated to detect PSAI. Less PGI₂ like activity was produced by the unexhausted umbilical arterial rings when incubated with platelet poor plasma from the six patients than with either phosphate buffered saline or control platelet poor plasma (Table).

Fig. 1 Ability of plasma to support vascular PGI₂ like activity (PSA) in patients with Henoch-Schönlein purpura.

Table Plasma factors influencing prostacyclin (PGI₂) like activity in Henoch-Schönlein purpura

Case No	PSA* (%)	PSAI†	Preservation of ZK 36-374‡ effect
<i>Patients</i>			
1	0	+	Normal
2	0	+	Decreased
3	0	+	Decreased
4	0	+	Decreased
5	0	+	Normal
6	23	+	Normal
<i>Controls</i>			
1	62	-	Normal
2	57	-	Normal
3	65	-	Normal

*Estimation of ability of plasma to support vascular PGI₂ like activity.
†Inhibition of vascular PGI₂ like activity: + = present, - = absent.
‡A stable PGI₂ like analogue.

Preservation of PGI₂ like effect of stable PGI₂ analogue. The effect of ZK36-374 was noticeably depressed after incubation with platelet poor plasma from three of the six patients in whose plasma PSAI was present (Table).

Plasma PGI₂m and serum TxB₂. There was a significant difference between the results of plasma PGI₂m concentrations obtained from 14 patients with Henoch-Schönlein purpura (16.6 (SD 8.7) pg/ml) and the control group (35.3 (SD 7.5) pg/ml)

Fig. 2 Plasma PGI₂ metabolite (PGI₂m) concentrations in patients with Henoch-Schönlein purpura.

($p < 0.05$) (Fig. 2). A positive correlation was found between PSA and PGI₂m values ($r = 0.52$, $p < 0.05$). There was no significant difference in serum TxB₂ concentration between the 16 patients (11.4 (SD 9.4) ng/ml) and the control group (13.2 (SD 8.6) ng/ml). No significant correlation was found between the results of the tests on serum TxB₂ and plasma PGI₂m concentrations.

Discussion

It is well known that the pathological defect in Henoch-Schönlein purpura is that of a predominantly small vessel vasculitis. Abnormalities of vascular prostaglandin metabolism have been documented in other vasculopathies, particularly in the haemolytic uraemic syndrome. There are, however, no published reports of studies in Henoch-Schönlein purpura. Conceivably, after an endothelial insult, such as deposition of immune complexes, a defect of PGI₂ generation may be present.

We have shown that plasma from patients with Henoch-Schönlein purpura in the acute phase of the illness has a diminished or absent ability to support PGI₂ generation in vitro from human umbilical arterial rings. Absent or depressed PSA concentra-

tions may theoretically be due to a number of causes, occurring singly or in combination. Firstly, a decreased ability of the plasma to stimulate PGI₂ production;⁹ secondly, the presence of inhibitory activity in the plasma either to the production¹⁵ or to the effect of PGI₂,²² and thirdly, rapid degradation of stimulated PGI₂.²³ All six patients in this study who had very low or absent concentrations of PSA, in whom further investigation was carried out, showed the presence of PSAI. The presence of PSAI may mean inhibitory activity against PGI₂ production, inhibitory activity against PGI₂ effect, or increased PGI₂ degradation. To attempt to differentiate these possibilities we looked at the ability of test plasma to preserve the effect of a stable PGI₂ analogue (ZK36-374) and found this to be absent in three of the six patients studied. The concentration of ZK36-374, although significantly higher than the physiological concentrations of PGI₂m in both patients and controls, was chosen because a definite antiplatelet effect was desired. It would be interesting to undertake further studies using different concentrations of the PGI₂ analogue; however, this was outwith the scope of this present study. On the basis of these data it would seem that in patients with low or absent concentrations of PSA inhibitory activity may exist to either PGI₂ production or PGI₂ effect.

We have also shown that most patients with Henoch-Schönlein purpura in this study had a decreased concentration of plasma PGI₂ metabolite (PGI₂m). It is of interest, however, that two patients showed a fairly high concentration of PGI₂m in combination with a low or absent PSA concentration and plasma inhibitory activity to the effect of the stable PGI₂ analogue. This would tend to confirm the presence of a circulating inhibitor to PGI₂ effect in these cases. These findings also show that platelet aggregation studies are more sensitive in detecting decreased PGI₂ like activity of plasma than determination of PGI₂m concentration.

We have found no significant difference in serum thromboxane (TxB₂) concentrations between the patient group and the control population. Furthermore, there was a poor correlation between the concentration of serum TxB₂ and plasma PGI₂m concentrations. These results do not suggest the presence of platelet activation in this syndrome. In view of the small numbers of patients in this study and the absence of any follow up data we felt unable to attempt a correlation between the demonstrated abnormalities and the clinical picture. It is of interest, however, that patients whose clinical picture included abnormal urinalysis and gastrointestinal involvement had a mean PSA concentration that was significantly depressed compared with

those patients in whom these clinical manifestations were absent.

The above findings suggest the abnormality of PGI₂ metabolism in Henoch-Schönlein purpura is rather complex and heterogeneous. Further studies will be required to define the biochemical nature of the inhibitors and to establish whether low or absent PSA concentrations always reflect the presence of inhibitory activity and whether there is any evidence of rapid PGI₂ degradation. It seems likely that these abnormalities, having been shown in yet another vasculitic syndrome, are most probably a secondary manifestation of endothelial damage. The disturbances, however, may well be of importance in extending the primary microvascular insult.

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