Probiotics in the prevention of eczema: a randomised controlled trial

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ABSTRACT

Objective To evaluate a multistrain, high-dose probiotic in the prevention of eczema.

Design A randomised, double-blind, placebo-controlled parallel group trial.

Settings Antenatal clinics, research clinic, children at home.

Patients Pregnant women and their infants.

Interventions Women from 36 weeks gestation and their infants to age 6 months received daily either the probiotic (Lactobacillus salivarius CUL61, Lactobacillus paracasei CUL08, Bifidobacterium animalis subspecies lactis CUL34 and Bifidobacterium bifidum CUL20; total of 1010 organisms/day) or matching placebo.

Main outcome measure Diagnosed eczema at age 2 years. Infants were followed up by questionnaire. Clinical examination and skin prick tests to common allergens were done at 6 months and 2 years.

Results The cumulative frequency of diagnosed eczema at 2 years was similar in the probiotic (73/214, 34.1%) and placebo arm (72/222, 32.4%; OR 1.07, 95% CI 0.72 to 1.6). Among the secondary outcomes, the cumulative frequency of skin prick sensitivity at 2 years was reduced in the probiotic (18/171; 10.5%) compared with the placebo arm (32/173; 18.5%; OR 0.52, 95% CI 0.28 to 0.98). The statistically significant differences between the arms were mainly in sensitisation to cow’s milk and hen’s egg proteins at 6 months. Atopic eczema occurred in 9/171 (5.3%) children in the probiotic arm and 21/173 (12.1%) in the placebo arm (OR 0.40, 95% CI 0.18 to 0.91).

Conclusions The study did not provide evidence that the probiotic either prevented eczema during the study or reduced its severity. However, the probiotic seemed to prevent atopic sensitisation to common food allergens and so reduce the incidence of atopic eczema in early childhood.

Trial registration number ISRCTN26287422.

INTRODUCTION

The major atopic disorders, eczema, allergic rhinitis and asthma, cause significant disease burdens worldwide. Symptoms of atopy were reported to occur in 15–40% of children aged 13–14 years living in the UK in the mid-1990s1 and prevalence has increased in many countries in recent years.2-4

Despite the high atopic disease burden, pathogenic mechanisms remain poorly understood. Atopic disorders are heterogeneous resulting from complex interactions between environmental factors, including exposure to microbes, and host genes modulating innate and acquired immune responses, and mucosal and skin integrity.5 A critical issue is the role of atopy defined as a genetic propensity to develop immunoglobulin E antibodies following exposure to allergens6 and assessed by skin prick tests (SPTs) or measurement of specific IgE in serum. For eczema7 and asthma,8 atopic sensitisation early in life has been associated with persistent disease.

The hygiene hypothesis, based on the observation of increased atopy in smaller families,9 postulates that reduced exposure to infections in early childhood results in aberrant immunological responses to allergens.10 Intestinal microbiota provide a critical source of early immune stimulation and variations in early gut colonisation have been associated with the development of atopic disease.11 Probiotics are defined as live microorganisms which, when administered in adequate amounts, confer health benefits.12 The term ‘probiotic’ is commonly used for microorganisms that are undergoing evaluation for possible health benefits and it is this sense that is used here. Clinical trials suggest a role for probiotics in the primary prevention of atopic eczema.13 The mechanisms whereby probiotics may prevent atopy are unclear but might involve reduced exposure to allergens through improved epithelial barrier function and modulation of the developing immune system to prevent IgE sensitisation.14

We evaluated the effect of a probiotic comprising two strains of lactobacilli and two strains of bifidobacteria, or an inert placebo, administered to
mother-infant dyads in the perinatal period on clinical and laboratory manifestations of atopy. Here, we report the clinical manifestations of atopy at age 0–2 years.

METHODS

The safety profile of the probiotic and factors determining compliance with the trial procedures have been published.

Participants

Women aged 16 years or more, with a normal singleton pregnancy and intending to give birth at Singleton Hospital, Swansea were invited to join the study. Details of the recruitment process are described elsewhere. Women with known adverse conditions likely to affect them, the fetus, or the outcome of the pregnancy, were excluded. Women were informed of the trial, by letter, at booking, and provided with further information when attending for antenatal care. Those returning an expression of interest were contacted by researchers. Signed, informed consent was obtained by a researcher at 35–36 weeks gestation.

We had intended to recruit infants at ‘high-risk’ of developing atopy as those with a first degree relative with either asthma or eczema diagnosed by a health professional or allergic rhinitis treated by a doctor. However, sufficient details of atopy in first degree relatives were often difficult to obtain at recruitment. Therefore, in practice, infants with and without an increased risk of atopy were recruited. To preserve the integrity of the random allocation sequence, all infants are included in this report. Key findings relating to those at increased risk of atopy are presented in Webtable 1.

Randomisation

Pregnant women were allocated on a 1:1 basis at 36 weeks gestation to either the active or placebo arm of the study according to a computer-generated, random sequence without blocks. The random sequence was generated by the commercial partners and held independently of the research team. Women were allocated consecutively to the next number in the randomisation sequence and provided with the corresponding, preprepared intervention pack.

Intervention

We selected a multispecies probiotic preparation on the basis of greater efficacy than either single species or single organism preparations in the prevention or treatment of diseases in animal models and clinical trials. Species that had been evaluated in previous clinical trials, including the prevention of eczema were selected. Organisms were selected on the basis of activity in animal models and published information from previous trials. The active intervention comprised a vegetarian capsule containing Lactobacillus salivarius CUL61 (National Collection of Industrial, Food and Marine Bacteria (NCIMB) 30211) 6.25×10⁹ colony forming units (CFUs), Lactobacillus paracasei CUL08 (NCIMB 30154) 1.25×10⁹ CFUs, Bifidobacterium animalis subspecies lactis CUL34 (NCIMB 30172) 1.25×10⁹ CFUs and Bifidobacterium bifidum CUL20 (NCIMB 30153) 1.25×10⁹ CFUs as a freeze-dried powder. The dose in both arms was one capsule daily from 36 weeks gestation until delivery. Infants received the same capsules as their mother once daily from birth to age 6 months. Women were asked to store the capsules in the fridge and either took the capsule by mouth or sprinkled the contents onto food. The contents of the capsule were sprinkled directly into the infant’s open mouth or mixed with expressed breast milk or formula feed. Mothers were asked not to consume any commercially available probiotics or live yoghurts or administer these to their infants.

Unused capsules returned by participants from intervention and placebo groups for compliance monitoring were tested by an independent laboratory on an opportunistic basis. No live bacteria were identified in the placebo capsules, conforming with the random allocation sequence. Viability of the bacteria in the active preparation was consistent with the storage conditions confirming no significant deterioration in the product.

Data collection and allergen testing

Demographic information and possible risk factors for atopy were recorded at recruitment. Follow-up questionnaires were scheduled every 6 weeks to age 6 months and at 1 year and 2 years. A research assistant completed questionnaires with the parent/carer during a home visit at age 6 weeks, during research clinics at 6 months and 2 years and by telephone interview at other times. Where a follow-up was missed, parents/carers were asked to provide information for the period since the last questionnaire was completed. Questionnaires recorded any atopic disorders diagnosed by health professionals, treatments received and the occurrence and duration of common skin, respiratory and gastrointestinal symptoms.

At research clinics, infants were examined by either a clinician or a trained researcher. If present, the severity of eczema was evaluated by the scoring atopic dermatitis index. SPTs using common food allergens (cow’s milk, hen’s egg), aeroallergens (house dust mite, cat dander, grass pollen) and positive (histamine) and negative controls were performed. The response to an allergen was considered positive if there was a wheal diameter ≥3 mm.

Outcome measures

The primary outcome was the cumulative frequency of diagnosed eczema at 2 year follow-up (reported by parents/carers to have been diagnosed by a health professional or diagnosed during a research clinic). Eczema was defined as an itchy rash affecting the face, scalp or extensor surfaces of the limbs in infants and flexures in older children and of duration ≥4 weeks and with ≥1 exacerbation by age 24 months based on the information from questionnaires. Secondary outcomes were responses to SPTs, atopic eczema defined as eczema with one or more positive SPTs, eczema of any duration and whether or not diagnosed by a health professional, the severity of eczema, treatment with topical steroid preparations, respiratory symptoms with asthma and allergic rhinitis and reported food allergy. We had intended to report outcomes up to age 2 years. However, scheduled follow-up assessments were often delayed particularly when mothers had returned to work. Therefore, we have included all information for children aged up to, but not including, 3 years.

Data management and analysis

Strict blinding of the clinical research teams regarding participant allocation was maintained until after databases were locked following completion of data collection. The period of follow-up, defined as the time from birth to the age of the last follow-up questionnaire, was calculated for each infant.

Demographic variables, possible risk factors for atopy, and primary and secondary outcomes were analysed by treatment allocated. Findings for binary outcomes were expressed in ORs with 95% CIs. A \( \chi^2 \) test or Fisher’s exact test was used to compare proportions. Continuous outcome variables had skewed distributions and were summarised using median values and IQR and compared by the Mann-Whitney U test.

We performed adjusted analyses to investigate whether the trends in atopic sensitivity or atopic eczema were modified by baseline demographic variables (table 1), possible risk factors for atopy (table 2) and mother and infant compliance. For compliance, participants were classified as no compliance and compliance levels 1–4 representing the quartiles of the total number of days that trial interventions were taken. We used a set of logistic regression models, including the study treatment in each of days that trial interventions were taken. We used a set of logistic regression models, including the study treatment in each of days that trial interventions were taken. We used a set of logistic regression models, including the study treatment in each of days that trial interventions were taken.

Table 1  Demographic characteristics and possible risk factors for atopy at baseline

<table>
<thead>
<tr>
<th>Variable</th>
<th>Probiotic arm*</th>
<th>Placebo arm*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Socioeconomic status (Townsend rank; N; median, IQR)</td>
<td>220; 949 (333–1514)</td>
<td>234; 864 (330–1558)</td>
</tr>
<tr>
<td>Mother smoked during pregnancy</td>
<td>41/205 (20.0%)</td>
<td>47/218 (21.6%)</td>
</tr>
<tr>
<td>Vaginal delivery</td>
<td>152/216 (70.4%)</td>
<td>157/232 (67.7%)</td>
</tr>
<tr>
<td>Cat, dog, rodent or bird kept indoors</td>
<td>112/220 (50.9%)</td>
<td>120/234 (51.3%)</td>
</tr>
<tr>
<td>House</td>
<td>59/217 (27.2%)</td>
<td>56/232 (24.1%)</td>
</tr>
<tr>
<td>No. households with 1 adult</td>
<td>10/220 (4.6%)</td>
<td>9/234 (3.9%)</td>
</tr>
<tr>
<td>≥3 adults</td>
<td>20/220 (9.1%)</td>
<td>25/234 (10.7%)</td>
</tr>
<tr>
<td>no other children</td>
<td>81/220 (36.8%)</td>
<td>93/234 (39.7%)</td>
</tr>
<tr>
<td>≥3 other children</td>
<td>16/220 (7.3%)</td>
<td>18/234 (7.7%)</td>
</tr>
<tr>
<td>No. infants at increased risk of atopy†</td>
<td>197/220 (89.6%)</td>
<td>205/234 (87.6%)</td>
</tr>
</tbody>
</table>

This table gives the number of participants (%) unless otherwise stated. *Denominator varies according to information provided by parents/carers and compliance with follow-up. †Defined as first degree relative with either asthma or eczema diagnosed by a health professional or allergic rhinitis treated by a doctor.

Table 2  Potential modulating factors for atopy during follow-up

<table>
<thead>
<tr>
<th>Variable</th>
<th>Probiotic arm</th>
<th>Placebo arm</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast fed (full or partial)—any duration</td>
<td>49/191 (25.7%)</td>
<td>47/205 (22.9%)</td>
</tr>
<tr>
<td>Age last breast fed in months (N; median, IQR)</td>
<td>185; 1 [0–7]</td>
<td>190; 1 [0–5]</td>
</tr>
<tr>
<td>Attended child-minder in first 12 months</td>
<td>10/163 (6.1%)</td>
<td>16/172 (9.3%)</td>
</tr>
<tr>
<td>Attended nursery in first 12 months</td>
<td>65/162 (40.1%)</td>
<td>61/170 (35.9%)</td>
</tr>
<tr>
<td>Any oral/systemic antibiotics</td>
<td>155/214 (72.4%)</td>
<td>154/225 (68.4%)</td>
</tr>
</tbody>
</table>

This table gives the number of participants (%) unless otherwise stated.

Figure 1  Trial profile *Reasons for exclusion: pregnancy complication (n=110); presented after 36 weeks gestation or at the end of the recruitment period and before 36 weeks gestation (83); multiple pregnancy (14); contacted again after recruitment terminated (9); previous infant participated in trial (6); unwilling to stop current probiotic (1). †Reasons for failure to participate: lost contact with research team (288); not sufficiently interested or disliked tests (184); concern that trial was “too much to take on” (129); unwillingness to take the investigation products (85); not prepared to be allocated to the placebo arm (21); developed a medical condition (16); language difficulties (10); not willing for this child to be treated differently from previous child (7); bereavement (2). *Follow-up was often delayed; includes children seen up to, but not including, age 3 years.

Role of the funding source
Cultech UK advised on study design, provided the trial interventions and generated the random allocation sequence but was not involved in data collection, analysis or interpretation of the findings.

Ethics approval
The study was approved by the local research ethics committee in February 2004 and registered with Current Controlled Trials (ISRCTN26287422).

RESULTS
Participants
Recruitment began in May 2005 and the last scheduled 2 year contact was in November 2009. Of 1419 women attending antenatal clinics who had expressed an interest in the study, 454 were recruited and randomised (figure 1; Trial profile). Demographic variables and possible risk factors for atopy were similar in the 220 women randomised to the probiotic component.
arm and 234 to the placebo arm (table 1). Maternal smoking during pregnancy, keeping pets and houses affected by damp or mould were common in both arms. Three hundred and seventy-four women were carrying an infant at high risk of developing atopy with a similar proportion in each study arm (table 1). Potential modifying factors for atopy during follow-up, such as feeding practice, exposure to other people and to antibiotics, were also similar in the two arms (table 2).

Completion of questionnaires and attendance at research clinics was similar in the two arms (figure 1). Median (IQR) age at follow-up was 2.11 years (2.01–2.28 years) in the probiotic arm and 2.09 years (2.01–2.24 years) in the placebo arm (p=0.28). We have reported previously that follow-up was more complete among less deprived families and mothers who were non-smokers.

The trial arm was not associated with retention or clinic attendance at 24 weeks and 2 years.16

Primary outcome measure
The cumulative frequency of diagnosed eczema (a composite of carers’ reports and research clinic findings) at 2 years was similar in the probiotic (73/214; 34.1%) and placebo arms (72/222, 32.4%; OR 1.07, 95% CI 0.72 to 1.6; p=0.71).

### Secondary outcome measures
Positive SPTs to one or more common allergens at either age 6 months or 2 years were significantly less frequent in the probiotic arm (OR 0.52, 0.28–0.98; p=0.036, table 3). The number needed to treat to prevent one infant becoming sensitised was 13 (95% CI 7 to 173). The reduced skin prick responses in the probiotic arm were mainly to food allergens (cow’s milk and egg proteins) and statistically significant differences were already apparent at age 6 months. In contrast, sensitisation to aeroallergens (house dust mite, cat dander and grass pollen) occurred mainly after 6 months and was similar in the two arms (table 3).

Atopic eczema at 2 years was significantly less frequent in the probiotic arm (OR 0.40, 0.18–0.91; p=0.024) and was apparent by age 6 months (table 3). Differences between the two groups in non-SPT positive eczema were not statistically significant. At age 6 months, non-SPT positive eczema occurred in 37/147 (25.2%) children in the probiotic and 24/143 (16.8%) in the placebo arm (χ^2=3.06; p=0.08). At age 2 years, non-SPT positive eczema occurred in 57/171 (33.3%) children in the probiotic and 46/173 (26.6%) in the placebo arm (χ^2=1.86; p=0.17). The cumulative frequency of all eczema to age 2 years reported by parents and carers, whether or not diagnosed by a

### Table 3 Secondary outcomes according to intervention group

<table>
<thead>
<tr>
<th>Variable</th>
<th>Probiotic arm</th>
<th>Placebo arm</th>
<th>OR (95% CI)</th>
<th>p Value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>SPT† positive at 6 m</td>
<td>6/151 (3.97%)</td>
<td>16/147 (10.88%)</td>
<td>0.34 (0.13 to 0.89)</td>
<td>0.023</td>
</tr>
<tr>
<td>cow’s milk</td>
<td>0/148 (0.0%)</td>
<td>5/147 (3.40%)</td>
<td>–</td>
<td>0.030*</td>
</tr>
<tr>
<td>egg</td>
<td>5/148 (3.4%)</td>
<td>14/147 (9.5%)</td>
<td>0.33 (0.11 to 0.95)</td>
<td>0.032</td>
</tr>
<tr>
<td>house dust mite</td>
<td>1/151 (0.66%)</td>
<td>0/147 (0.0%)</td>
<td>–</td>
<td>0.51*</td>
</tr>
<tr>
<td>cat</td>
<td>0/151 (0.0%)</td>
<td>2/145 (1.4%)</td>
<td>–</td>
<td>0.24*</td>
</tr>
<tr>
<td>grass</td>
<td>1/150 (0.67%)</td>
<td>0/147 (0.0%)</td>
<td>–</td>
<td>0.49*</td>
</tr>
<tr>
<td>SPT† positive at either 6 m or 2 yrs</td>
<td>18/171 (10.5%)</td>
<td>32/173 (18.5%)</td>
<td>0.52 (0.28 to 0.98)</td>
<td>0.036</td>
</tr>
<tr>
<td>cow’s milk</td>
<td>1/171 (0.6%)</td>
<td>6/173 (3.5%)</td>
<td>0.16 (0.02 to 1.4)</td>
<td>0.12*</td>
</tr>
<tr>
<td>egg</td>
<td>9/171 (5.3%)</td>
<td>19/173 (11.0%)</td>
<td>0.45 (0.2 to 1.0)</td>
<td>0.052</td>
</tr>
<tr>
<td>house dust mite</td>
<td>9/171 (5.3%)</td>
<td>11/173 (6.4%)</td>
<td>0.82 (0.3 to 2.0)</td>
<td>0.66</td>
</tr>
<tr>
<td>cat</td>
<td>3/171 (1.8%)</td>
<td>7/173 (4.0%)</td>
<td>0.42 (0.1 to 1.7)</td>
<td>0.20</td>
</tr>
<tr>
<td>grass</td>
<td>2/171 (1.2%)</td>
<td>2/173 (1.2%)</td>
<td>1.0 (0.14 to 7.2)</td>
<td>0.99*</td>
</tr>
<tr>
<td>Skin</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atopic eczema at 6 m</td>
<td>4/151 (2.7%)</td>
<td>13/147 (8.8%)</td>
<td>0.28 (0.089 to 0.88)</td>
<td>0.021</td>
</tr>
<tr>
<td>Severity of eczema at 6 m clinic; median (IQR)</td>
<td>14.3 (7.5–17.9)</td>
<td>14.4 (10.6–24.9)</td>
<td>–</td>
<td>0.46</td>
</tr>
<tr>
<td>Atopic eczema at 2 yrs</td>
<td>9/171 (5.3%)</td>
<td>21/173 (12.1%)</td>
<td>0.40 (0.18 to 0.91)</td>
<td>0.024</td>
</tr>
<tr>
<td>Severity of eczema at 2 yr clinic; median (IQR)</td>
<td>11.1 (7.2–20.1)</td>
<td>14.2 (7.2–14.2)</td>
<td>–</td>
<td>0.85</td>
</tr>
<tr>
<td>All reported eczema§, ¶</td>
<td>119/214 (55.6%)</td>
<td>132/226 (58.4%)</td>
<td>0.90 (0.61 to 1.3)</td>
<td>0.55</td>
</tr>
<tr>
<td>Received topical steroid preparation</td>
<td>30/214 (14.0%)</td>
<td>40/226 (17.7%)</td>
<td>0.76 (0.45 to 1.2)</td>
<td>0.29</td>
</tr>
<tr>
<td>Respiratory</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>All reported asthma¶</td>
<td>23/171 (11.9%)</td>
<td>20/179 (10.0%)</td>
<td>1.2 (0.63 to 2.3)</td>
<td>0.57</td>
</tr>
<tr>
<td>Night-time cough</td>
<td>156/214 (72.9%)</td>
<td>164/226 (72.6%)</td>
<td>1.0 (0.67 to 1.5)</td>
<td>0.93</td>
</tr>
<tr>
<td>Night-time or daytime cough</td>
<td>178/214 (82.2%)</td>
<td>188/226 (82.2%)</td>
<td>1.0 (0.6 to 1.6)</td>
<td>0.99</td>
</tr>
<tr>
<td>Wheezing without symptoms of a virus infection</td>
<td>50/214 (23.4%)</td>
<td>55/171 (24.3%)</td>
<td>0.95 (0.61 to 1.47)</td>
<td>0.81</td>
</tr>
<tr>
<td>Inhaled bronchodilator or steroid</td>
<td>26/214 (12.1%)</td>
<td>27/226 (11.9%)</td>
<td>1.0 (0.57 to 1.8)</td>
<td>0.94</td>
</tr>
<tr>
<td>Allergic rhinitis¶</td>
<td>10/190 (5.3%)</td>
<td>10/201 (5.0%)</td>
<td>1.1 (0.43 to 2.6)</td>
<td>0.90</td>
</tr>
<tr>
<td>Sneezing and/or snuffling</td>
<td>207/214 (96.7%)</td>
<td>212/226 (93.8%)</td>
<td>1.95 (0.77 to 4.93)</td>
<td>0.15</td>
</tr>
<tr>
<td>Any reported food allergy</td>
<td>22/200 (11.0%)</td>
<td>31/204 (15.2%)</td>
<td>0.69 (0.38 to 1.2)</td>
<td>0.21</td>
</tr>
</tbody>
</table>

This table gives the number of participants (%) unless otherwise stated.

* χ^2 test for contingency tables or Fisher’s exact test* where expected values in cells were <5.

† SPT valid if diameter of wheal for the positive control was ≥1 mm than diameter for the negative control; SPT positive if diameter of wheal for antigen was ≥3 mm than diameter for negative control.

‡ SCORAD index.19

§ Whether or not diagnosed by a health professional.

¶ SCORAD, scoring atopic dermatitis; SPT, skin prick test.
health professional, its severity, the duration of rash and the use of topical steroids were similar in the two arms. The cumulative frequency of asthma and allergic rhinitis and symptoms and treatments consistent with these conditions and of reported food allergy were also similar in both arms (table 3).

The differences between the two arms in the risk of atopic eczema and SPT positivity among infants with one or more relatives with atopy were very similar to those in the whole cohort (webtable 1). However, with the exception of reduced sensitivity to cow’s milk protein at 6 months, differences in the smaller cohort did not reach statistical significance. Median (IQR) compliance with the trial interventions in pregnant women was 20 (12–28) days in the probiotic arm and 20 (12–28) days in the placebo arm (p=0.98, Mann-Whitney U test). In infants, the corresponding values were 106 (30–141) days and 103 (11–151) days (p=0.97, Mann-Whitney U test). In logistic regression analyses, in almost all cases, additional variables (table 1) were not significantly related to clinical outcomes and did not change the estimated OR for the effect of the probiotic. The exception was the presence of a smoker in the household; this was a significant risk factor for atopic eczema and atopic sensitivity (OR at 2 years: 2.48, 95% CI 1.08 to 5.70; p=0.033 and OR 2.25, 95% CI 1.16 to 4.37; p=0.017 respectively). This effect was independent of the treatment and did not significantly modify the univariate OR for the treatment effect. There was no evidence for an interaction effect between treatment and smoking allocation.

We have reported previously that probiotic administration was not associated with adverse effects in either mothers or their infants.15

DISCUSSION

Our findings do not support the administration of a multistrain, high-dose probiotic to mothers during late pregnancy and their infants from birth to age 6 months for the prevention of eczema in early childhood. However, probiotic administration was associated with a reduced frequency in sensitivity to food antigens and atopic eczema.

The strengths of our study are the relatively large number of mother-infant dyads recruited, the pragmatic nature of the trial, and confirmation of identity and viability of the probiotic organisms.

The study had several weaknesses. We had intended to evaluate clinical outcomes in infants at increased risk of atopy identified by having one or more first degree relatives with atopy. In practice, this was difficult to determine accurately at recruitment. However, the frequency of atopy and effects of the probiotic were similar in infants with and without a first degree relative with atopy. We expected that administering a novel intervention to healthy young infants would be difficult and, although compliance with the trial interventions in mothers was good, compliance was lower in infants. Finally, many mothers returned to work and this resulted in missed or delayed follow-up questionnaires and study clinic attendances. Although we compensated for missed follow-ups by subsequently asking parents and carers for information since the last assessment was done, this may have resulted in some loss of accuracy of information.

Our finding that the study probiotic did not affect the frequency of eczema is inconsistent with a meta-analysis of 13 randomised, placebo-controlled trials (3092 infants/children), where probiotic administration was associated with modest reductions in the frequency of eczema (fixed effects analysis; RR 0.79; 95% CI 0.71 to 0.88).24 Despite agreement between studies in the meta-analysis (I²=24.0%),24 variability in outcomes between studies are likely to result from differences in the probiotics used,25 the contribution of atopic and other pathogenic factors including variations in host mucosal and skin integrity,26 the diagnostic criteria, and outcome measures used for eczema.27

Our observations of reduced frequency of skin prick sensitivity to food allergens and eczema in children with demonstrated allergic sensitivity in the probiotic arm are consistent with those of a meta-analysis (20 cohorts; 4031 participants, including our findings) where probiotic administration during infancy and childhood was associated with reduced serum IgE and atopic sensitisation to food or inhalant allergens in random effects analysis with limited heterogeneity between studies and without evidence of publication bias.28

In our study, in contrast to food allergens, sensitivity to Aeroallergens was similar in the two arms. This tends to develop after infancy29 and, with the exception of two studies,30 31 most researchers have also reported that probiotic administration during early infancy did not reduce aeroallergen sensitivity.32–34 Our observation that probiotic administration did not reduce the frequency of asthma or wheeze in young children is consistent with other studies.28

CONCLUSION

Administration of probiotics in early life may have a role in the prevention of atopic sensitisation. However, a better understanding of strain-specific probiotic effects35 40 and how these address underlying atopic mechanisms is needed to guide the selection of strains for evaluation in clinical trials. Long-term follow-up to determine the effect of probiotic administration on the allergic march is also a priority.41

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