Isoniazid pharmacokinetics in children treated for respiratory tuberculosis

H S Schaaf, D P Parkin, H I Seifart, C J Werely, P B Hesseling, P D van Helden, J S Maritz, P R Donald

More than 50 years after its introduction as an antituberculosis agent, isoniazid (INH) continues to form the cornerstone of all “first-line” antituberculosis regimens and remains the only agent recommended for tuberculosis chemoprophylaxis in children. INH is most valued for its powerful bactericidal effect against the metabolically active organisms most commonly encountered in the sputum of adults with cavitating pulmonary tuberculosis and is the most valuable agent for preventing the development of resistance in companion agents. There is also evidence that INH suppresses the growth of non-multiplying organisms and that prolonged exposure of such organisms to concentrations above the minimal inhibitory concentration (MIC) leads to bacteriolysis.

INH is well absorbed from the gastrointestinal tract, but is subject to significant first pass metabolism that may impact on its systemic concentrations. INH does not bind appreciably to plasma proteins, crosses membranes readily, and distributes into a compartment that approximates the total body water. Very little of the parent compound is excreted unchanged in the urine and the greater proportion is acetylated in the liver and the small intestine to acetylsalicylic acid prior to excretion in the urine. Acetylation capacity in any individual is genetically determined, and although the initial studies of INH pharmacokinetics seemed to indicate that INH was bimodally eliminated, there was the suspicion that its elimination was in fact trimodal. Modern molecular biology techniques and improved analytical methods have now established, beyond doubt, that INH is eliminated in accordance with a trimodal distribution of subtypes, fast (FF), intermediate (FS), and slow (SS), the fast (F) and slow (S) alleles being co-dominant.

The pharmacokinetic characteristics of INH have been extensively studied in adults, but data in respect of children, and especially younger children are limited. Where such data are available, cognisance has not been taken of the genotype or of the trimodality of INH elimination.

This study was undertaken to improve our understanding of INH pharmacokinetics in children being treated for tuberculosis, making use of improved analytical technology and advances in our understanding of the polymorphisms governing INH metabolism.

PATIENTS AND METHODS

The study was undertaken in the Western Cape Province of South Africa, an area with a particularly high incidence of tuberculosis (>600/100 000 population at the time of this study). Children less than 13 years of age with primary respiratory tuberculosis and some with abdominal tuberculosis were included in the study following informed written consent of the parent or legal guardian. If not already hospitalised for other reasons the children enrolled in the study were temporarily admitted to hospital on the morning of the study.

The age and weight of each child were recorded and the extent of pulmonary involvement and the presence of extrapulmonary tuberculosis noted. Children who were severely ill were excluded from the study. With appropriate counselling and written informed consent the human immunodeficiency virus (HIV) status of the children was assessed.

The INH used for the study was standard pharmaceutical grade in powder form obtained from Fluka Chemie AG (Buchs, Switzerland). The INH was accurately weighed to

Abbreviations: AUC, area under the concentration curve; FF, homozygous fast acetylator; FS, heterozygous fast acetylator; INH, isoniazid; NAT2, N-acetyltransferase 2; SS, homozygous slow acetylator
give a dose of 10 mg/kg according to the child's weight the previous day. The INH powder was dissolved in 5–10 ml of water and administered by one of the study personnel orally with a syringe or, in the case of very young children, through a nasogastric tube and washed down with water. A light breakfast was permitted 60–90 minutes later.

Four blood specimens of 1–1.5 ml each, taken at 2, 3, 4, and 5 hours post-dose, were collected in ethylenediaminetetra-acetate (EDTA) coated tubes, chilled, and delivered on ice to the laboratory within one hour of taking the last specimen. INH concentrations were determined by the high performance liquid chromatography (HPLC) method of Seifart and colleagues. A further single 3 ml sample was collected into an EDTA coated tube for DNA analysis.

### NAT2 genotyping

Genomic DNA (gDNA) was prepared via the salting out procedure of Miller and colleagues. This gDNA was subsequently analysed for the \( \text{NAT2}^*5, \text{NAT2}^*6, \text{NAT2}^*7, \text{NAT2}^*12, \text{NAT2}^*13, \text{and} \text{NAT2}^*14 \) alleles, via a polymerase chain reaction (PCR) based strategy, as previously described. Separate PCR aliquots were restricted with the \( \text{Msp}^I, \text{Fok}^I, \text{Bam}^I, \text{Pst}^I, \text{Dde}^I, \) and \( \text{BamHI} \) restriction enzymes (according to the manufacturer's recommendations) to delineate the polymorphisms at nucleotide positions 191, 282, 481, 590, 803, and 857, respectively. According to the Vatsis nomenclature the wild type fast allele (F) is assigned as \( \text{NAT2}^*2, \text{NAT2}^*12, \) or \( \text{NAT2}^*13. \) These alleles confer normal enzyme activity on the NAT2 protein, while the mutant slow alleles (S), classified as \( \text{NAT2}^*5, \text{NAT2}^*6, \text{NAT2}^*7, \) and \( \text{NAT2}^*14 \) in humans, confer a decreased enzyme activity on the NAT2 protein.

The \( \text{T341C} \) mutation of the \( \text{NAT2}^*5 \) allele was typed via an allele specific PCR protocol, employing the confronting primer PCR method of Hamajima and colleagues. In this case the standard PCR mixture contained two primer sets, primer set I \([5'\text{AGCCCATTTCTGCAGCAT} \text{A}1063\text{nt-3} \) and \( \text{GAGCCTGAC} \) \( \text{CAT1063nt-3} \) and \( \text{GAGCCTGAGGAGGCTTGAG1349nt-3} \) which initiates amplification in the case of the \( \text{NAT2}^*5 \) allele (a 187bp product) and primer set II \([5'\text{1045nt-TTCTGCGAGA} \text{CAT1063nt-3} \) and \( \text{GAGCCTGAC} \) \( \text{CAT1063nt-3} \) and \( \text{GAGCCTGAGGAGGCTTGAG1349nt-3} \), which only amplifies in the case of the \( \text{NAT2}^*14 \) allelic sequence (a 323bp product). In addition, outermost primers \([876ntTTAGAG \text{GCTATTTTTGATCACA}897\text{nt} \) and \( \text{GAGCCTGAGGAGGCTTGAG1349nt} \) also amplify a gene specific PCR product (475bp) in the reaction mixture which serves as an internal amplification control.

The \( \text{PstI} \) restriction sites occur at nucleotide positions 1050 and 1347 of the \( \text{NAT2} \) gene sequence (GenBank accession number D10870); we have found these sites to be conserved in all samples that we have analysed thus far (more than 700). The use of this second enzymatic restriction improves the sensitivity of the analysis of the polymorphisms at 481, 857, and 191 respectively.

### Pharmacokinetic parameters

The apparent first order elimination rate constants \((k, \text{h}^{-1})\) of the individual patients were calculated from the linear regression of \( \ln(C_t) \) and \( t \), where \( \ln(C_t) \) is the natural logarithm of the concentration at time \( t \); observations of \( C_t \) were made at 2, 3, 4, and 5 hours after oral administration of 10 mg/kg body weight of INH. The area under the curve (AUC) over the interval 2–5 hours after the dose was calculated by integration of the AUC of each of the constituent subintervals (2–3, 3–4, and 4–5 hours after the dose) by standard methods.

### Statistical methods

Tests of homogeneity of group means were performed by one way analysis of variance. The general linear model approach was used for data subject to more than one classification criterion. The association between quantitative variables was examined by regression; in the case of the relation between age and \( k \) values the heteroscedasticity of the data required

### Table 1: Clinical features and special investigation results of the study children

<table>
<thead>
<tr>
<th>Clinical feature or special investigation</th>
<th>Number</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age group</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0–2 years</td>
<td>18</td>
<td>28</td>
</tr>
<tr>
<td>2–5 years</td>
<td>24</td>
<td>38</td>
</tr>
<tr>
<td>5–13 years</td>
<td>22</td>
<td>34</td>
</tr>
<tr>
<td>Weight loss</td>
<td>50</td>
<td>78</td>
</tr>
<tr>
<td>Weight &lt;3rd centile</td>
<td>31</td>
<td>48</td>
</tr>
<tr>
<td>Cough</td>
<td>39</td>
<td>61</td>
</tr>
<tr>
<td>Household TB contact</td>
<td>35</td>
<td>55</td>
</tr>
<tr>
<td>Mantoux test</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0–4 mm</td>
<td>13</td>
<td>20</td>
</tr>
<tr>
<td>5–14 mm</td>
<td>6</td>
<td>9</td>
</tr>
<tr>
<td>&gt;15 mm</td>
<td>41</td>
<td>64</td>
</tr>
<tr>
<td>Not done</td>
<td>4</td>
<td>6</td>
</tr>
<tr>
<td>HIV infected</td>
<td>13</td>
<td>20</td>
</tr>
<tr>
<td>Chest radiograph</td>
<td>44</td>
<td>69</td>
</tr>
<tr>
<td>Lymphadenopathy</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Collapse/aplasia</td>
<td>33</td>
<td>52</td>
</tr>
<tr>
<td>Pleural effusion</td>
<td>14</td>
<td>22</td>
</tr>
<tr>
<td>Milary</td>
<td>8</td>
<td>13</td>
</tr>
<tr>
<td>Extra-pulmonary TB</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Peripheral lymph nodes</td>
<td>14</td>
<td>22</td>
</tr>
<tr>
<td>Pleural effusion</td>
<td>14</td>
<td>22</td>
</tr>
<tr>
<td>Milary</td>
<td>8</td>
<td>13</td>
</tr>
<tr>
<td>TB meningitis stage 1</td>
<td>4</td>
<td>6</td>
</tr>
<tr>
<td>Pericardial effusion</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Culture or histology confirmed</td>
<td>41</td>
<td>64</td>
</tr>
</tbody>
</table>

### Table 2: The mean first order elimination rate constant \((k)\), the area under the curve (AUC) during the period 2–5 hours after dosing, and the mean serum INH concentrations at 2, 3, 4, and 5 hours after dosing with 10 mg/kg isoniazid (INH)

<table>
<thead>
<tr>
<th>Genotype ( \text{[n]} )</th>
<th>Mean INH concentration ((\text{SD})) ((\text{mg/l}))</th>
<th>2 h after dosing</th>
<th>3 h after dosing</th>
<th>4 h after dosing</th>
<th>5 h after dosing</th>
</tr>
</thead>
<tbody>
<tr>
<td>SS ( [25] )</td>
<td>0.254 (0.046)</td>
<td>18.356 (4.692)</td>
<td>8.599 (1.974)</td>
<td>6.585 (1.615)</td>
<td>5.099 (1.355)</td>
</tr>
<tr>
<td>FS ( [24] )</td>
<td>0.513 (0.074)</td>
<td>8.246 (3.349)</td>
<td>5.131 (1.864)</td>
<td>3.168 (1.294)</td>
<td>1.955 (0.885)</td>
</tr>
<tr>
<td>FF ( [15] )</td>
<td>0.653 (0.117)</td>
<td>5.371 (3.076)</td>
<td>3.938 (1.754)</td>
<td>2.045 (1.060)</td>
<td>1.125 (0.640)</td>
</tr>
<tr>
<td>Fl(d,df)</td>
<td>88.92 (2.61)</td>
<td>58.42 (2.55)</td>
<td>33.39 (2.59)</td>
<td>61.94 (2.61)</td>
<td>84.91 (2.61)</td>
</tr>
<tr>
<td>p value</td>
<td>&lt;0.0005</td>
<td>&lt;0.0005</td>
<td>&lt;0.0005</td>
<td>&lt;0.0005</td>
<td>&lt;0.0005</td>
</tr>
</tbody>
</table>

SD, standard deviation; SS, homoyzogous slow acetylators; FS, heterozygous fast acetylators; FF, homoyzogous fast acetylators.
the use of median regression. This is essentially robust regression that entails the fitting of parallel straight lines to the three groups of data.21

The Institutional Review Board of the Faculty of Health Sciences, Stellenbosch University, approved the study. The parents or legal guardians of all of the children gave written informed consent for their children’s participation in the study.

RESULTS

Sixty four children with a median age of 3.8 years (lower quartile 1.8 years, upper quartile 7.8 years) were included in the study. Table 1 summarises details of age and radiological and clinical features. Although 50 (78%) children had experienced recent weight loss, confirmed by reference to a “Road to Health” card, only 31 (48%) had a weight for age of less than 3rd centile (National Center for Health Statistics, USA). Thirteen (20%) children were HIV infected and 15 (23%) children suffered from abdominal tuberculosis. In 41 children (64%) tuberculosis was confirmed by isolation of M tuberculosis from one or more clinical specimens or identification of acid fast bacilli following tissue biopsy.

Twenty five children (39%) with median age of 2.8 years were genotyped as homozygous slow acetylators of INH, 24 (38%) with median age of 3.9 years heterozygous fast acetylators, and 15 (23%) with median age of 4.1 years as homozygous fast acetylators. With respect to the variable age, the differences between the genotype groups are not statistically significant.

Table 2 presents the mean first order elimination rate constant (k), the area under the concentration versus time curve (AUC) for the period 2–5 hours after dosing, and the mean INH concentration at 2, 3, 4, and 5 hours after administration of INH. The means of each of the variables differed significantly between genotypes (p < 0.0005 in each instance). In the case of the variable k the group variance increases with group means and for formal testing of homogeneity of group means the k values were transformed to logarithms; this had the effect of stabilising the variances. The ratio of the largest to the smallest variance gave observed F (25.22) = 1.49; this conservative test gives p = 0.18. A formal test of normality of residuals was not performed but dotplots indicated no obvious departure from normality. Although one way ANOVA is sensitive to lack of homoscedasticity it is robust against departures from normality. A Kruskal-Wallis test applied to k (or its logarithms) results in a similar χ² (49.17; degrees of freedom = 2, p < 0.0005).

Similar checks of the results relating to other variables in table 2, confirmed the statistical significance statements. Confidence limits (95%) for the location of the log transformed data are: SS (0.234 to 0.268), FS (0.477 to 0.538), FF (5.93 to 0.700). They are clearly well separated.

At the extremes the two hour INH concentrations varied from a high of 12 mg/l in a homozygous slow acetylator to a low of <2 mg/l in a homozygous fast acetylator. Within each genotype the means of k, AUC, and 2-hour, 3-hour, 4-hour, and 5-hour INH concentrations were not significantly associated with either HIV infection or the presence of abdominal tuberculosis.

With regard to the relation between age and k a regression of median values, taking into account heteroscedasticity of the data, confirms a significant decline in k with increasing age for each genotype. The results of fitting straight line regressions, k on age, with common slope parameter, were: slope = −0.00521, standard error 0.00114, giving p < 0.001. The fitted median values at the common mean age are 0.248, 0.500, 0.620 respectively for the three genotypes; they differ significantly from each other according to a global test (p < 0.0005), and the two values closest to each other, FS and FF also differ significantly. Table 3 summarises the result of fitting straight line regressions, INH concentration at each of the time points, 2-hours, 3-hours, 4-hours, and 5-hours on age with a common slope for the genotypes, and confirms a significant rise in each of the 2-hour, 3-hour, 4-hour, and 5-hour INH concentrations with age (p < 0.0005 in each instance).

In order to explore the relation of INH elimination and age further we compared the findings in this group of children with those obtained by our research group in a similar ethnic population of adults who were also genotyped and phenotyped after receiving 5 mg/kg and 10 mg/kg doses of INH. Among the adult patients INH concentrations were

<table>
<thead>
<tr>
<th>Genotype</th>
<th>k (SD) (h⁻¹)</th>
<th>AUC (SD) (mg/l/h)</th>
<th>2 h after dosing</th>
<th>3 h after dosing</th>
<th>4 h after dosing</th>
<th>5 h after dosing</th>
</tr>
</thead>
<tbody>
<tr>
<td>SS</td>
<td>0.193 (0.026)</td>
<td>24.870 (4.077)</td>
<td>10.942 (1.740)</td>
<td>8.943 (1.448)</td>
<td>7.433 (1.268)</td>
<td>6.169 (1.141)</td>
</tr>
<tr>
<td>FS</td>
<td>0.430 (0.080)</td>
<td>15.338 (4.018)</td>
<td>8.702 (1.841)</td>
<td>5.789 (1.446)</td>
<td>3.991 (1.225)</td>
<td>2.631 (1.015)</td>
</tr>
<tr>
<td>FF</td>
<td>0.678 (0.056)</td>
<td>8.139 (2.167)</td>
<td>6.031 (1.431)</td>
<td>3.043 (0.896)</td>
<td>1.748 (0.510)</td>
<td>0.877 (0.257)</td>
</tr>
</tbody>
</table>

SD, standard deviation; SS, homozygous slow acetylators; FS, heterozygous fast acetylators; FF, homozygous fast acetylators.

Table 3: Significance of a straight line regression of INH concentrations on age with a common slope for genotype

<table>
<thead>
<tr>
<th>Time after INH dosing (hours)</th>
<th>Ordinates at mean age</th>
<th>Slope</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>SS</td>
<td>FS</td>
<td>FF</td>
</tr>
<tr>
<td>2</td>
<td>8.622</td>
<td>5.078</td>
<td>3.882</td>
</tr>
<tr>
<td>3</td>
<td>6.650</td>
<td>3.128</td>
<td>2.001</td>
</tr>
<tr>
<td>4</td>
<td>5.150</td>
<td>1.924</td>
<td>1.092</td>
</tr>
<tr>
<td>5</td>
<td>4.048</td>
<td>1.129</td>
<td>0.604</td>
</tr>
</tbody>
</table>

SS, homozygous slow acetylators; FS, heterozygous fast acetylators; FF, homozygous fast acetylators.

*Significance of the slope coefficient.
determined at 0.5, 1, 2, 3, 4.5, and 6 hours after dosing. Table 4 presents the adults’ mean INH concentrations at 2 and 3 hours after dosing, together with the calculated 4- and 5-hour INH concentrations, k values, and AUC for the period 2–5 hours after dosing. These means may be compared with the paediatric data in table 2. However, for a more formal comparison of the two data sets we recall the basic model relating concentration and time of observation according to which log (concentration) is a linear function of time. The slope coefficient is the rate constant k, and the implication is that the data of any individual can be summarised in this constant and the intercept of the linear function. Equivalently the data can be summarised in k and the fitted ordinate at any chosen time. For this analysis we use the ordinate at 2 hours, and for convenience refer to it as A.

Genotype and adult/child status are treated as factors with three and two levels, respectively, in a two way analysis of variance to test for the significance of these factors. Table 5 shows the results of the ANOVA of variable k.

All effects are clearly highly significant. As a check a rank regression analysis was performed, as implemented in MINITAB, and essentially the same results were obtained. Inspection of tables 2 and 4 shows that the significant interaction is explained by the mean values of k being different between children and adults at genotypes SS and FS, but not at FF. To confirm, the Bonferroni confidence intervals with global confidence coefficient 0.95 for the differences between the adult and child means are: SS (0.0029 to 0.1201), FS (0.0274 to 0.1386), FF (−0.1017 to 0.0517). According to the ANOVA the overall means of the genotypes differ significantly. Bonferroni intervals, as above, for the differences are: SS–FS (−0.2826 to −0.2060), SS–FF (−0.4855 to −0.3895), FS–FF (−0.2423 to −0.1481).

Table 6 present the results of an analysis of variance of variable A. The genotype and adult/child main effects are clearly significant. These results have also been confirmed by a rank regression analysis. The Bonferroni confidence intervals with global confidence coefficient 0.95 for the genotype mean differences are: SS–FS (1.628 to 3.402), SS–FF (3.668 to 5.782), FS–FF (1.172 to 3.248). The conclusion is that the data of any individual can be summarised in A.

Finally, as further confirmation of the significance of the genotype and adult/child differences a multivariate ANOVA was performed with the joint response variables k and A. For the test of the main effect of factor adult/child the Wilk F-statistic is F(2,117) = 31.10, p < 0.0005; for the test of the main effect of factor genotype the Wilk F-statistic is F(4,234) = 80.28, p < 0.0005; and for the interaction it is F(4,234) = 2.40, p = 0.051.

After transforming the children’s weight for age to T scores, analysis of variance did not reveal any significant association of the pharmacokinetic parameters with body weight.

**DISCUSSION**

The results of this study illustrate once again the considerable differences in exposure to INH that exist between homozygous slow acetylators of INH and the heterozygous and homozygous fast acetylators, and show that these differences also exist in children. Our data also confirm that younger children eliminate INH faster than older children and in a trimodal model of INH elimination there is a significant age related decline in the first order elimination rate constant (k, h⁻¹) with age in all three genotypes. Furthermore the exposure of the children to INH, as reflected by the first order elimination rate constant, AUC for the period 2–5 hours after dosing, and INH concentrations at different time intervals after dosing, is significantly less than that of a group of adults drawn from the same population and receiving the same mg/kg body weight dose of INH. These findings, taking into account the NAT2 genotype for the first time, confirm the suggestions of earlier workers, based on phenotypification, that younger children eliminate INH faster than older children, and children, as a group, faster than adults.

The significantly faster elimination of INH by infants and younger children has been ascribed to the relatively greater mass of the liver in proportion to total body weight and it has been proposed that more optimal doses would be calculated on the basis of body surface area rather than body weight. Given that INH will most often be used in developing countries under programme conditions it is unlikely that this will be possible.

The normal range of INH concentrations two hours after dosing has been given as 3–5 mg/l; alternatively it has been suggested that a 3-hour concentration of 1.5 mg/l is desirable. It is therefore noteworthy that seven (35%) of the homozygous fast acetylators had a 2-hour INH concentration of less than 3 mg/l and nine (45%) did not reach a 3-hour post-dose concentration of 1.5 mg/l. It is inevitable that using a lower dose of 5 mg/kg body weight will lead to an even greater proportion of homozygous fast acetylators and probably a significant proportion of heterozygote fast acetylators failing to achieve the recommended concentrations.

The above findings provide justification for the official recommendations of some professional bodies that children should receive higher mg/kg doses of INH than adults. Thus while the International Union Against Tuberculosis and Lung Disease,28 the World Health Organisation,30 and the Joint Tuberculosis Committee of the British Thoracic Society41 recommend an INH dosage of 4–6 mg/kg body weight, the American Academy of Pediatrics recommends an INH dose of 10–15 mg/kg body weight. In debating the most appropriate dose of INH for use in children, some caution is necessary. Many of the studies documenting a satisfactory clinical response to an INH dose of 5 mg/kg emanate from populations with a predominance of homozygous slow acetylators; the relative therapeutic disadvantage of the heterozygote fast acetylator will also be concealed by the synergistic potential of a multidrug regimen. Under adverse circumstances, however, such as failure of full compliance, compromised absorption of INH itself, or its companion drugs, in particular rifampicin,31 or the sequestration of organisms in poorly perfused lesions,31–33 this relative disadvantage may be exposed. In the light of our findings, and in agreement with the recommendations of the American Academy of Pediatrics, we suggest that young children less than 5 years of age should receive an INH dose of at least 10 mg/kg to ensure that the faster acetylators of INH are exposed to adequate serum concentrations of INH.
Competing interests: none declared

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