LETTERS TO THE EDITOR

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Fludarabine in the treatment of an active phase of a familial haemophagocytic lymphohistiocytosis

EDITOR,—Familial haemophagocytic lymphohistiocytosis (FHL) is a lethal disease with an uncontrolled activation of T lymphocytes and macrophages due to a perforin gene defect.

The only current curative treatment is bone marrow transplantation. However, favourable outcome is associated with clinical remission status at the time of the procedure.

Unfortunatly, the use of steroids, etoposide (VP16), cyclosporin A, and antithymocyte globulins alone or in association frequently fails to control recurrent active phases.

BL, a 2 month old boy, was admitted in June 1999 for an active phase of FHL. His elder brother had died of FHL. The diagnosis was established on clinical (vomiting, fever, pallor, hepatosplenomegaly) and biological features (pancytopenia, hypertriglyceridaemia (3.82 mmol/l), haemodilution, hypofibrinogenaemia (0.95 g/l), a moderate elevation of aspartate transaminases (2N) and haemophagocytosis on bone marrow aspiration). No central nervous system abnormality was observed on cerebrospinal fluid analysis and cerebral magnetic resonance imaging.

A first remission was obtained with the combination of steroids: prednisolone (2 mg/kg/day), VP16-phosphate (150 mg/kg/day, d1–d7), cyclosporin A (4 mg/kg/day, continuous infusion), and antithymocyte globulins (10 mg/kg/day, d1–d5) three weeks after diagnosis. Despite maintenance treatment, relapse occurred one month later with severe pancytopenia. No remission was obtained with a second course of steroids, VP16-phosphate, and antithymocyte globulins. Two and a half months (day 76) after diagnosis, a course of fludarabine (30 mg/m²/day for four days) was initiated and dramatically improved our patient’s condition regarding the clinical and all biological criteria of FHL. An additional course was given on day 92. Transient neutropenia and a noticeable lymphopenia were observed. After a busulfan (120 mg/m²/day for four days) and cyclophosphamide (30 mg/kg/day for four days) conditioning regimen, one month after the last course of fludarabine, we performed a haemopoietic stem cell transplant with the father’s CD34+ HLA-half-identical peripheral cells (Miltenyi, Germany). Haematological reconstitution was observed from day 24 post transplant. There was transient grade II acute graft versus host disease (skin, liver). No relapse of FHL has occurred to this date (day 330 post transplant).

Treatment of active phases of FHL is based on drugs killing immunocompetent cells. Fludarabine is a purine antimetabolite with a strong immunosuppressive action. During treatment with fludarabine for B cell malignancies, an important decrease in the T cell subpopulations, particularly of the natural killer phenotype (CD16/56+), was observed. Our patient’s response to the first course of this drug was dramatic and allowed bone marrow transplant when in good clinical condition. Nevertheless a series of patients is needed to assess the efficacy of fludarabine for the treatment of active phases of FHL.

We thank Dr JL Stephan for his helpful clinical advice.

Correspondence to: Prof JP Vannier (Jean-Pierre.vannier@chu-rouen.fr)

Table 1

<table>
<thead>
<tr>
<th>Patient 1</th>
<th>Patient 2</th>
<th>Patient 3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age at diagnosis (years)</td>
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<td>1.5</td>
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<tr>
<td>Holiday destination where infected</td>
<td>South of France</td>
<td>Elba</td>
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<tr>
<td>Interval from exposure to appearance of symptoms (months)</td>
<td>7</td>
<td>12</td>
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<tr>
<td>Interval between appearance of symptoms and diagnosis (weeks)</td>
<td>6</td>
<td>3</td>
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<tr>
<td>Hepatomegaly (cm)</td>
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<td>1</td>
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<tr>
<td>Splenomegaly (cm)</td>
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<td>3</td>
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<td>LDH (U/l)</td>
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<td>Triglycerides (mmol/l)</td>
<td>3.64</td>
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</tr>
<tr>
<td>Serum IgG (g/l)</td>
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<td>15.6</td>
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</table>

LDH = lactate dehydrogenase

Fludarabine: A new therapy for familial haemophagocytic lymphohistiocytosis

Editors,—We read with interest the report by Baker et al. in *J Pediatr* [1] that investigated the potential of fludarabine for the treatment of active phases of FHL. Their conclusion was that fludarabine has potent activity in FHL, but that it needs to be established as an effective therapy by a phase III trial. We have recently treated a patient with FHL with fludarabine and would like to report our results.

Our patient was a previously healthy 2-month-old boy (Patient 1, Table 1) who presented with a high fever, pancytopenia, and haemophagocytosis. He was treated with standard therapy, including steroids, etoposide, cyclos porin A, and antithymocyte globulin. Despite this, his condition did not improve and he eventually required a bone marrow transplant.

Fludarabine was then initiated as a salvage therapy. His symptoms improved dramatically after the first course of fludarabine and he was able to undergo a bone marrow transplant. This confirmed our patient’s condition regarding the efficacy of fludarabine.

Fludarabine is a purine antimetabolite with a strong immunosuppressive action. It has been shown to be effective in the treatment of B cell malignancies. While the eradication of stray dogs may go a long way to reduce the incidence of VL, vaccination will be more desirable. Although resistance and immunity against the leishmania parasite is not well understood, the seemingly increasing incidence of VL in children travelling from Northern Europe might be because they have no transplacental immunity against the parasite and are therefore more prone to develop this condition than local children. There is much in common between the presentation features of the haemophagocytic syndromes and VL. It is noteworthy that all three of our patients showed signs of macrophage activation and haemophagocytosis was observed in their bone marrow smears. With increased awareness of this condition by physicians in non-endemic countries, the time required to reach the correct diagnosis and institute treatment should be reduced.
I have explored the validity of this assay for use in supernatants of faecal homogenates in children with cystic fibrosis and found it wanting. Recovery of a 500 pg/ml spike of IL-8 progressively increased from 41% in samples which were a 12-fold dilution of faeces to 189% in samples which were a 120 000-fold dilution of faeces, when used according to manufacturer’s instructions. Precluding the samples 50/50 in newborn calf serum, and using calf serum for further dilutions gave this assay (R&D catalogue no DB8000) mean (SD) spike recovery of 92.1 (12.5%) and coefficients of variation of 3.46% (intra-assay) and 6.85% (interassay). Without knowledge of IL-8-ELISA validation data of Smyth et al, I assume that this assay returns similarly spuriously high IL-8 concentrations in polyethylene glycol based wholegut lavage fluid to my 120 000-fold dilution faecal supernatant. The absence of a significant difference between CF patients and controls in their α1 antitrypsin output suggests that intestinal inflammation was not present in the CF patients. Overestimation of the WGLF IL-8 concentration would explain the apparently implausibly large volumes of swallowed sputum that the authors estimate would be required to account for their findings. In this study which could not turn on the mucociliary escalator, but did dramatically increase the rate of intestinal transit and exclude exogenous pancreatic enzymes, swallowed sputum is the most likely explanation for the results.

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Intestinal inflammation in cystic fibrosis

EDITOR,—Following their studies of whole gut lavage fluid, Smyth et al have suggested that a non-idiosyncratic intestinal inflammation occurs constitutively in patients with cystic fibrosis (CF), as a consequence of a proinflammatory effect of the patient’s CFTR mutations. They reported marginally elevated excretion of IgG, IgM, interleukin 1 (IL-1), neutrophil elastase, and eosinophil cationic protein, and much more significant increase in excretion of IL-8 and albumin, but no increase in excretion of α1 antitrypsin or IgA. In this study where lavage fluid was administered continuously, and intestinal effluent was collected in discrete samples, pooling of the effluent before analysis would have allowed small differences in calculated inflammatory marker outputs to be interpreted as representative of gastrointestinal output. Of all the inflammatory markers presented, only IL-8 shows a range of cytokine outputs in CF patients with or without fibrosing colonopathy that did not extend into the range seen in controls, in these non-parametric datasets. The author’s evidence for intestinal inflammation therefore relies heavily on the validity of their IL-8 Quantikine assay (R&D Minneapolis) protocol.

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Intestinal inflammation in cystic fibrosis: an alternative hypothesis

EDITOR,—I was interested by the report of Smyth and colleagues on the finding of markers of inflammation in whole gut lavage in patients with cystic fibrosis. As the α1 antitrypsin levels were not elevated when compared to controls, perhaps another hypothesis needs to be considered. Conceivably the inflammatory markers are not increased within the bowel, but rather, they are not degraded due to the lack of intestinal enzymes. α1 antitrypsin, which is resistant to proteolytic enzyme activity, would not be affected by such a phenomenon and, therefore, would be the same in patients with cystic fibrosis and controls. Perhaps the authors would need to resort to the somewhat dated technique of radio labelled albumin to definitively answer this question.

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Intestinal inflammation in cystic fibrosis

EDITOR,—We thank Dr Briars for his recent comments and are aware of his opinions regarding the potential source of the intestinal cytokines that we discussed in paper, including reference to his previous paper.1

We do not agree that our data is dependant upon IL-8 alone. We have shown statistically significant differences for a whole range of proteins and types of assays. Due to the large number of proteins and types of assays that we have performed, we have not carried out the extensive experiments for IL-8, as reported by Dr Briars. We do know that the polyethylene glycol used in a key constituent of the lavage fluid does not affect the IL-8 assay. There are two reasons why variable recovery is unlikely to be a major factor in our results. Firstly, by collecting whole gut lavage, any intestinal secretions present, including bile, intestinal substrates found in faeces (for example, sterocobulin) are effectively absent. Secondly, whole gut lavage is a perfusion system found to be equivalent to balloon perfusion systems. Thus, the dilution of any components would be very similar between the subjects and controls. Using whole gut lavage minimises any interference from intestinal material as much as is feasible in vivo.

Assuming the worst case scenario from Dr Briars data (that is, a two fold overestimate of IL-8 in the cystic fibrosis patients, which is not found in the controls), this still shows significantly increased IL-8 output in the cystic fibrosis patients (p<0.0001) and unfeasible volumes of sputum would still be required. For these, and reasons detailed in our paper and previous correspondence, we do not believe that sputum is the primary source of abnormalities found. Our observations concerning the increase in intestinal inflammatory markers in the whole gut lavage of cystic fibrosis patients have now been supported by a study which investigates intestinal inflammation within mucosal biopsy samples.3 This provides additional support to the hypothesis that the basic defect of cystic fibrosis transmembrane regulator can be proinflammatory.

Dr Eisenberg correctly points out the potential influence of pancreatic enzymes and degradation. The results we found for α1 antitrypsin were unexpected, given differences for albumin and IgG. Some discordance in data has been found previously in whole gut lavage from subjects with active inflammatory bowel disease who are pancreatic sufficient and who also can have raised intestinal permeability.1

However, our data that showed raised albumin and IgG are consistent with well established data showing raised intestinal permeability in children with cystic fibrosis. As we discussed, it has been found that protein outputs from balloon perfusion experiments (which exclude upper intestinal secretions) are similar to those found in whole gut lavage, which suggests that any potential effect of degradation from pancreatic IgG may be minimal.3 We also showed eosinophilic cationic protein to be raised in children with cystic fibrosis. As with α1 antitrypsin, this is relatively stable in faeces at room temperature (approx 21% loss over 24 hours). This loss would be considerably lower during whole gut lavage. Thus, degradation would be unlikely to explain this difference.

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Lumbar puncture should not be performed in meningococcal disease

Editor,—I was dismayed to see your publication of the letter by Dr Sam regarding the role of lumbar puncture in meningococcal disease.1 While fully understanding the need to get as much information as possible, the benefits of isolating the causative organism need to be weighed against the risk of causing clinical deterioration. It is time that textbooks of emergency paediatrics stated clearly that lumbar punctures on children with a haemorrhagic rash, and clinical signs of meningococcal infection, should not be carried out until the clinical condition has been stabilised, and only if the procedure will add further valuable information that cannot be obtained elsewhere.

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1 Sam WIC. The role of lumbar puncture in meningococcal disease. Arch Dis Child 2000;83:370.

Intestinal inflammation in cystic fibrosis: an alternative hypothesis

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Arch Dis Child 2001 84: 373
doi: 10.1136/adc.84.4.373d

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