Impact of HIV on mortality from acute lower respiratory tract infection in rural Zambia

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Abstract

Aims—To establish the prevalence and clinical correlates of HIV among children with acute lower respiratory tract infection.

Methods—Children admitted to a rural Zambian hospital were studied over an eight month period. The diagnosis of acute lower respiratory tract infection was made clinically, according to World Health Organisation (WHO) criteria. Clinicians, who were unaware of the children’s HIV status, prescribed antibiotic and supportive treatment according to WHO guidelines. HIV status was established using the polymerase chain reaction (AmpliCor HIV1, Roche) applied to dried blood spots.

Results—Acute lower respiratory tract infection was diagnosed in 132 children (median age 8 months, range 1 month to 4 years). The WHO criteria for severe or very severe pneumonia were met by 96/132 patients (73%) and 21 patients (16%) died. HIV dried blood spot PCR was positive in 14 cases (11%), of whom four fulfilled the WHO clinical case definition for paediatric AIDS and five died. The group as a whole were malnourished, but the HIV positive children were more severely malnourished (mean z score for weight = −3.01) than the HIV negative children (mean z score = −1.73, p < 0.001). The relative risk of death was 2.6 in the HIV positive group but this was not significant (p = 0.079).

Conclusions—An important minority of children with acute lower respiratory tract infection in rural Zambia will be infected with HIV. However, most HIV positive children presenting with respiratory infection will survive given simple antibiotic and supportive treatment.

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Keywords: HIV; acute lower respiratory tract infection; polymerase chain reaction; malnutrition

The diagnosis of HIV infection is difficult in children under 12 months of age. The World Health Organisation (WHO) has proposed a clinical case definition for paediatric AIDS but this has been shown to have a low sensitivity. Serological methods of testing for HIV are unreliable due to the possibility of transplacental transmission of antibodies from the seropositive mother. The use of the polymerase chain reaction (PCR), which detects HIV proviral DNA, has been established as a reliable method of identifying true HIV infection in young infants. This may be performed on whole blood or dried blood spots on filter paper. Dried blood spots are easy to collect, require only a small volume of blood, and store well at ambient temperatures.

The purpose of this study was to establish the prevalence and clinical correlates of HIV infection in a group of children with acute lower respiratory tract infection. We have employed the polymerase chain reaction for HIV proviral DNA on dried blood spots, collected on filter paper.

Methods

Patients with acute lower respiratory tract infection, admitted to the paediatric ward at St Francis Hospital over an eight month period from November 1994 to June 1995, were studied. The admission criteria for children with respiratory infections were severe or very severe pneumonia according to WHO guidelines (table 1), or pneumonia with another indication for hospital admission, for example severe malnutrition. Less severely ill children were treated as outpatients and were not eligible for entry into the study. Only children under 5 years were admitted to the children’s ward. Children under 4 weeks of age were not studied because of the difficulty of making a clinical diagnosis in this age group and to reduce the possibility of false negative results on PCR. Children were excluded if widespread wheeze was audible on auscultation of the chest or if their parents took them away from the hospital before treatment was completed.

Patients were examined clinically within 24 hours of admission by one of us (AS). Clinical features described in the WHO definition of severe and very severe pneumonia were noted (table 1), as were findings required for the HIV clinical case definition of paediatric AIDS (table 2). Each child was weighed and the oxygen saturation recorded using an Ohmeda Biox 3740 pulse oximeter (Ohmeda, Louisville, USA). The normal range for oxygen saturation at this altitude (1150 m) had previously
For the child age 2 months up to 5 years with cough or difficult breathing (who does not have stridor, severe undernutrition, or signs suggesting meningitis)

*Very severe pneumonia

Central cyanosis

Not able to drink

*Severe pneumonia

No central cyanosis and able to drink but chest indrawing

*Admission advised for both these categories

All patients with very severe pneumonia should receive oxygen. Where supply of oxygen is ample and makes use of a commercial PCR kit—the Amplicor HIV-1 kit (Roche Diagnostic Systems, New Jersey, USA) to amplify a sequence on a highly conserved region of the gag gene. Briefly, this uses a chelate extraction, after removal of haemoglobin from the sample. The amplification step uses biotinylated primers (Bio-SK431 and Bio-SK462) and 35 amplification cycles. The PCR product is detected by hybridisation to a DNA probe (SK102) bound to a microwell plate and uses an avidin–horseradish peroxidase conjugate to produce a colour change. This is quantified using a microwell plate reader, with a cut off value for optical density of 0.35 or greater A492 units. A serial dilution was performed with HIV seronegative blood ‘spiked’ with 8E5 cells—a cell line containing one copy of HIV proviral DNA per cell. This revealed that the assay could detect one copy of HIV proviral DNA per 6 mm filter paper disc. One 6 mm filter paper disc was analysed per patient and any equivocal results were repeated.

Statistical analysis was by means of the χ2 test and t test for unpaired data, using the Arcus Pro-stat statistical package; z scores for weight were calculated using the ‘Epi Nut’ program on the ‘Epi Info’ statistical package.

Ethical permission for the study was given by the ethics committee of St Francis Hospital and by the research ethics committee of the Royal Liverpool Children’s Hospital NHS Trust.

### Results

Over the eight month study period, 139 patients had a clinical diagnosis of acute lower respiratory infection. Wheeze was present in three patients, and four were taken away by their parents before treatment was complete. These patients were excluded from the study, leaving 132 patients. There were 64 boys (48%) and the median age was 8 months (range 1 month to 4 years). The WHO criteria for severe or very severe pneumoniaa were fulfilled by 96 patients (73%). There were 21 deaths (overall mortality rate 16%). A chest radiograph was taken in 84 patients, and unilateral or bilateral lobar or segmental consolidation was present in 68 (81%).

HIV dried blood spot PCR was positive in 14 cases (11%), of which only four fulfilled the WHO clinical case definition for paediatric AIDS and five died. The WHO clinical case definition was also fulfilled by three children who were HIV PCR negative. The WHO clinical case definition was found both in the HIV PCR positive and HIV PCR negative groups. Three mothers had previously been identified as HIV positive when serological testing was performed for clinical indica-

### Table 1  WHO clinical criteria for very severe and severe pneumonia

<table>
<thead>
<tr>
<th>Symptoms</th>
<th>Definition</th>
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<tbody>
<tr>
<td>Central cyanosis</td>
<td>Not able to drink</td>
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</tbody>
</table>

### Table 2  WHO clinical case definition for paediatric AIDS

| Major signs | Weight loss or failure to thrive |
| Minor signs | Generalised lymphadenopathy* |
| Oropharyngeal candidiasis | Repeated common infections (otitis, pharyngitis, etc) |
| Persistent cough (> 1 month) | Generalised dermitis |
| Confirmed maternal HIV infection |

Paediatric AIDS is suspected in an infant or child presenting with at least two major signs associated with at least two minor signs in the absence of known causes of immunosuppression. *Generalised lymphadenopathy = lymph nodes measuring at least 0.5 cm and present in two or more sites, with bilateral lymph nodes counting as one site.

been established by studying 85 asymptomatic children; an oxygen saturation below the 2.5 centile (92%) was taken to indicate hypoxaemia. Oxygen was given to those children who had an oxygen saturation less than 92%.

Antibiotics were given according to WHO guidelines. Laboratory facilities for microbiology were very limited, and blood and sputum culture were not performed. Chest radiographs were taken when the clinical diagnosis was uncertain or when complications, such as empyema, were thought to have occurred. Where clinical response to antibiotics was poor after seven days and the chest radiograph was suggestive of primary tuberculosis, then an empirical trial of antituberculous chemotherapy was begun. All chest radiographs were later examined by a radiologist (HC) who was not aware of the clinical features or the HIV status of the child.

Blood is routinely collected from all children admitted to the paediatric ward at St Francis Hospital for haemoglobin estimation and thick blood film for malaria parasites. A 50 μl aliquot of blood from this routine specimen was used to prepare a dried blood spot, using standard newborn screening papers (Schleicher and Schuell 903). These were allocated a code number and stored in 1.5 ml sealed plastic tubes (Sarstedt) at room temperature. Samples were returned to the United Kingdom for analysis. All records relating to children in this study were identified by a code number only.

The method used for the HIV PCR was as described by Cassol et al 7 and makes use of a commercial PCR kit—the Amplicor HIV-1 kit (Roche Diagnostic Systems, New Jersey, USA) to amplify a sequence on a highly conserved region of the gag gene. Briefly, this uses a chelate extraction, after removal of haemoglobin from the sample. The amplification step uses biotinylated primers (Bio-SK431 and Bio-SK462) and 35 amplification cycles. The PCR product is detected by hybridisation to a DNA probe (SK102) bound to a microwell plate and uses an avidin–horseradish peroxidase conjugate to produce a colour change. This is quantified using a microwell plate reader, with a cut off value for optical density of 0.35 or greater A492 units. A serial dilution was performed with HIV seronegative blood ‘spiked’ with 8E5 cells—a cell line containing one copy of HIV proviral DNA per cell. This revealed that the assay could detect one copy of HIV proviral DNA per 6 mm filter paper disc. One 6 mm filter paper disc was analysed per patient and any equivocal results were repeated.

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### Table 3  Frequency of occurrence of the clinical features of HIV infection (WHO clinical case definition) in HIV PCR positive and HIV PCR negative groups

<table>
<thead>
<tr>
<th>Medical Signs</th>
<th>HIV PCR positive (n=14)</th>
<th>HIV PCR negative (n=118)</th>
</tr>
</thead>
</table>
| Failure to thrive
  (>4 weeks) | 11 | 67 |
| Diarrhoea
  (>4 weeks) | 3 | 12 |
| Fever
  (>4 weeks) | 6 | 17 |
| Lymphadenopathy | 1 | 0 |
| Oral candidiasis | 3 | 12 |
| Repeated common infections
  (>4 weeks) | 6 | 7 |
| Cough | 6 | 25 |
| Generalised dermitis | 0 | 0 |
| Maternal HIV | 3 | 0 |
Impact of HIV on mortality from respiratory infection

The clinical case definition had a low sensitivity when compared with HIV PCR (sensitivity 31%, specificity 98%, positive predictive value 57%).

The HIV PCR positive and HIV PCR negative groups did not show significant differences in either age or gender. The two groups were similar when comparing the proportion from each group who fell into the categories of severe or very severe pneumonia, who had chest radiograph abnormalities consistent with pneumonia, and who were hypoxaemic. The chest radiographs of two children showed reticulonodular infiltrates consistent with lymphoid interstitial pneumonitis. Both were HIV PCR positive.

The group as a whole were malnourished (mean z score for weight = −1.86). The HIV positive children were considerably more malnourished (mean z score = −3.01) than the HIV negative group (mean z score = −1.73). The difference between the means was 1.28 (95% confidence interval −1.98 to −0.58, p < 0.001 when the unpaired two sample t test was applied).

Of those patients who were HIV PCR positive, five of 14 (36%) died compared with 16 of 118 (14%) who were HIV PCR negative. The odds ratio for death in the HIV PCR positive patients was 2.6 but this was not significant (p = 0.079).

Among the HIV PCR positive children, three of 14 (21%) were started empirically on a trial of antituberculous chemotherapy, compared with 11 of 118 (9%) of the HIV PCR negative group. However, the difference was not statistically significant (χ^2 = 0.35, p = 0.351). Other complications observed were pleural effusion, empyema, pneumothorax, pneumomediastinum, and pyogenic pericarditis. The numbers with each complication were small and in none of these was there a significant difference between the HIV positive and HIV negative groups.

Discussion

This study has shown that, in a rural hospital in Africa, the prevalence of HIV in children with acute lower respiratory tract infection is approximately 11%. This is, to our knowledge, the first time the PCR has been used to establish the prevalence of HIV among a group of African children with acute lower respiratory tract infection. The majority of children enrolled in this study were malnourished, but the HIV positive children were more malnourished than their HIV negative peers. Although the risk of death from pneumonia was increased by more than twofold in the HIV positive children, the majority survived when given simple antibiotic and supportive treatment.

The prevalence of antibodies to HIV among women living in rural areas in Zambia is known to be around 13%. The rate of vertical transmission of HIV from mother to infant in Zambia is approximately 39%. Therefore the prevalence of vertically acquired HIV infection among young children in rural Zambia would be expected to be around 5%. The relative risk of HIV among children admitted to hospital with acute respiratory infection in rural Zambia is therefore approximately twice what would be expected in the background population.

Lepage et al studied Rwandan children and found the sensitivity of the WHO clinical case definition for paediatric AIDS was low. They proposed a simplified clinical case definition in which respiratory distress secondary to lower respiratory infection was an AIDS defining feature. All the children described in the present study had some degree of respiratory distress secondary to lower respiratory tract infection and 73% had severe or very severe pneumonia by WHO criteria. However, only 11% had evidence of HIV infection as determined by PCR, and so the simplified clinical case definition proposed by Lepage et al performs poorly in our study population (specificity = 0%).

Chintu et al studied the seroprevalence of HIV in 1266 children admitted to hospital in Lusaka, the Zambian capital. They found that the overall prevalence of HIV in paediatric admissions was 28% and that the rate was the same among the subgroup of children with pneumonia. This prevalence is much higher than the rate using PCR, in our study. However, the seroprevalence of HIV among women of childbearing age in Zambian cities is approximately 30%, whereas in rural Zambia the rate is around 13%. The lower prevalence of HIV in children found in our study is therefore in proportion to the lower seroprevalence seen in the adult population in rural areas. Chintu et al also found a high seroprevalence of HIV among children with malnutrition (41%). Mortality from acute lower respiratory infection was similar in children who were HIV positive (11%) and HIV negative (4%) in their study.

Nathoo et al, in Zimbabwe, found clinical or serological evidence of HIV in 31% of urban children with acute lower respiratory tract infection, and moderate to severe malnutrition (weight for age < 80%) in 30%. The case fatality rate was 16% and the risk of death was increased over threefold in children who had evidence of HIV infection. In our study 71% of the children were less than 80% of weight for age. The reason for such poor nutrition is obscure. Chronic diarrhoea occurred in only 15/131 patients (11%) (table 3) and was no more common in the HIV positive group. However, in Katete district alone over 2400 children (3.4%) have lost both parents to HIV related illness (JM Cairns, personal communication). Although these children are cared for by the extended family, the financial strain is considerable and they may be at greater risk of malnutrition as a result. We observed similar mortality rates to the children studied in Zimbabwe, and a similar increase in mortality in the HIV positive group.

The present study employed the polymerase chain reaction, using dried blood spots, to detect infection with HIV. This method has been shown to be highly sensitive (detecting 99.4% of infants vertically infected with HIV
when the sample is taken after 15 days of age) and specific (specificity = 100%). An assay of dried blood spots ‘spiked’ with 8E5 cells, performed as part of the present study, showed that one copy of HIV proviral DNA could be detected in a 6 mm filter paper punch. As the target sequence is a highly conserved region, the probability of false negative results due to strain variation is low. Dried blood spot samples are robust. Storage at 22°C for three months and freeze-thawing twice have been shown to have no effect on PCR reactivity. In the present study samples were stored for up to eight months at temperatures of around 22°C and it is possible that some loss of PCR reactivity and reduction in the sensitivity of the assay may have occurred. However, there are no published data on storage for longer periods.

In the first year of life, the possibility of transplacental transmission of antibodies to HIV makes the use of HIV serology unreliable. Although 75% of infants will lose maternal antibody to HIV by 1 year of age, a significant proportion of children who become antibody negative will be infected with HIV. HIV PCR represents a more reliable way of making the diagnosis of HIV infection in young children. This study used clinical criteria to diagnose acute lower respiratory tract infection and this may have lead to diagnostic imprecision. However, chest radiographs are expensive to perform in the developing world. Most health workers will have to make this diagnosis on clinical grounds alone and so it seemed appropriate to use clinical criteria in this study. The observation that consolidation was present in 81% cases where a chest radiograph was taken suggests that the clinical diagnosis was correct in most cases.

We did not study children over the age of 5 years. The oldest child found to be HIV PCR positive was aged 3 years, and the median age of the HIV PCR positive children was 11 months. Of the children over 1 year, three of six died while still hospital inpatients. In a survey of children in hospital in Kampala, Uganda, 80% of HIV positive children were under 2 years and 23% died within three months. We found that HIV infected children had a good response to conventional treatment for pneumonia, it seems unlikely that many children will survive to reach their fifth birthday. Long term follow up data are sorely needed on children vertically infected with HIV in the developing world.

In conclusion, an important minority of children with acute lower respiratory tract infection in rural Zambia will be infected with HIV. However, most HIV positive children presenting with respiratory infection will survive given simple antibiotic and supportive treatment. Malnutrition was a more severe problem in the HIV infected group and more research is needed to establish whether intensive nutritional rehabilitation can reduce mortality in these children. Further work now needs to be done to establish if the pathogens associated with acute lower respiratory tract infection in the developing world have changed in association with the HIV epidemic.

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