Inaccuracy of glomerular filtration rate estimation from height/plasma creatinine ratio


Abstract

Use of a height/plasma creatinine formula to estimate glomerular filtration rate (GFR) is simpler and less invasive than renal or plasma clearance methods. The aim of this study was to determine whether these formulas enabled accurate prediction of GFR measured from the plasma clearance of $^{51}$Cr labelled ethylenediaminetetra-acetic acid ($^{51}$Cr-EDTA). Thirty nine patients underwent GFR measurement at least six months after potentially nephrotoxic chemotherapy. Altman-Bland analysis was performed on the measured GFR and that estimated simultaneously using the original and a modified Counahan-Barratt formula and the Schwartz formula. The limits of agreement of the estimated GFR with the measured GFR were unacceptably wide in each case, despite highly significant correlation coefficients. The bias was smallest for the modified Counahan-Barratt formula. Use of these formulas to estimate GFR in children is insufficiently accurate for research purposes and has limitations in clinical practice. Furthermore, use of correlation coefficients to evaluate different methods of measuring GFR is inappropriate.

(Arch Dis Child 1994; 70: 387–390)

Measurement of the glomerular filtration rate (GFR) in children and adolescents is an important part of the assessment of many renal diseases, and may facilitate safer prescribing of fluids, electrolytes, and drugs. It is of particular value in children with malignant disease in whom it may provide an important measure of toxicity, and influence the type and dose of cytotoxic drug treatment.

In paediatric practice, however, both the acceptability and accuracy of conventional renal clearance methods of GFR measurement are reduced by the need for carefully timed and complete urine collections. The utility of the inulin clearance method is limited further by the requirement for a continuous intravenous infusion, and the cumbersome nature of the inulin assay. The utility of endogenous creatinine clearance measurement is diminished by variability in the dietary intake and tubular secretion of creatinine, and by interference of non-creatinine chromogens with the plasma creatinine assay. There is often considerable discrepancy between GFR values from the two techniques in individual patients, with the latter method usually giving higher results due to tubular creatinine secretion.

These disadvantages prompted the introduction of clinical measurement of GFR from the plasma clearance of ethylenediaminetetraacetic acid labelled with $^{51}$chromium ($^{51}$Cr-EDTA), without the necessity for urine collection. Initial studies in adults and in children showed that $^{51}$Cr-EDTA plasma clearance corresponded closely to simultaneously measured renal inulin clearances. The simplicity, wide availability, and low radiation exposure of radioisotopic plasma clearance methods have led to their widespread adoption in paediatric practice.

Several attempts have been made to introduce and validate simpler methods whereby GFR can be estimated from the plasma creatinine concentration ($P_c$), assuming steady state conditions. Two well known formulas have been published for use in children, both of which allow determination of GFR from $P_c$ and height. These formulas have been validated by the use of correlation coefficients to compare the measured and estimated GFR, but this method is inappropriate and does not indicate whether the estimated GFR is an acceptably accurate substitute for measured GFR.

The aim of this study was to determine whether GFR measured from the plasma clearance of $^{51}$Cr-EDTA could be predicted accurately and reliably by use of a formula relating the patient’s height and $P_c$.

Patients and methods

Data were collected prospectively during a study of renal function after treatment in a cohort of 39 consecutive children and adolescents (22 male) with malignant solid tumours completing treatment with potentially nephrotoxic chemotherapy (cisplatin (23 patients), ifosfamide (13), or both (three)) at the Paediatric Oncology Unit in Newcastle upon Tyne. All patients were investigated at least six months after completion of chemotherapy for neuroblastoma (10 patients), rhabdomyosarcoma (six), soft tissue sarcoma (five), Ewing’s sarcoma (five), osteosarcoma (four), brain tumour (four), germ cell tumour (three), and hepatic tumour (two).

The patient’s height was measured using a stadiometer (Raven Equipment Ltd). No patient was underweight, but four were overweight. No patient had clinically detectable oedema, pleural or ascitic fluid.

The $^{51}$Cr-EDTA plasma clearance determination of GFR was performed using a single
intravenous injection of radioisotope at time 0, with subsequent collection of plasma samples at 1, 2, and 4 hours. A straight line was fitted to the log activity time curve, assuming adequate distribution of the tracer by 1 hour, and the half time (t1/2) of the radioisotope determined. The volume of distribution (Vd) was found from the estimate at time 0. The plasma clearance was calculated from

\[
\text{Plasma clearance (ml/min) = } \frac{V_d \times 0.693}{t_{1/2}}
\]

and then expressed per 1.73 m² surface area, giving the measured GFR.

The Pcr of one of the above blood samples was measured by a Technicon SMA Auto-Analyzer (Jaffe method with initial dialysis). The estimated GFR was calculated in three ways:

1. the Counahan-Barratt formula:

\[
\text{GFR} = \frac{38 \times \text{ht}}{\text{P}_{\text{cr}}}
\]

where GFR = glomerular filtration rate (ml/min/1.73 m²), ht = height (cm), and Pcr = plasma creatinine concentration (µmol/l).

2. a modified Counahan-Barratt formula (Morris et al):

\[
\text{GFR} = \frac{40 \times \text{ht}}{\text{P}_{\text{cr}}}
\]

3. the Schwartz formula:

\[
\text{GFR} = \frac{k \times \text{ht}}{\text{P}_{\text{cr}}}
\]

where k = 48.6 except in males ≥13 years age, in whom k = 61.9.

The values of k (48.6 and 61.9) used in formula (3) are derived by multiplying those figures (0.55 and 0.70) quoted by Schwartz et al by a factor of 88.4 to allow for expression of Pcr in µmol/l in this study, rather than mg/dl. Counahan et al used the true Pcr (either measured directly or derived by subtracting 12.4 µmol/l from the Pcr measured by the Technicon method). Morris et al used a reaction rate method for Pcr measurement, and Schwartz et al used a Technicon method modified to reduce interference by non-creatinine chromogens. Therefore, each of the above formulas was calculated twice in all patients – once using the measured Pcr, and once using the 'corrected' Pcr, which was calculated by subtraction of 12.4 µmol/l from the measured value.

Two methods of statistical analysis were used. First, an Altman-Bland analysis was performed. The difference between measured and estimated log GFR was plotted against the average of the measured and estimated log GFR. Further, the ratio geometric mean of measured GFR: geometric mean of estimated GFR was calculated together with 95% 'reference ranges' for this ratio (the 'limits of agreement' of the measured and estimated GFR). Logarithmic transformation of the data was necessary since the spread of the difference between measured and estimated GFR appeared to increase with GFR. Therefore, the limits of agreement are expressed in terms of ratios rather than absolute differences.

Second, the correlation coefficient (r) was calculated, as in previous studies.

### Results

The median (range) height, non-corrected Pcr, measured GFR, and each of the estimated GFRs (using the non-corrected Pcr) are shown in table 1.

<table>
<thead>
<tr>
<th>Age at study (years)</th>
<th>7.1 (2.2-18.5)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Height (cm)</td>
<td>118.0 (88.0-177.5)</td>
</tr>
<tr>
<td>Plasma creatinine (µmol/l)</td>
<td>59 (26-202)</td>
</tr>
<tr>
<td>GFR (ml/min/1.73 m²)</td>
<td>5.5 (3.6-17.3)</td>
</tr>
</tbody>
</table>

### Table 1: Patient details and GFR results; figures are median (range)

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CB-38 = GFR estimated from formula (1); CB-40 = GFR estimated from formula (2); Schwartz = GFR estimated from formula (3); all are calculated using non-corrected plasma creatinine concentration.

### Figure 1: An Altman-Bland plot of the difference between the log GFR measured from ⁵¹Cr-EDTA and the log GFR estimated from the Counahan-Barratt formula (using the non-corrected plasma creatinine concentration) against the average of the measured and estimated log GFRs. The mean (1.96 SD) value of the difference is shown.
Table 2 Bias and limits of agreement of measured and estimated GFRs

<table>
<thead>
<tr>
<th>Bias*</th>
<th>Mean</th>
<th>95% CI</th>
<th>p Value</th>
<th>Limits of agreement†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Using non-corrected plasma creatinine concentration</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Measured GFR vs CB-38</td>
<td>1.12</td>
<td>0.97-1.22</td>
<td>0.009</td>
<td>0.67-1.89 (see fig 1)</td>
</tr>
<tr>
<td>Measured GFR vs CB-40</td>
<td>0.84</td>
<td>0.70-0.92</td>
<td>&lt;0.001</td>
<td>0.48-1.42</td>
</tr>
<tr>
<td>Using corrected plasma creatinine concentration</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Measured GFR vs CB-38</td>
<td>0.86</td>
<td>0.78-0.95</td>
<td>0.005</td>
<td>0.47-1.58</td>
</tr>
<tr>
<td>Measured GFR vs CB-40</td>
<td>0.82</td>
<td>0.74-0.91</td>
<td>&lt;0.001</td>
<td>0.45-1.50</td>
</tr>
<tr>
<td>Measured GFR vs Schwartz</td>
<td>0.64</td>
<td>0.57-0.71</td>
<td>0.001</td>
<td>0.33-1.22</td>
</tr>
</tbody>
</table>

*Bias is the ratio geometric mean of measured GFR: geometric mean of estimated GFR (see text); 95% CI = 95% confidence interval of the bias; p = probability that the bias is equal to unity. †Limits of agreement are the 95% reference range for the bias.

be surprising if the results obtained from two methods designed to measure the same quantity were not related. Furthermore, as the value of the correlation coefficient (for this application) depends on both the variation between individuals and that within individuals (that is, measurement error), large interindividual variation relative to measurement error will give a high correlation coefficient, with a highly significant p value.

The bias is significantly different from unity, except in the case of the modified Counahan-Barratt formula using non-corrected Pcr. As expected, use of the corrected Pcr increases the mean estimated GFR. However, this confers no advantage as the corrected bias is even further from unity than the non-corrected bias with each of the three formulas used.

Data presented by Burghard et al gives a correlation coefficient of 0.91 for 50 subjects, comparing renal clearance of inulin and GFR estimated from the Counahan-Barratt formula. However, inspection of their figure still shows considerable scatter. The higher value of the correlation coefficient is largely due to the wider range of the measured GFR in their patients. When the data of Burghard et al are analysed in the same way as ours, the results are comparable — although the bias (0.97; with 95% confidence interval 0.89 to 1.05) is close to unity, the limits of agreement are just as wide (0.53-1.75).

Reference to the initial publications describing the derivation and validation of estimated GFR formulas shows that correlation and regression analyses were used, and that considerable scatter was present despite the high correlation coefficients. Counahan et al and other authors have commented on the wide confidence limits for estimated GFRs in individual patients. Although it has been suggested that the use of estimated GFR may be appropriate to screen children by identifying those who are very likely to have a normal or an abnormal GFR, it has also been acknowledged that a renal inulin clearance or radioisotopic plasma clearance method is necessary when an accurate GFR measurement is needed for either clinical or research purposes.

In general, inaccuracy of estimated GFR may be due to interindividual differences in the modified Counahan-Barratt formula when the non-corrected Pcr is used.

Furthermore, the limits of agreement are extremely wide in all three formulas, using either the non-corrected or the corrected Pcr. For example, using the Counahan-Barratt formula and non-corrected Pcr, the measured GFR will be between 0.69 and 1.89 times the estimated value in 95% of cases; when the corrected Pcr is used, the equivalent figures are 0.47 and 1.58.

The correlation coefficients between the measured GFR and each of the estimated GFR values are shown in table 3, and the scatter plot of measured GFR versus the estimated GFR (from the Counahan-Barratt formula, using the non-corrected Pcr) is shown in fig 2. Although all the correlation coefficients are highly statistically significant, the figure illustrates the considerable discrepancies between the measured and estimated GFR in many individual patients.

Discussion

The most important finding of this study is the considerable disagreement between measured and estimated GFR in individual patients. The width of the limits of agreement with measured GFR prevents the use of the Counahan-Barratt and Schwartz formulas in circumstances where accurate determination of the GFR is required for either clinical or research purposes. Use of the corrected (rather than the non-corrected) Pcr fails to narrow the limits of agreement.

The use of the correlation coefficient is misleading and meaningless in this situation, and the significance value is irrelevant. Moreover, it is apparent from table 3 and fig 2 that a highly significant correlation coefficient may exist despite considerable scatter. The correlation coefficient measures linear association rather than agreement. Two variables may be related without being in agreement; indeed, as Altman and Bland have commented, it would
body habitus, including reduced muscle bulk in children with cancer, and the use of different methods of PCr measurement. This study comprised an apparently typical group of children who had completed treatment for cancer. None were still malnourished as judged by their weight for height for age, and the estimated GFR of the four overweight patients did not appear to be unrepresentative of the group as a whole. A regression equation between height and PCr derived from one group of children might not be applicable to another group with a different range of body habitus, and the convenience of GFR estimation is lost if validation is required in individual populations before it can be confidently applied.

As there is no clear guidance on desirable levels of accuracy in either clinical or research practice, individual judgment must be exercised for each particular application of a GFR measurement or estimation method. Nevertheless, the results of this study demonstrate that it is not possible to predict accurately the GFR measured from the plasma clearance of 51Cr-EDTA in children treated with potential nephrotoxins by using a formula relating height and PCr. Although the method may be adequate for some clinical purposes (for example, identifying those children in whom GFR is unlikely to be below a certain threshold\(^1\)), the limits of agreement shown in table 2 are clearly far too wide to allow confidence in these GFR estimations for either guidance in the clinical decision of whether or not to give further potentially nephrotoxic chemotherapy, or for comparison of the glomerular toxicity of particular treatments.

Furthermore, comparison of GFR measurement methods (including the initial studies of 51Cr-EDTA plasma clearance\(^5-8\)) has usually been based on the use of correlation and regression analysis, which is not an adequate tool to determine whether two methods of measuring the same variable can be used interchangeably. Reappraisal of currently used methods of GFR measurement in children should include appropriate statistical evaluation.

Dr R Skinner was a MRC Training Fellow. We also thank the Special Trustees of Newcastle Health Authority and the North of England Children’s Cancer Research Fund for additional financial support.

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Arch Dis Child 1994 70: 387-390
doi: 10.1136/adc.70.5.387

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