Defective alloantigen-presenting capacity of ‘Langerhans cell histiocytosis cells’

Raymond C H Yu, Jenny F Morris, Jon Pritchard, Tony C Chu

Abstract
The functional activity of skin cells derived from an infant who died of multisystem Langerhans cell histiocytosis (LCH) was examined. Involved and non-involved skin was obtained at postmortem examination within three hours of death; normal epidermal Langerhans cells and ‘LCH cells’ were separated by means of dispase digestion. The functional activity of different populations of CD1a positive cells was assessed using the conventional six day allogeneic mixed cell reaction. Compared with Langerhans cells from a healthy control, LCH cells showed minimal functional activity. However, Langerhans cells from non-involved skin showed normal and Langerhans cells overlying involved skin showed augmented functional activity. These findings suggest that LCH is a disease in which abnormal Langerhans cells accumulate and/or proliferate in various tissues but it does not affect the entire Langerhans cell population. (Arch Dis Child 1992;67:1370–2)

Langerhans cell histiocytosis (LCH), a rare disorder of unknown cause affecting both children and adults, can involve many different organs and present to a wide range of medical specialties. LCH is now regarded as a ‘reactive’ disorder in which cells bearing the Langerhans cell phenotype accumulate and/or proliferate in physiologically normal and abnormal sites.12 The subsequent local tissue damage may cause organ dysfunction and in severe cases, death. The tissue destruction in LCH is believed to be at least partly the result of excess cytokine production.3 Although lesional ‘LCH cells’ express most of the phenotypic markers as epidermal Langerhans cells including HLA-DR and the CD1a complex, it is not known whether these cells carry the functional capacity of Langerhans cells. Functional studies have been severely hampered by the scarcity of fresh pathological specimens.

Case report
An 8 month old boy presented with a skin rash, discharging right ear, and easy bruising from the age of 3 months. Moderate hepatosplenomegaly was evident. Investigations showed anaemia (haemoglobin concentration 65 g/l) and thrombocytopenia (platelet count 3·0·10^11/l). A skin biopsy specimen and bone marrow aspirate both demonstrated patchy infiltration by large cells with cleaved nuclei which showed positive immunostainings to S100 and peanut agglutinin. A diagnosis of multisystem LCH was made according to the criteria of the Histiocyte Society.4

The child’s illness progressed over the next nine months with pancytopenia requiring frequent transfusions and worsening hepatosplenomegaly. Several systemic agents including high dose systemic prednisolone, etoposide, vincristine, and cyclosporin were tried with only transient benefit. From October 1990 he developed persistent unexplained abdominal pain, distension, and diarrhoea and in February 1991 he died of progressive disease. Postmortem examination carried out within three hours of death showed involvement of the bone marrow, lungs, skin, gastrointestinal tract, pancreas, liver, and spleen by LCH.

In order to assess the functional activity of LCH cells, samples of clinically normal skin and skin affected by LCH were taken. Small samples were snap frozen for immunohistochemistry using anti-CD1a monoclonal antibodies (MAbs) to identify the presence and localisation of Langerhans cells and LCH cells. Skin obtained from an adult patient who underwent abdominoplasty provided control epidermal Langerhans cells.

ASSESSMENT OF LCH AND LANGERHANS CELL FUNCTION
An allogeneic mixed cell reaction was used to assess the functional activity of Langerhans cells and LCH cells. Ninety six well, round bottomed microtitre plates were used in a conventional six day assay. ‘Stimulator cells’ were prepared from both patient and control skin. The epidermis was separated using dispase (2 mg/ml) and a single cell suspension prepared by trypsinisation (trypsin 0·05% and EDTA 0·02%). Dermal cells were isolated by collagenase digestion (0·5 mg/ml). Cells were resuspended in Roswell Park Memorial Institute medium containing 10% heat inactivated normal human serum with epidermal cells at 3·10^6 ml and dermal cells at 10^7 ml and irradiated with 3000 cGy. Aliquots of 100 μl of doubling dilutions of these cells were used per well in the functional assay. The four different stimulator cell populations were analysed using MAbs against CD1a and HLA-DR with a standard indirect immunofluorescence technique.

Peripheral blood mononuclear cells (PBMC) separated from venous blood from three healthy volunteers were used as the ‘responder cells’ at 3·10^5 well.
Results

PHENOTYPIC ANALYSIS

Non-involved skin showed a dense dermal infiltrate with rounded, CD1a-positive LCH cells but with no epidermal invasion. Epidermal stimulator cells bearing CD1a and HLA-DR antigens as demonstrated by indirect immunofluorescence technique

<table>
<thead>
<tr>
<th></th>
<th>CD1a</th>
<th>HLA-DR</th>
<th>Viability</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patient's normal epidermal cells (%)</td>
<td>1</td>
<td>1</td>
<td>90</td>
</tr>
<tr>
<td>Patient's lesional epidermal cells (%)</td>
<td>1</td>
<td>1</td>
<td>90</td>
</tr>
<tr>
<td>Patient's lesional dermal cells (%)</td>
<td>40</td>
<td>41</td>
<td>90</td>
</tr>
<tr>
<td>Normal control epidermal cells (%)</td>
<td>1</td>
<td>1</td>
<td>85</td>
</tr>
</tbody>
</table>

Langerhans cells, which could be differentiated from LCH cells microscopically by their dendritic nature, were identified in epidermis at normal densities. Analysis of the stimulator cell populations is shown in the table.

FUNCTIONAL ACTIVITY OF CELL POPULATIONS (FIGURE)

CD1a positive cells in lesional dermis demonstrated minimal functional activity on a per cell basis to the PBMC of all three responders. Langerhans cells from the non-involved epidermis showed similar dose related functional activity to Langerhans cells from the control skin. Langerhans cells from the epidermis overlying the involved dermis demonstrated an exaggerated functional activity against all three responder cell populations.

Discussion

Langerhans cells represent a well defined bone marrow derived cell population in the human epidermis. They constitutively express CD1a and HLA-DR and have a vital role as antigen-presenting cells in the skin. LCH is a reactive disorder in which cells bearing the Langerhans cell phenotype accumulate and/or proliferate in physiologically normal and abnormal sites. The subsequent local tissue damage may cause organ dysfunction and in severe cases, death. The in vitro functional capacity of LCH cells as antigen presenting cells has not so far been examined.

In the present study we use the allogeneic mixed cell reaction to assess the functional activity of LCH cells; this assay has been used extensively as an investigative tool for dissecting the functional activities of antigen-presenting cells. Antigen-presenting cells are believed to provide unique stimulatory signals to resting T cells in primary allogeneic mixed cell response. In vitro allogeneic mixed cell reaction is believed to represent the sensitisation and effector phases of graft rejection in vivo.

In this study we have demonstrated for the first time that, compared with normal Langerhans cells, lesional LCH cells from a patient with multisystem LCH are unable to present alloantigens to naive T cells in vitro. Although LCH cells share the same phenotype as Langerhans cells, they show defective functional activity. This discrepancy was not due to the different ways in which the epidermal and dermal cells were treated as we found that collagenase treatment does not significantly affect the function of normal Langerhans cells (unpublished data).

The Langerhans cells in normal skin from our patient showed normal functional activity. This finding suggests that the defective functional characteristic of LCH cells is localised to these cells and does not involve the entire Langerhans cell population. The significance of the increased functional activity of Langerhans cells overlying the involved dermal infiltrate is unclear but it is possible that cytokines generated in the underlying dermal infiltrate activate the overlying Langerhans cells.
The profound restrictions on work in this disease due to the difficulties in obtaining fresh viable tissues make this study unique. Further studies will concentrate on purifying LCH cells in order that the present results can be validated. The results of this study do, however, strongly suggest that LCH is a disease in which functionally abnormal Langerhans cells accumulate and/or proliferate in various tissues but the abnormality seems to be confined to the lesional LCH cells.

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**Notes**