diagnosis of the orocraniodigital syndrome, our patient has a more widespread skeletal dysplasia, with absence of thumbs, and isolated growth hormone deficiency. As he is the only child of non-consanguineous parents, no further information is gained regarding the mode of inheritance.

References


Neonatal rickets in one of identical twins

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SUMMARY We report a case of rickets in one of identical low birthweight twin infants. Plasma 25-hydroxycholecalciferol concentrations were normal in both infants; there was good clinical response to 1-a hydroxycholecalciferol. We suggest that there was a delay in maturation of renal 25-hydroxycholecalciferol-1-a hydroxylase enzyme.

The problem of rickets in the low birthweight infant has been known for some time.1 The diagnosis is made on clinical, biochemical, and radiological criteria; serial measurement of plasma alkaline phosphatase activity is useful to screen for disease.2 The aetiology of neonatal rickets is multi-factorial. We report a case of rickets in one of identical twins.

Case report

Female twins (I vertex, II breech) were born at 30 weeks' gestation after a normal pregnancy to a 23-year-old healthy well-nourished white primigravida. The membranes of twin I ruptured spontaneously 36 hours before, and those of twin II at delivery. The placenta weighed 680 g and was monochorionic and diamniotic. The blood group of both infants was ORh⁺ (mother ARh⁺) and identical antigens for the common groups, including Lutheran, Lewis, and Duffy were found. The genotype was M⁺ N⁺ S⁺ S⁻ P⁺ Lu⁺-Le⁻ (a⁻b⁻) Fy₂(a⁺b⁻) for both children.

Plasma alkaline phosphatase, calcium, and inorganic phosphorus were measured serially from age 2 weeks.3 Rickets was diagnosed in twin II at age 6 weeks (36 weeks' post-conceptional age). There was no evidence of rickets in twin I either at this age


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Childhood reference ranges:2 Plasma alkaline phosphatase upper limit 850 U/l, plasma calcium 2.20–2.67 mmol/l, plasma inorganic phosphorus 1.02–1.96 mmol/l.

Conversion: SI to traditional units - plasma calcium: 1 mmol/l = 4.0 mg/100 ml. Plasma inorganic phosphorus: 1 mmol/l = 3.0 mg/100 ml.

Figure Serial plasma alkaline phosphatase, calcium, and inorganic phosphorus in both twins.
or subsequently. Twin II had craniotabes and slowed linear growth; twin I continued to grow parallel to her centile. The clinical details and investigations done at the time of diagnosis in twin II are listed in the Table; serial biochemical data are shown in the Figure.

Both infants had similar intakes of calciferol, calcium, and phosphorus; oral feeding was established by the end of the first week of life and was initially mother's expressed breast milk and later commercial formula*. The volume of feeding in both infants was between 150 and 200 ml/kg a day after the first week.

Twin II was treated with 0.08 increasing to 0.20 μg/kg l × 25-hydroxycholecalciferol†, and no additional calcium or phosphorus was given. Longitudinal growth increased with evidence of healing on the x-ray films by age 20 weeks; treatment was stopped at 38 weeks.

Discussion

The aetiology of rickets in the neonatal period is often unknown, but has been attributed to prenatal or postnatal calciferol deficiency, calcium or phosphorus deficiency, a disturbance in the metabolic pathway of cholecalciferol, or defective mineralisation despite adequate substrate.

Although both infants came from the same antenatal nutritional environment, twin II was lighter than twin I, but with no obvious intrauterine growth retardation. The placenta was single and the blood group genotypes identical. The chance of being non-identical with these criteria alone is less than 0.03.4

There was no clinical or biochemical evidence of malabsorption, nor was there hepatic or renal disease which might affect cholecalciferol metabolism; the generalised aminoaciduria that was seen is common in preterm infants. The exchange transfusions performed on days 4 and 7 may have prevented rickets in twin I by providing additional calciferol or inorganic phosphorus; we can only speculate on this possibility. The initial plasma inorganic phosphorus concentrations were however comparable in both infants on day 14 which would argue against this. Serum copper concentration in twin II was normal making copper deficiency unlikely as a cause for the bony changes seen.

Both these infants had similar intakes of calcium, phosphorus, and calciferol before the development of rickets in twin II; a deficiency is therefore not the likely aetiology. The parathyroid hormone concentration in twin II was compatible with secondary hyperparathyroidism seen in infants (A Fairney, 1980, personal communication); we suggest that the low plasma inorganic phosphorus concentration in the affected twin was due to the phosphaturic effect of secondary hyperparathyroidism. The plasma alkaline phosphatase activity in twin I, although

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Table: Clinical information on twins I and II; same day results at time of diagnosis in twin II

<table>
<thead>
<tr>
<th>Clinical problems</th>
<th>Twin I</th>
<th>Twin II</th>
</tr>
</thead>
<tbody>
<tr>
<td>Weight (kg)</td>
<td>1.2</td>
<td>1.0</td>
</tr>
<tr>
<td>Apgar score (at one and five min)</td>
<td>5, 9</td>
<td>8, 8</td>
</tr>
<tr>
<td>(1) Hyaline membrane disease</td>
<td>positive pressure ventilation</td>
<td></td>
</tr>
<tr>
<td>(2) Hyperbilirubinaemia</td>
<td>normal liver transaminases treatment: exchange transfusion (x 2), phototherapy</td>
<td></td>
</tr>
<tr>
<td>(3) Anaemia (lowest Hb 7.8 g/dl)</td>
<td>mild haemolysis (physiological)</td>
<td></td>
</tr>
<tr>
<td>(4) Neonatal apnoea</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Results</th>
<th>Twin I</th>
<th>Twin II</th>
</tr>
</thead>
<tbody>
<tr>
<td>X-ray film</td>
<td>No rickets</td>
<td>Rickets</td>
</tr>
<tr>
<td>Plasma alkaline phosphatase (U/l)</td>
<td>1500</td>
<td>2500</td>
</tr>
<tr>
<td>Plasma alkaline phosphatase isoenzyme pattern</td>
<td>Bone</td>
<td>Bone</td>
</tr>
<tr>
<td>25-OHCC μg/l (ARR 25-43)</td>
<td>53</td>
<td>38</td>
</tr>
<tr>
<td>Parathyroid hormone ng/l (ARR 275-1200)</td>
<td>260</td>
<td>940</td>
</tr>
<tr>
<td>Copper μmol/l (RR 11-25)</td>
<td>——</td>
<td>12-9</td>
</tr>
<tr>
<td>Vitamin E μmol/l (RR 11-5-35)</td>
<td>11-6</td>
<td>4-1 (after treatment 17-5)</td>
</tr>
<tr>
<td>Generalised aminoaciduria</td>
<td>++</td>
<td>+</td>
</tr>
<tr>
<td>Viral/bacterial studies</td>
<td>——</td>
<td>——</td>
</tr>
<tr>
<td>Treatment</td>
<td>Antibiotics</td>
<td>Antibiotics, oral fat soluble vitamin E, 1-ac-hydroxycholecalciferol</td>
</tr>
</tbody>
</table>

increased above childhood reference ranges, is commonly present in the preterm infant and we do not believe that it alone represents subclinical rickets. The 25-hydroxycholecalciferol concentrations were within a normal range for both infants; assays for 1,25 and 24,25-dihydroxycholecalciferol were not available. End-organ unresponsiveness to the active 1,25 dihydroxycholecalciferol is an unlikely cause in this case because of the rapid clinical response to 1α hydroxycholecalciferol (Leo). In favour of a transient 25-hydroxycholecalciferol-1α hydroxylase enzyme deficiency in twin II is the presence of adequate initial plasma concentrations of 25-hydroxycholecalciferol, the absence of obvious renal disease, and the favourable response to 1α hydroxycholecalciferol, so that this treatment could subsequently be stopped. Although both infants should have had identical enzyme complements there may have been a difference in the timing of enzyme maturation. The only major difference between the two infants was the severity of the early clinical course in twin I and the development of rickets in twin II. This is contrary to what one might expect.

We suggest that in this infant the cause was a maturational delay in hydroxylation of 25-hydroxycholecalciferol. We do not know of any previous report of neonatal rickets in one of identical twins.

We thank Dr D Bartrop for advice, Ms Julie Patton for performing the genotype studies, and Dr Angela Fairney for the 25-hydroxycholecalciferol and parathyroid hormone assays.

References


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