Serum immunoglobulins IgG, IgM, and IgA in maternal cord blood pairs from infants of normal and low birthweights in Tanzania

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SUMMARY Serum total protein, albumin, IgG, IgM, and IgA were determined in cord blood of 54 term infants appropriate-for-gestational-age (AGA), 14 preterm AGA infants, and 21 small-for-dates infants, and in their mothers immediately after delivery in Dar-es-Salaam. The mean serum levels of total protein, albumin, and IgG in mothers who delivered a term AGA infant were 6.8 g/100 ml, 2.9 g/100 ml, and 1840 mg/100 ml respectively, whereas those from their infants were 6.9 g/100 ml, 4.1 g/100 ml, and 1471 mg/100 ml. The de novo synthesis of IgM and IgA during fetal life seems to be activated at an earlier gestational age than in infants in Western countries; this fact subsequently resulted in a higher detection rate for IgM and IgA in cord blood of term AGA infants. IgG in Tanzanian mothers was generally higher than in corresponding cord blood sera, which is contrary to the finding in Europe.

The incidence of low birthweight (<2500 g) infants in Tanzania is high (15%) compared with Western countries (6–7%). The high incidence of infections in underdeveloped countries has been related to unfavourable conditions for intrauterine growth of the fetus, although in an industrialised country such a relationship could not be shown.

Although some authors have been unable to show a relationship between malaria during pregnancy and neonatal mortality, the importance of malarial prophylaxis for the prevention of severe maternal anaemia and fetal loss (as reviewed by Kortman) has led to the widespread use of malarial prophylaxis during pregnancy in endemic areas. The question of why a healthy child is usually born despite severe placental malarial infection remains unresolved.

We wished to learn more about the development of humoral immune status (IgG, IgM, and IgA) during fetal life in relation to the immune status in the mother in an area where socioeconomic and nutritional conditions are very different from those in the West. IgM and IgA, unlike IgG, are synthesised by the fetus, which make them suitable for assessing the development of the humoral immune status of the newborn infant. In the Tanzanian population that we studied the IgG, IgM, and IgA levels were determined in maternal and cord blood of term and preterm appropriate-for-gestational-age (AGA) and small-for-dates (SFD) infants immediately after delivery. These data were compared with results from similar studies in Western countries.

Definitions and abbreviations

A term AGA infant was an infant born within 38–42 weeks of gestation with birthweight within ±1 SD from the mean for that particular gestational age, using the local standards (Fig. 1). A preterm AGA infant was an infant born before 37 completed weeks of gestation and with birthweight within ±1 SD from the mean for the gestational age, using the same local standards. An SFD infant was an infant with a birthweight <10th centile for gestational age, using the local standards.

Subjects and methods

The survey was part of a study on the interaction between malaria and pregnancy carried out between September 1976 and September 1977 at the University Hospital Muhimbili Medical Centre and the Government Maternity Ocean Road Hospital.
Details of local health facilities, population, socio-economic and geographical circumstances have been given previously. Maternal venous blood and cord blood was collected from the following groups of mothers and their offspring immediately after delivery:

**Group 1.** The control group comprised 54 randomly selected mothers and their term AGA infants. The gestational age ranged from 38 to 41 weeks.

**Group 2.** This group comprised 14 mothers and their preterm AGA infants. The gestational age ranged from 31 to 37 weeks.

**Group 3.** This group comprised 21 mothers and their SFD infants. The gestational age ranged from 37 to 40 weeks.

Any mother with hypertension or any condition complicating pregnancy—such as diabetes, urinary tract infection, multiple pregnancies, or antepartum haemorrhage—was excluded. All infants were born by normal vaginal delivery and their 1-minute Apgar score was 9 or 10. No external malformation or other obvious abnormality was found when each infant was examined. Even in the absence of clinical suspicion of intrauterine infection, such an infection cannot be positively excluded, although none of the low birthweight infants had malarial infection, or the thick smear of his peripheral blood. Gestational age of each infant was assessed within 8 hours of delivery using the physical and neurological scales of maturation suggested by Dubowitz et al. The various anthropometric and clinical data for the 3 groups of mothers and infants are summarised (Table 1). At delivery the umbilical cord was cleaned carefully before about 10 ml whole cord blood was collected. Immediately after delivery 5 ml venous blood was taken from the cubital vein of the

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**Table 1** Anthropometric and clinical data (mean ± SD) of 54 term AGA, 14 preterm AGA, and 21 SFD infants and their mothers. Number of determinations in parentheses

<table>
<thead>
<tr>
<th>Data</th>
<th>Term AGA</th>
<th>Preterm AGA</th>
<th>SFD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mothers</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>24·2±4·5</td>
<td>21·8±4·8</td>
<td>21·3±4·7</td>
</tr>
<tr>
<td></td>
<td>(52)</td>
<td>(13)</td>
<td>(19)</td>
</tr>
<tr>
<td>Parity</td>
<td>3·5±1·9</td>
<td>1·8±1·1</td>
<td>2·5±2·2</td>
</tr>
<tr>
<td></td>
<td>(51)</td>
<td>(14)</td>
<td>(21)</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>153·5±7·1</td>
<td>153·0±3·0</td>
<td>152·5±4·9</td>
</tr>
<tr>
<td></td>
<td>(50)</td>
<td>(14)</td>
<td>(19)</td>
</tr>
<tr>
<td>Arm circumference (cm)</td>
<td>25·0±2·6</td>
<td>25·2±2·3</td>
<td>24·3±1·9</td>
</tr>
<tr>
<td></td>
<td>(51)</td>
<td>(12)</td>
<td>(16)</td>
</tr>
<tr>
<td>Infants</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ratio (boys:girls)</td>
<td>26·28</td>
<td>6·8</td>
<td>10·11</td>
</tr>
<tr>
<td>Birthweight (g)</td>
<td>3036±262</td>
<td>2035±500</td>
<td>1535±476</td>
</tr>
<tr>
<td></td>
<td>(54)</td>
<td>(14)</td>
<td>(21)</td>
</tr>
<tr>
<td>Length (cm)</td>
<td>48·5±1·7</td>
<td>44·2±3·0</td>
<td>42·8±4·0</td>
</tr>
<tr>
<td></td>
<td>(54)</td>
<td>(14)</td>
<td>(21)</td>
</tr>
<tr>
<td>Head circumference (cm)</td>
<td>34·4±1·0</td>
<td>31·3±2·1</td>
<td>30·4±1·6</td>
</tr>
<tr>
<td></td>
<td>(54)</td>
<td>(14)</td>
<td>(21)</td>
</tr>
<tr>
<td>Gestational age (weeks)</td>
<td>40·1±0·6</td>
<td>35·0±1·9</td>
<td>37·7±2·1</td>
</tr>
<tr>
<td></td>
<td>(54)</td>
<td>(14)</td>
<td>(21)</td>
</tr>
</tbody>
</table>

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![Intraterine growth curves for weight, length, and head circumference for Tanzania.](image)
mother. Consent was obtained from each mother. After sampling serum was separated by centrifugation and stored and transported at –20°C until analysed. The samples were analysed within 3 months of collection. Serum protein was analysed by the Biuret reaction. IgA, IgG, and IgM levels were measured by single radial immunodiffusion, using commercial rabbit antisera to human IgG, IgM, and IgA, and their specific standards (Behringwerke). Because low concentrations for IgA and IgM were expected in cord blood, low-concentration radial immunodiffusion techniques were used. Electrophoresis of serum proteins was performed on cellulose acetate strips using the Beckman microzone electrophoresis equipment and the Beckman densitometer (R–110).

All laboratory investigations were checked by the laboratories of Sophia Children’s Hospital, Rotterdam, by the same methods. The correlation coefficient (r) of the investigations performed in Dar-es-Salaam and those in Rotterdam was 0.9 or better. Significance of differences between means were measured by Student’s t test.

Results

Maternal blood tests. Mean serum concentration (± SD) of total protein, immunoglobulins (IgG, IgM, and IgA) for mothers of term AGA, preterm AGA, and SFD infants are shown in Table 2. Total protein, IgG, IgM, and IgA showed a nearly symmetrical distribution in all three, except for IgM in mothers of SFD infants. Higher levels of total protein were found in mothers of preterm AGA and SFD infants compared with mothers of term AGA infants (P<0·001 and P<0·05 respectively). In mothers of term AGA infants 42% of the total protein concentration (6·8 g/100 ml) was albumin (2·9 g/100 ml), and 20% was γ-globulin (1·4 g/100 ml). IgG levels were slightly higher in mothers of preterm AGA than in mothers of term AGA and SFD infants. In mothers of SFD infants, higher values of IgA (P<0·01) and IgM were found than in mothers delivering term AGA infants. No noticeable changes in maternal IgG were observed during the last 8 weeks of pregnancy as calculated by multiple regression analysis (r=0·246).

Cord blood tests. Total proteins and IgG levels showed a nearly symmetrical distribution in the 3 groups of infants, whereas for IgM and IgA a nearly normal distribution was obtained after logarithmic transformation of the values. Mean serum concentration (± SD) of total protein and immunoglobulins (IgG, IgM, and IgA) in term AGA, preterm AGA, and SFD infants in cord blood are shown in Table 2. In term AGA infants the mean serum total protein was 6·9 ± 0·5 g/100 ml. This was slightly higher than that in preterm AGA or SFD infants (P<0·05). In this group of infants, albumin accounted for 60% and γ-globulins for 20% of the total protein values (4·1 ± 0·2 and 1·4 ± 0·2 g/100 ml respectively). No differences were found in the mean levels of immunoglobulins IgG, IgM, and IgA in the 3 groups of newborn babies. The IgM was >20 mg/100 ml at birth in 40% of the SFD, in 25% of the preterm AGA, and in 14% of the term AGA infants. Individual values of serum IgG, IgM, and IgA from cord blood of term AGA, preterm AGA, and SFD babies are plotted against gestational age in Fig. 2. IgM was detectable in the 31st week of gestation, whereas IgA levels were not detected until the 35th week of gestation.

In normal term infants, mean IgG in cord serum was higher in children of primiparous than of multiparous mothers (1723 v. 1327 mg/100 ml). This difference was not observed for IgA or IgM. In this group of infants IgM was detectable in all the cord sera. In 13 (27%) of the term AGA children, IgM values were 20 mg/100 ml or more. Among this group of infants, mean birthweight was slightly higher compared with mean birthweight of the whole group, but not significantly so. The mean IgA level in the term AGA infants was 5·5 mg/100 ml. In 17 (35%) infants, mean IgA level was higher than in children of multiparous mothers.

Table 2 Total protein, IgG, IgM, and IgA of maternal/cord serum pairs in 54 term AGA, 14 preterm AGA, and 21 SFD infants and their mothers (mean ± SD). Number of determinations in parentheses

<table>
<thead>
<tr>
<th>Data</th>
<th>Mothers</th>
<th>Babies</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Term AGA</td>
<td>Preterm AGA</td>
</tr>
<tr>
<td>Total protein (g/100 ml)</td>
<td>6·8±0·8 ± 0·8 ± 0·8 (49)</td>
<td>7·8±0·6 ± 0·6 (14)</td>
</tr>
<tr>
<td>IgG (mg/100 ml)</td>
<td>1840±419 (49)</td>
<td>2059±530 (14)</td>
</tr>
<tr>
<td>IgM (mg/100 ml)</td>
<td>155±31 (49)</td>
<td>180±57 (14)</td>
</tr>
<tr>
<td>IgA (mg/100 ml)</td>
<td>152±78 ± 78 ± 78 (49)</td>
<td>177±79 (14)</td>
</tr>
</tbody>
</table>

*Difference between means P<0·001, **difference between means P<0·01, ***difference between means P<0·05.

a = comparison of values between mothers of term and preterm AGA, b = comparison of values between mothers of term and SFD infants, c = comparison of values between preterm AGA and SFD infants, d = comparison of values between term and preterm AGA, e = comparison of values between term and SFD infants.

Conversion: traditional to SI units—total protein, IgG, IgM, and IgA 1 mg/100 ml =0·01 g/l.
of these infants IgA could not be detected in cord serum.

**Maternal/fetal blood ratios.** The mean concentration of IgG in the mother was 1.25 times greater than in her child in the term AGA group, 1.47 times greater in the mothers of preterm AGA, and 1.27 times greater in the mothers of SFD babies. Generally higher values of IgG in the mother corresponded to higher values in the fetus.

**Discussion**

Two assumptions were made in an attempt to relate the levels of IgG, IgM, and IgA in maternal and cord blood in Tanzania with those of other studies in Western countries. Firstly, it was assumed that the immunoglobulin levels in preterm AGA infants and their mothers reflected the normal events during pregnancy. Secondly, it was assumed that the methods used would justify comparison with other studies. Regarding the first assumption it should be remembered that the term 'preterm appropriate-for-gestational-age' is in a sense paradoxical, as events leading to or accompanying preterm birth must be 'abnormal' and may well affect the relative proportions of some components in the mother or her fetus. However the same criteria were used by studies in Western countries. The last assumption is probably valid since the low concentration radial immunodiffusion techniques were used for the IgA and IgM determinations in cord blood, with a detection limit of 1 mg/100 ml for IgM.

In this study, among mothers delivering a term AGA infant, the mean value for total protein (6.8 g/100 ml) and albumin (2.9 g/100 ml) was higher than found in white women. Our range of means was 5.9–6.4 for total protein and 2.4–2.75 g/100 ml for albumin. Somewhat higher values for total protein (7.2–8.0 g/100 ml) were found in well-nourished African, Hindu, and Bantu women towards the end of pregnancy. Mean IgG values at the end of normal pregnancy (1840 mg/100 ml) were higher than is normal for white women (1000–1400 mg as reviewed by de Mural18), whereas IgA and IgM levels were similar. This was found also in another study in Africa. The reason for the higher IgM and IgA levels among mothers of SFD infants remains unclear, although it might be explained by a higher incidence of (subclinical) infections.

In the group of Tanzanian term AGA infants we studied, mean total protein level was higher than that reported in infants from Western countries (average 5.8 g/100 ml, Schultze and Heremans), which may partly be explained by the higher albumin levels, 4.1 v. 3.5–4.0 g/100 ml, in Western term infants.

When the individual values in the three groups of Tanzanian infants in relation to their gestational age (Fig. 1) were compared with studies in the USA the de novo synthesis of IgM and IgA by the fetus seems to be activated at an earlier gestational age. This assumption is further substantiated by the higher incidence of detectable IgM (100%) and especially of IgA levels (65%) in the term AGA infant at birth compared with the infant in Western countries (75% for IgM, and 0–30% for IgA as reviewed by de Mural18).

Since the fetus synthesises most of its proteins, with the exception of IgG, from amino-acids which have been transferred across the placenta it can be assumed that the rate of synthesis of albumin, IgM, and IgA by the Tanzania fetus is greater than that found in Western countries during the last stage of pregnancy. The high endogenous synthesis of IgM and IgA in the fetus might be explained by increased
induction from antigenic stimuli in the maternal environment, although a genetic origin cannot be excluded. It was beyond the scope of this study to evaluate the incidence of intrauterine infection among the infants, although it can be said that none of the infants in the low birthweight groups, and only one infant in the term AGA group had malarial parasites in the cord blood smear at birth. In this particular infant, IgM level was 20 mg/100 ml; the child was clinically well and went home without any treatment: at the postnatal follow-up no abnormalities were noted. Among the preterm AGA infants all were discharged home in good condition, except one with a gestational age of 31 weeks who died at age 3 weeks, severely dehydrated. In the SFD group all were discharged home in good condition.

No attempts were made to evaluate the cellular immunity in this community of mothers and infants.

In conclusion, in paired Tanzanian maternal/fetal blood, generally higher values were detected for total protein, albumin, and IgG in maternal blood at the end of pregnancy, compared with Western standards. The concentration of total protein, albumin, and IgG in the fetal circulation was also increased compared with the same standards, with a higher detection rate for IgM and IgA, which could be explained by an increased induction of the fetal humoral antibody synthesis from antigenic stimuli in the maternal environment. Contrary to findings in Western countries, IgG in the maternal sera tended to be higher than in the corresponding fetal circulation.

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