Effect of early oral calcium supplementation on serum calcium and immunoreactive calcitonin concentration in preterm infants

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SUMMARY Oral calcium supplements (80 mg/kg per 24 h) were given to 23 preterm infants, and the course of serum calcium, magnesium, immunoreactive calcitonin, and gastrin was compared with a control group of 23 matched infants. In the supplemented group, serum calcium concentrations remained at the baseline level (2·31 mmol/l ± 0·18 SD) while a fall (from 2·27 ± 0·18 to 1·91 ± 0·24 mmol/l) was observed at 12–16 hours of age in the control group, with 4 values <1·75 mmol/l. There was no change in serum magnesium concentration in either group. The postnatal rise of serum immunoreactive calcitonin concentrations in the control group (from 171 ± 135 to 493 ± 273 pg/ml at 12–48 hours of age) was not found in the supplemented group. There was a negative correlation between serum calcium and immunoreactive calcitonin levels in the control group, but not in the supplemented group. There was no correlation between serum immunoreactive calcitonin and gastrin concentrations. These data show that oral calcium supplementation can prevent early neonatal hypocalcaemia, and suggest that this effect is achieved at least in part through a reduction of the postnatal rise of serum immunoreactive calcitonin.

After birth there is a fall in serum calcium (Ca) and a rise in serum immunoreactive calcitonin (iCT) levels. Preterm infants are particularly at risk from early hypocalcaemia. In order to compensate for the interruption of the large maternal supply, early supplements of oral calcium were given to preterm neonates. Its effect on serum Ca, magnesium (Mg), iCT, and immunoreactive gastrin (iGT) concentrations was compared with a control group. Our aim was to find out whether hypocalcaemia can be prevented in preterm neonates, and if it can, how early.

Patients and methods

Subjects. The study was carried out between February and September 1977 on 46 preterm infants. The study was approved by the ethical committee of the hospital.

Gestational age was assessed by the score of Dubowitz et al. Birthweight was between the 25th and 75th centiles. Any infant with an Apgar score <7 at one minute was excluded. On admission to the unit (between 2 and 8 hours of age) each infant was placed randomly in one of two groups: a group of 23 infants supplemented with calcium (80 mg/kg per 24 hours for 5 days) or a group of 23 control infants. The calcium was given as 10% calcium gluconate in water (osmolality: 297 mosmol/kg H2O), divided into between 6 and 8 doses a day and added to milk feeds.

Feeds were started at age 6–9 hours, either with breast milk collected from a milk bank (13 infants in the supplemented group and 10 infants in the control group) or by a 'humanised' formula. Mean calcium concentration was 24·5 mg/100 ml for breast milk and 57 mg/100 ml for the formula; mean Ca/P ratio was 1·9 for breast milk and 1·6 for the formula.

Measurements. For serum Ca and Mg determinations blood was collected at regular intervals by heel prick before feeds: 2–8, 12–16, 18–24, 44–52, 62–76, and 92–100 hours of age. Serum Ca and Mg were measured by atomic absorption (IL 343).

For serum iCT and iGT determinations, again blood samples were collected in microtubes by heel prick at admission (that is, 2–8 hours of age) and then at ages 12–16, or 18–24 hours. Blood collections were performed sequentially in 11 infants of the supplemented group and 13 of the control group. After centrifugation at 4°C sera were stored at −28°C. Samples were analysed in the same order to...
avoid interassay variation. Serum iCT was measured by the method of Tashjian,12 and David et al.,6 the lower limit for the assay being 150 pg/ml, with serum iCT undetectable in normal children and adults. For practical reasons, levels of serum iCT <150 pg/ml were given a value of 75 when calculating the mean. Serum iGT was measured according to the technique of Ganguli and Hunter,13 and Sann et al.,14 with a sensitivity of 15 pg/ml.

Basal levels of serum Ca were analysed by Student's t test and the levels of serum iCT by χ2 test. The postnatal course of serum Ca and iGT and iCT were analysed by paired t tests.

Results

Clinical data. The main clinical data are shown in Table 1. There was no difference in gestational age, birthweight, or the volume of milk intake between the two groups.

Tolerance of calcium supplementation. The daily number (mean ± 1 SD) of regurgitations was 1.6 ± 0.48 in the control group and 2.0 ± 0.44 in the supplemented group. The daily number of stools was 4.39 ± 0.26 in the control group and 5.65 ± 0.25 in the supplemented group. The maximum loss of weight since birth was 153 ± 20 g in the control group and 147 ± 21 g in the supplemented group. None of these differences between the two groups was significant. The later development of all infants was normal.

Postnatal course of serum Ca and Mg. This is shown on Fig. 1. The baseline Ca levels were similar in both groups. In the control group there was a decrease from (mean ± 1 SD) 2.27 ± 0.18 to 1.91 ± 0.24 mmol/l at age 12–16 hours (P<0.001). However, there was only a minor and nonsignificant decrease in the supplemented group, from 2.31 ± 0.18 to 2.29 ± 0.14 mmol/l. Serum Ca remained significantly lower in the control group than in the supplemented group until 92–100 hours of life. The incidence of hypocalcaemia (that is serum Ca <1.75 mmol/l) was 4 of 23 in the control group and none of 23 in the supplemented group. In the supplemented group 2 infants exhibited hypercalcaemia on one occasion, 2.92 and 2.82 mmol/l. All other values were <2.75 mmol/l. 24 hours after stopping calcium supplementation, serum Ca levels (2.39 ± 0.16 mmol/l) were not significantly different from those of the previous day, 2.32 ± 0.18 mmol/l.

Serum Mg was similar in both groups for the baseline levels, 0.84 ± 0.07 mmol/l. No significant difference was observed at 12–24 hours (0.82 ± 0.01 in the supplemented group v. 0.81 ± 0.07 mmol/l in the control group) or later.

Postnatal course of serum iCT and iGT concentrations. The baseline levels for iCT (Fig. 2) (mean ± 1 SD) were similar in the control group (161 ± 125 pg/ml) and the supplemented group (186 ± 155 pg/ml) (χ2 0.621, NS). At 12 to 16 hours there was only one undetectable value in the control group, the detectable values being 553 ± 232 pg/ml; in the supplemented group there was also one undetectable value, the detectable values being 254 ± 67 pg/ml. The difference between the two groups was highly significant P<0.001. At age 18 to 24 hours, there was still only one undetectable value in each group: the detectable values were 505 ± 282 pg/ml in the control group and 372 ± 145 pg/ml in the supplemented group. Despite the higher level of the mean concentration in the control group, the difference was not significant. The course of individual serum iCT concentrations is shown in Fig. 3. In the control group, serum iCT levels increased from 171 ± 135 to

Table 1

<table>
<thead>
<tr>
<th>Clinical data</th>
<th>Calcium supplemented</th>
<th>Control</th>
</tr>
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<tbody>
<tr>
<td>(n=23)</td>
<td>(n=23)</td>
<td></td>
</tr>
<tr>
<td>Gestational age (weeks)</td>
<td>35.0 ± 0.35*</td>
<td>34.6 ± 0.44</td>
</tr>
<tr>
<td>Birthweight (g)</td>
<td>(32-37)</td>
<td>(32-37)</td>
</tr>
<tr>
<td>Daily milk intake (ml/kg)</td>
<td>1992 ± 72</td>
<td>1985 ± 70</td>
</tr>
<tr>
<td>Daily calcium intake (mg/kg)</td>
<td>(1400-2450)</td>
<td>(1510-2650)</td>
</tr>
<tr>
<td>Days 1 to 5</td>
<td>46 ± 2.5 → 109 ± 2.1</td>
<td>51 ± 5 → 112 ± 3</td>
</tr>
<tr>
<td>Daily calcium intake (mg/kg)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Days 1 to 5</td>
<td>86 ± 3 → 122 ± 5</td>
<td>19 ± 3 → 45 ± 4</td>
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*Mean ± SD, ranges in parentheses.
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493 ± 273 pg/ml at 12 to 24 hours of age (paired t test, P<0.01); in the supplemented group, there was a slight but nonsignificant increase from 188 ± 144 to 303 ± 164 pg/ml. Between 12 and 24 hours, there was no significant difference in iCT levels when the infants were fed breast milk or formula.

The course of serum iCT is shown on Table 2. Baseline levels were similar in the control group and the supplemented group. In the control group there was an increase at 12-16 and at 18-24 hours (P<0.05). In the supplemented group however, the values at 12-16 and at 18-24 hours were not significantly different from the baseline levels or from the values in the control group.

Correlation analysis. At the nadir of serum Ca concentrations in the control group, there was a highly significant negative correlation between serum Ca and iCT concentrations (r = -0.849; P<0.001). In the supplemented group, no correlation was observed between serum Ca and iCT concentration at any time. There was no correlation between serum iCT and iGT levels or between serum Ca and iGT concentrations in either group.

<table>
<thead>
<tr>
<th>Postnatal age (hours)</th>
<th>Supplemented group</th>
<th>Control group</th>
</tr>
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<tbody>
<tr>
<td>2-8</td>
<td>60 ± 28</td>
<td>63 ± 20</td>
</tr>
<tr>
<td>12-16</td>
<td>93 ± 55</td>
<td>124 ± 74*</td>
</tr>
<tr>
<td>18-24</td>
<td>84 ± 31</td>
<td>132 ± 87*</td>
</tr>
</tbody>
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*Significant (P<0.05) difference from the baseline level.
Discussion

During the transition from fetal to neonatal life, the preterm infant is removed from an environment where he received a net transplacental influx of 130 to 150 mg/kg per 24 hours of calcium. This removal results in a rapid fall in serum Ca which sometimes leads to hypocalcaemic convulsions. These data show that when these infants are supplemented with a large amount of calcium, the early fall in serum Ca can be prevented. This confirms the preliminary report of Brown et al. Recently, Moya and Doménech also described the prevention of early neonatal hypocalcaemia with a formula supplemented with calcium. They suggested that the preterm neonate, like the fetus, is dependent on a large supply of calcium in order to maintain a normal serum Ca. Our data also suggest that such large supplies of calcium can be achieved by the oral route; this is in agreement with the results of Sutton et al. who showed that calcium given as a food supplement is almost completely absorbed by preterm infants.

Oral calcium supplements were well tolerated. Although there was a tendency for these infants to have a greater frequency of bowel movements than the control group, the difference was not significant. This is in contrast with the results of Brown et al. and attributable to the use of pure calcium gluconate instead of gluconate in syrup. Willis et al. recorded an increased incidence of necrotising enterocolitis in preterm infants with oral calcium supplements, but not if the calcium was given mixed with food. The only serious side effect in our study was the occurrence of transient hypercalcaemia in 2 of the supplemented infants, which disappeared when the supplements were stopped. Thus we favour oral calcium supplementation to prevent early hypocalcaemia in preterm neonates, provided that calcium is mixed with the foods and that serum Ca levels are regularly monitored.

Our results from the control group confirm previous work showing that hypercalcitonaemia participates in the occurrence of early hypocalcaemia. By contrast, the postnatal rise in serum iCT was limited in the supplemented group, and there was no correlation between serum Ca and iCT concentrations. It is unlikely that phosphorous or parathyroid hormone is involved since Brown et al. found no effect of oral calcium on serum levels; nor was there any change in plasma 25-hydroxycholecalciferol levels. The fall in serum iCT levels may therefore be attributed to calcium gluconate administration.

The effect of calcium gluconate was surprising, since in adults intravenous or oral administration of calcium gluconate stimulates the secretion of calcitonin. In baby rats, Cooper et al. observed a rise in serum iCT in response to postprandial hypercalcaemia. However, in human preterm neonates Salle et al. could detect no change in the postnatal rise of serum iCT during continuous infusion of calcium gluconate, despite an increase in serum Ca. Our data thus suggest that the effect of oral calcium gluconate on serum iCT concentrations may be attributed to a digestive factor. According to some reports, gastrin is a potent calcitonin secretagogue in man although this has not been confirmed by others. In the present study, the postnatal course of serum iCT and iGT was the same in both groups, but no correlation was observed. Our findings agree with the results of Cooper et al. in newborn rats. We conclude that the effect of oral calcium gluconate on serum iCT may be attributed to other digestive factors.

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