Comparison of hexachlorophane and chlorhexidine powders in prevention of neonatal infection

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SUMMARY The protective effect of treating the skin of newborn infants with powders containing 1% chlorhexidine or 0.33% hexachlorophane was compared. Each was equally effective in preventing colonisation and infection by Staphylococcus aureus. In contrast, the skin became profusely colonised by coagulase-negative staphylococci, irrespective of the powder used. Venous blood concentrations of chlorhexidine were low or undetectable in the few infants whose blood was analysed.

The value of topical chemoprophylaxis with hexachlorophane against staphylococcal colonisation and infection in the newborn is well known.1–4 The fact that some hexachlorophane is absorbed through the skin5–7 led to a withdrawal of hexachlorophane prophylaxis in many hospitals. This withdrawal was often followed by a resurgence of staphylococcal sepsis, thus confirming the value of the chemoprophylaxis.8–13

Toxic effects have been caused by absorption of hexachlorophane but only if used in excessive quantities14 or if applied to skin that had been damaged by burns or disease.15–17 No side effects have been reported in more then 200 000 infants since a talcum powder containing 0.33% hexachlorophane was introduced in Bristol 20 years ago,1 nor have side effects been reported in the many infants so treated elsewhere. The blood concentration of hexachlorophane in powder-treated infants is very low, greatly below the level found to be toxic in animals.7,16 Similarly, Plueckhahn et al.17 failed to find evidence of damage in very large numbers of infants treated with a hexachlorophane emulsion. On this evidence it would be unwise to stop using hexachlorophane as it gives considerable protection against staphylococcal infection until an equally effective alternative can be found that is still safer.

Materials and methods

Infants of at least 37 weeks’ gestation and born by vaginal delivery were randomly selected from two lying-in wards of this hospital. All infants were white, of normal birthweights, and remained in the ward for at least 7 days. At birth, the cord was cut at 4 cm from the skin, ligated with a rubber band, and sprayed with a chlorhexidine preparation (Hibispray No. 1, ICI Ltd). The management of all infants was identical, except for the disinfectant powder used. One group was treated with a talcum powder containing 1% chlorhexidine (Hibitane, ICI Ltd) and the other with 0.33% hexachlorophane powder (Ster-zac, Hough, Hoseason and Co. Ltd) already in routine use in this hospital. Both contain 3% zinc oxide. There were no differences in gestational ages, sex, duration of labour, or birthweights of infants in either group. There was no untreated group of babies as it was not considered justifiable to withhold antistaphylococcal prophylaxis. Each infant was dusted liberally with powder all over the trunk, once daily; in addition the umbilical and perineal areas were powdered at each napkin change after wiping the umbilical cord with alcohol.

Swabs were taken from the umbilicus, perineum, nose, and axillae on the day of birth and on two days subsequently—on day 3, 5, or 7. These were placed immediately in Stuart’s transport medium to which was added disinfectant neutralisers (2% Tween 80 and 0.3% Azoletcin). Swabs were inoculated on to blood agar containing 10% neutraliser, and on to MacConkey's agar and Sabouraud's agar using a serial dilution technique.
Plates were incubated aerobically for 48 hours at 37°C. Clinical signs of infection were recorded, particularly umbilical or eye infections, and septic lesions of the skin or paronychiae. Swabs from all lesions were cultured. Skin infections other than staphylococci, such as erythema and oedema, were also noted.

Heparinised venous blood samples were taken from each infant when venous blood was required for other clinical investigations. Venepuncture was justified for experimental purposes only; capillary blood samples were not taken since contamination with chlorhexidine on the skin could not have been excluded. The blood samples were stored at −20°C and sent to the Huntingdon Research Centre, Cambridgeshire, for estimation of chlorhexidine by gas–liquid chromatography.18

Results

A total of 189 infants entered the trial but 15 who left hospital before the 7th day were withdrawn. Of those remaining, 87 were treated with hexachlorophane and 87 with chlorhexidine powder.

The aerobic microbial flora of nose, axillae, perineum, and umbilicus is shown in Table 1. Staphylococcus aureus was rarely isolated and there was no difference between the two groups in colonisation by this organism. Staphylococcus aureus colonisation of the nose was usually greater than at other sites and increased with age (Fig. 1). Only 3 proved staphylococcal infections, paronychiae, or eye infections were found in each group.

However, colonisation by coagulase-negative staphylococci was heavy, 70–90% for all sites from the day of birth, irrespective of the powder (Fig. 1). Other organisms that were isolated are shown in Table 1. The only difference between the effects of the two treatments was in the lower colonisation by Escherichia coli of the umbilicus and perineum in the chlorhexidine-treated group (Fig. 2). However, no infection was caused by any organism other than S. aureus.

The incidence of skin reactions was very low in both groups. One infant in the chlorhexidine-treated group developed a slight transient erythema and 6 infants in each group had skin reactions unrelated to the powders—such as napkin rash and neonatal urticaria.

The separation time of the umbilical cord did not significantly differ between the chlorhexidine- and hexachlorophane-treated groups (Table 2).

Fig. 1 Colonisation of infants by Staphylococcus aureus and coagulase-negative staphylococci.

| Table 1 Microbial flora: percentage of infants colonised at any sampling time during the study |
|---------------------------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|
| Organism                        | Percentage of infants colonised |
|                                 | Nose (Chlorhexidine)           | Umbilicus (Chlorhexidine)       | Perineum (Chlorhexidine)        | Axillae (Chlorhexidine)         | Nose (Hexachlorophane)           | Umbilicus (Hexachlorophane)       |
| S. aureus                       | 21·2                           | 34·0                           | 2·4                            | 22·2                           | 4·8                             | 5·5                             |
| Coagulase-negative staphylococci| 92·0                           | 87·0                           | 77·5                           | 83·0                           | 92·0                           | 88·0                           |
| Escherichia coli                | 24·0                           | 19·2                           | 42·0                           | 56·0                           | 62·5                           | 75·0                           |
| Other Gram-negatives            | 5·4                            | 2·0                            | 9·4                            | 10·1                           | 12·0                           | 18·2                           |
| Streptococcus faecalis          | 22·6                           | 13·8                           | 12·3                           | 11·7                           | 15·5                           | 20·8                           |
| Candida albicans                | 0                              | 1·2                            | 4·9                            | 2·0                            | 4·7                            | 0                              |
| Others                          | 1·2                            | 0                              | 2·4                            | 0                              | 0                              | 4·6                            |

**Fig. 1 Colonisation of infants by Staphylococcus aureus and coagulase-negative staphylococci.**
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**Fig. 2** Colonisation of infants by *Escherichia coli.*

The concentration of chlorhexidine was determined in the blood of 12 infants (Table 3). The level was slightly raised above the limit of detection of the assay method in two (Cases 2 and 12). One of these was in the hexachlorophane-treated group, and it may be significant that his mother was the only one to have had chlorhexidine solution instilled into her bladder before delivery. Hibitane cream is used for vaginal examinations and the mothers of the two infants with significant chlorhexidine levels had had more examinations than the others.

**Discussion**

The use of chlorhexidine or hexachlorophane dusting powder in this study was associated with equally low levels of colonisation and infection by *S. aureus*. In contrast, colonisation of all sites by coagulase-negative staphylococci was strikingly high from the first day of life, irrespective of the disinfectant. This consistent and tenacious colonisation may represent a protective role of the normal skin flora. The only significant differences in bacteriological findings between the two powders was the lower colonisation of the umbilicus by *E. coli* in the chlorhexidine-treated group. Gillespie and colleagues observed delayed umbilical separation time using 0.2% chlorhexidine in talcum without zinc oxide, but in the present study we observed no difference between infants treated with chlorhexidine and hexachlorophane powders, both of which contain 3% zinc oxide. The zinc oxide, with its astringent properties, is probably more important than the disinfectant in reducing cord separation time.

Plueckhahn et al. expressed concern that the widespread use of 4% chlorhexidine gluconate would increase the incidence of species of *Pseudomonas* and *Proteus* with associated problems of cross infection. However, using a lower concentration of chlorhexidine (1%) we found no increase in colonisation by these organisms.

The very low blood levels found in the few infants from whom samples were obtained are in line with the results of Cowen et al. who studied percutaneous

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**Table 3** Blood chlorhexidine concentrations in infants treated with hexachlorophane or chlorhexidine powder

<table>
<thead>
<tr>
<th>Case</th>
<th>Age (days)</th>
<th>Blood-chlorhexidine (mg/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Hexachlorophane</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>5</td>
<td>None detected</td>
</tr>
<tr>
<td>2</td>
<td>6</td>
<td>0.146*</td>
</tr>
<tr>
<td>3</td>
<td>9</td>
<td>0.062</td>
</tr>
<tr>
<td>4</td>
<td>11</td>
<td>0.047</td>
</tr>
<tr>
<td>5</td>
<td>13</td>
<td>0.053</td>
</tr>
<tr>
<td><strong>Chlorhexidine</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>3</td>
<td>None detected</td>
</tr>
<tr>
<td>7</td>
<td>6</td>
<td>0.032</td>
</tr>
<tr>
<td>8</td>
<td>6</td>
<td>0.017</td>
</tr>
<tr>
<td>9</td>
<td>6</td>
<td>0.034</td>
</tr>
<tr>
<td>10</td>
<td>7</td>
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</tr>
<tr>
<td>11</td>
<td>7</td>
<td>None detected</td>
</tr>
<tr>
<td>12</td>
<td>10</td>
<td>0.098*</td>
</tr>
</tbody>
</table>

*Values significantly different from the background peak heights at 95% confidence level.
absorption in infants bathed with chlorhexidine solution. Case et al. failed to demonstrate percutaneous absorption from handwashing in 5% aqueous chlorhexidine solution. Chow et al. in studies on rats found that less chlorhexidine was absorbed than hexachlorophane under similar conditions.

Even when chlorhexidine is absorbed, its toxicity appears to be low. The LD50 values for chlorhexidine gluconate administered to laboratory animals by oral, subcutaneous, or intravenous routes are very high. There is a lack of information on toxic effects caused by chlorhexidine in man, and we can find no evidence of systemic toxicity from topical use.

We believe that a topical disinfectant should be used to prevent staphylococcal infection of infants in maternity hospitals. Doctors who prefer to use the disinfectant in powder rather than in liquid form may choose either 1% chlorhexidine or 0·3% hexachlorophane in presterilised talcum powder and apply it to the whole body as well as to the umbilicus. The choice will depend on the risks of toxic side effects. Hexachlorophane powder has been consistently effective for more than 20 years and has caused no harm when correctly used. Blood levels in mature infants after 8 days' treatment were well below presumed toxic levels based on animal experiments. Pluckhahn et al. found similarly low blood levels in infants treated with hexachlorophane either as 0·75% emulsion or 0·5% in powder. However, there have been serious consequences when hexachlorophane was grossly misused or wrongly formulated.

Chlorhexidine was as effective as hexachlorophane and since evidence suggests that it has a wider margin of safety than hexachlorophane, many will probably prefer it. Chlorhexidine powder should soon be available commercially for use by those who prefer to avoid hexachlorophane.

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References

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