Viral haemorrhagic fever in children

Nathalie E MacDermott,1 Surjo De,2 Jethro A Herberg1

ABSTRACT

Viral haemorrhagic fevers (VHFs) are currently at the forefront of the world’s attention due to the recent Zaire ebolavirus epidemic in West Africa. This epidemic has highlighted the frailty of the world’s public health response mechanisms and demonstrated the potential risks to nations around the world of imported cases of epidemic diseases. While imported cases in children are less likely, the potential for such a scenario remains. It is therefore essential that paediatricians are aware of and prepared for potential imported cases of tropical diseases, VHFs being of particular importance due to their propensity to cause nosocomial spread. Examining the four families of viruses—Filoviridae, Arenaviridae, Bunyaviridae and Flaviviridae, we describe the different types of VHFs, with emphasis on differentiation from other diseases through detailed history-taking, their presentation and management from a paediatric perspective.

The 2014–2015 Ebola virus disease (EVD) outbreak has been the largest epidemic of a filoviral haemorrhagic fever (VHF) on record.1 It threatened the stability of three West African nations and, several months into the epidemic, the potential implications of imported cases led to urgent contingency planning worldwide. As of November 2015, there had been 27 imported cases. This epidemic has highlighted the global vulnerability of an increasingly interconnected world to public health threats, and the requirement for all nations to maintain readiness for future imported cases of highly pathogenic, transmissible diseases.

Approximately 20% of EVD cases in the current epidemic were children under the age of 15 years,2 emphasising the need for vigilance for imported childhood cases. While Ebola has dominated the headlines for many months and the Zaire ebolavirus is one of the more lethal species of VHF viruses,3 healthcare professionals should be aware of several other VHF diseases, as early management in a controlled and safe healthcare environment improves survival of patients, protects healthcare workers and reduces potential for community spread.

PATHOGENS CAUSING VHF IN CHILDREN

Pathogens causing VHF encompass a broad spectrum of viruses causing a severe multisystem syndrome with vascular dysregulation and damage, which may result in overt haemorrhage.4 The viruses most commonly classified as causing VHF are listed in table 1,3,4–24 and include species from four families (Arenaviridae, Filoviridae, Bunyaviridae and Flaviviridae). This article will focus on those classified as hazard group 4 pathogens by the UK Advisory Committee on Dangerous Pathogens (ACDP), owing to their potential for spread within the healthcare setting and their significant public health implications. These are pathogens that, for the safety of healthcare workers and the general public, are the most important to be recognised.

CLINICAL PRESENTATION OF VHF IN CHILDREN

VHF diseases are caused by several unrelated viruses, and they cause a range of clinical symptoms. The best characterised paediatric data are for EVD (table 2).2,4–25 Each VHF virus is associated with a stronger likelihood of particular clusters of symptoms, emphasised in table 1, but any feature can occur with any VHF virus. Percentage figures shown in table 2 reflect the occurrence of these symptoms in children under 15 years of age during the current Ebola virus epidemic in West Africa.6 Percentage figures for symptom incidence in children with non-Ebola VHF’s are largely unavailable and where available are limited by small sample sizes.

The broad, non-specific nature of most symptoms makes careful travel and epidemiological history-taking essential for prompt recognition. Key questions to include in the history are shown in box 1,26 some of which are location-specific. Boxes 2 and 3 give case history examples emphasising the broad nature of presentations of VHF. Clinicians must bear in mind the potential for other non-VHF, but potentially life-threatening, diseases that may present with haemorrhagic symptoms, and should consider these within the differential diagnosis and arrange appropriate investigation when investigating any suspected cases of VHF.

Filoviridae

Ebola virus disease

Ebola viruses are a genus in the filovirus family. They were first identified during an outbreak of haemorrhagic fever in the Democratic Republic of Congo (DRC, then Zaire) in 1976, and have subsequently been responsible for approximately 21 further epidemics of haemorrhagic fever affecting humans in DRC, Uganda, Sudan and Gabon.3 The most devastating epidemic to date is the recent epidemic centred in Guinea, Liberia and Sierra Leone, which has affected over 28 000 people with more than 11 000 deaths.26 See figure 1 for regions of the world known to be at risk from Ebola viruses.

Zaire, Sudan and Bundibugyo ebolaviruses are responsible for all human epidemics. Zaire ebolavirus causes the most lethal disease, with mortality rates of 40–90%, while Sudan and Bundibugyo ebolaviruses have mortality rates of approximately 40–70% and 25–50%, respectively.3 It remains unclear how Ebola viruses enter human transmission, although fruit bats may act as a reservoir for

Table 1 Features of the viral haemorrhagic fevers

<table>
<thead>
<tr>
<th>Virus</th>
<th>Disease</th>
<th>Geography</th>
<th>Human infection source</th>
<th>Incubation (days)</th>
<th>Treatment</th>
<th>Clinical precautions</th>
<th>Common clinical features and case fatality rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Family: Filoviridae; Genera: Ebola virus, Marburgvirus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Zaire ebolavirus, Sudan ebolavirus, Bundibugyo ebolavirus, Tai Forest ebolavirus</td>
<td>Ebola virus disease</td>
<td>Africa (central, western and eastern)</td>
<td>Fruit bats, infected non-human primates, human-to-human, nosocomial, sexual transmission</td>
<td>2–21</td>
<td>Supportive and antibody based therapies</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, weakness, headache, profuse diarrhoea, vomiting, conjunctival injection, hiccoughs, haemorrhage CFR=25–90%</td>
</tr>
<tr>
<td>Marburg marburgvirus</td>
<td>Marburg HF</td>
<td>Africa (central, eastern and southern)</td>
<td>Fruit bats, infected non-human primates and forest antelope, human-to-human, nosocomial</td>
<td>2–21</td>
<td>Supportive</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, weakness, headache, myalgia, vomiting, diarrhoea, conjunctival injection, haemorrhage CFR=24–88%</td>
</tr>
<tr>
<td>Family: Arenaviridae; Genus: Mamarenavirus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lassa mammarenavirus</td>
<td>Lassa fever</td>
<td>West and central Africa</td>
<td>Rodents, human-to-human, nosocomial, sexual transmission</td>
<td>6–21</td>
<td>Supportive and ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, sore throat, retrosternal pain, myalgia, severe cases—gross oedema, haemorrhage, deafness CFR=Approximately 15% (hospitalised patients)</td>
</tr>
<tr>
<td>Junin mammarenavirus</td>
<td>Argentine HF</td>
<td>Argentina</td>
<td>Rodents, human-to-human, nosocomial</td>
<td>5–19</td>
<td>Supportive and convalescent plasma</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, headache, myalgia, vomiting, retro-orbital pain, haemorrhage, irritability, ataxia CFR=up to 30%</td>
</tr>
<tr>
<td>Machupio mammarenavirus</td>
<td>Bolivian HF</td>
<td>Bolivia</td>
<td>Rodents, human-to-human (rare), nosocomial (rare)</td>
<td>5–19</td>
<td>Supportive and ? ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, headache, myalgia, vomiting, retro-orbital pain, haemorrhage, irritability, ataxia CFR=up to 30%</td>
</tr>
<tr>
<td>Guanarito mammarenavirus</td>
<td>Venezuelan HF</td>
<td>Venezuela</td>
<td>Rodents, human-to-human, nosocomial</td>
<td>7–14</td>
<td>Supportive and ? ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, headache, sore throat, arthralgia/myalgia, diarrhoea, conjunctivitis, haemorrhage CFR=up to 30%</td>
</tr>
<tr>
<td>Sabia mammarenavirus</td>
<td>Brazilian HF</td>
<td>Brazil</td>
<td>Rodents, human-to-human, nosocomial</td>
<td>7–14</td>
<td>Supportive and ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, headache, myalgia, vomiting, retro-orbital pain, haemorrhage, jaundice, hepatic necrosis CFR=up to 30%</td>
</tr>
<tr>
<td>Chapare mammarenavirus</td>
<td>Chapare HF</td>
<td>Bolivia</td>
<td>Rodents, human-to-human, nosocomial</td>
<td>7–14</td>
<td>Supportive and ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Unclear, little known, similar clinical presentation to Bolivian HF</td>
</tr>
<tr>
<td>Lujo mammarenavirus</td>
<td>Lujo</td>
<td>Zambia</td>
<td>Rodents, human-to-human, nosocomial</td>
<td>9–13</td>
<td>Supportive and ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, myalgia, headache, vomiting, diarrhoea, bleeding (nose, gums) CFR=30% (15–70%), approximately 5% in children &lt;18 years</td>
</tr>
<tr>
<td>Family: Bunyaviridae; Genera: Nairovirus, Phlebovirus, Hantavirus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crimean-Congo hemorhagic fever virus</td>
<td>Crimean-Congo HF</td>
<td>Africa, Balkans, Russia, Middle East, central Asia, western China</td>
<td>Ticks, livestock animals (cattle, sheep, goats), ostriches, human-to-human, nosocomial</td>
<td>1–13</td>
<td>Supportive and ribavirin</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, malaise, headache, vomiting, diarrhoea, bleeding (nose, gums) CFR=30% (15–70%), approximately 5% in children &lt;18 years</td>
</tr>
</tbody>
</table>

Continued
<table>
<thead>
<tr>
<th>Virus</th>
<th>Disease</th>
<th>Geography</th>
<th>Human infection source</th>
<th>Incubation (days)</th>
<th>Treatment</th>
<th>Clinical precautions</th>
<th>Common clinical features and case fatality rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Severe fever with</td>
<td>Severe fever with</td>
<td>China, Japan, South Korea and</td>
<td>Ticks, human-to-human, nosocomial</td>
<td>5–15</td>
<td>Supportive</td>
<td>Isolation of patient, Full PPE for healthcare workers</td>
<td>Fever, thrombocytopenia, leukopenia, myalgia, vomiting, diarrhoea, headache with evolving disseminated intravascular coagulation, haemorrhage and multiorgan failure. CFR=8%</td>
</tr>
<tr>
<td>thrombocytopenia</td>
<td>thrombocytopenia syndrome</td>
<td>North Korea</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Omsk hemorrhagic</td>
<td>Omsk HF</td>
<td>Russian federation</td>
<td>Ticks, infected muskrats</td>
<td>3–14</td>
<td>Supportive</td>
<td>No known interhuman spread—Standard IP&amp;C measures</td>
<td>Fever, myalgia, headache, rash, haemorrhage CFR≤1% (up to 10% for severe (haemorrhagic Dengue)</td>
</tr>
<tr>
<td>virus</td>
<td>HF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Kyasanur forest</td>
<td>Kyasanur Forest</td>
<td>India, Western China</td>
<td>Ticks, infected monkeys</td>
<td>3–8</td>
<td>Supportive</td>
<td>Suggestion of nosocomial spread. Consider isolation and full PPE. High risk of infection in laboratory setting—full PPE</td>
<td>Biphasic—first phase: fever, myalgia, headache, diarrhoea, vomiting, haemorrhage second phase: fever, encephalitic symptoms CFR=0.4–2.5%</td>
</tr>
<tr>
<td>disease virus</td>
<td>disease</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alkhurma hemorrhagic</td>
<td>Alkhurma HF</td>
<td>Saudi Arabia, Egypt</td>
<td>Ticks, mosquitoes, infected animals (sheep, camels), unpasteurised milk</td>
<td>3–8</td>
<td>Supportive</td>
<td>No known interhuman transmission—Standard IP&amp;C measures</td>
<td>Fever, headache, myalgia/arthritis, vomiting, haemorrhage, encephalitis CFR=Up to 25%</td>
</tr>
<tr>
<td>fever virus (variant of</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Kyasanur)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

CFR, case fatality rate; IP&C, infection prevention & control; PPE, personal protective equipment.
the virus without being affected by it. Non-human primates also host the virus, but have high mortality rates. Once the Ebola virus enters human circulation, transmission is maintained through human-to-human infections.

In the current outbreak of EVD in West Africa mortality rates have been particularly high (approximately 80%) in children under the age of 5 years, with mortality in infants under 1 year of age close to 90%. In contrast children aged 10–15 years have a low mortality rate when compared with all age groups (approximately 50%).

The incubation period for the virus is from 2 days to 21 days. Detection of Ebola viral RNA by PCR in peripheral blood is likely to occur within the first week of illness. The virus is more likely to be isolated from the blood late in the course of the disease. 

The virus is often spread through hospital contacts. When present, the virus is spread by direct contact with the blood or body fluid of an infected patient. EVD is not spread by air. 

Transmission of EVD is high in areas where health care and sanitation are poor. The virus can be spread to a household member by maintaining direct contact with the virus. 

The incubation period for the virus is from 2 days to 21 days. Detection of Ebola viral RNA by PCR in peripheral blood is likely to occur within the first week of illness. The virus is more likely to be isolated from the blood late in the course of the disease. 

Transmission of EVD is high in areas where health care and sanitation are poor. The virus can be spread to a household member by maintaining direct contact with the virus. 

Management of patients with EVD is generally supportive, including symptomatic treatment of fever, pain and vomiting. Profound hypovolaemia can occur from profuse diarrhoea and vomiting. While oral fluids may normally be preferred, in the context of wet symptoms in EVD, intravenous fluids are essential. Remaining ahead of the fluid loss appears key to improved outcomes and is likely to reduce the risk of prerenal failure. Due to the copious diarrhoea, electrolyte abnormalities are common and regular monitoring is essential where available. 

Due to the copious diarrhoea, electrolyte abnormalities are common and regular monitoring is essential where available. 

A key feature of EVD is its propensity to cause thrombocytopenia, liver enzyme derangement and disseminated intravascular coagulation, all of which may result in profuse bleeding. These blood parameters should be monitored regularly, to expedite timely intervention with platelets, vitamin K or fresh frozen plasma. It is important to remain vigilant for signs of evolving bacterial sepsis, particularly Gram-negative sepsis from translocation of gut pathogens. There is no proven treatment for EVD, however outcomes of early trials are anticipated in the near future. WHO advises the use of convalescent plasma in children with EVD on compassionate grounds at a dose of 10 mL/kg whole blood or plasma with close monitoring of efficacy. 

There are unpublished clinical trials into the use of monoclonal antibody products (ZMAPP) and antiviral medications such as favipiravir and TKM-Ebola. Although a dose regimen for favipiravir use in children has been described, there is no published evidence to support the efficacy of this or any other antiviral for treatment of Ebola in children. Ribavirin does not appear effective in EVD. 

There are unpublished clinical trials into the use of monoclonal antibody products (ZMAPP) and antiviral medications such as favipiravir and TKM-Ebola. Although a dose regimen for favipiravir use in children has been described, there is no published evidence to support the efficacy of this or any other antiviral for treatment of Ebola in children. Ribavirin does not appear effective in EVD. 

Clinical trials of two potential vaccinations are ongoing.

1. Which countries have you or your child travelled to in the last 3 weeks and to which regions in these countries?
2. Were you in rural areas with very basic accommodation?
3. Did you work in or did your child attend a healthcare facility in this location?
4. Have you attended any funerals in the last 3 weeks? Did you or your child have any direct contact with the body of the deceased or with someone who had handled the body?
5. Have you or your child come into physical contact with anyone who was unwell in the last 3 weeks?
6. Have you or your child visited any enclosed areas where bats may have lived such as mines or caves?
7. Have you or your child come into contact with any monkeys or other 'bush' animals? Have you or your child prepared or eaten any 'bush meat' such as bat, monkey or forest antelope?
8. Have your children been in an environment where ticks may be present such as in forests or long grass? Have they sustained a tick bite or crushed a tick with their bare hand/foot?
9. Have you or your child visited an abattoir or tannery or been involved in the slaughter of animals?

### Table 2 Clinical features of viral haemorrhagic fevers in children

<table>
<thead>
<tr>
<th>General/systemic</th>
<th>Gastrointestinal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fever (90%)</td>
<td>Loss of appetite (73%)</td>
</tr>
<tr>
<td>Headache (59%)</td>
<td>Abdominal pain (47%)</td>
</tr>
<tr>
<td>Myalgia (38%)</td>
<td>Vomiting (62%)</td>
</tr>
<tr>
<td>Arthralgia (35%)</td>
<td>Diarrhoea (60%)</td>
</tr>
<tr>
<td>Fatigue/weakness (79%)</td>
<td>Jaundice (11%)</td>
</tr>
<tr>
<td>Difficulty swallowing (20%)</td>
<td></td>
</tr>
<tr>
<td>Sore throat (16%)</td>
<td>Respiratory</td>
</tr>
<tr>
<td>Hiccups (7%)</td>
<td>Difficulty breathing/tachypnoea (20%)</td>
</tr>
<tr>
<td>Rash (6%)</td>
<td>Cough (31%)</td>
</tr>
<tr>
<td></td>
<td>Chest pain (29%)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Haemorrhagic</th>
<th>Neurological</th>
</tr>
</thead>
<tbody>
<tr>
<td>Unexplained bleeding (10%)</td>
<td>Confusion (10%)</td>
</tr>
<tr>
<td>Epistaxis (1.3%)</td>
<td></td>
</tr>
<tr>
<td>Bleeding from gums (2.1%)</td>
<td>Unresponsive/obtunded (5%)</td>
</tr>
<tr>
<td>Bleeding from eyes</td>
<td>Seizures (later stages)</td>
</tr>
<tr>
<td>Haematemesis (1.4–1.9%)</td>
<td></td>
</tr>
<tr>
<td>Melaena/haemorrhagic diarrhoea (3.2%)</td>
<td>Ocular</td>
</tr>
<tr>
<td>Vaginal bleeding (0.7%)</td>
<td>Conjunctivitis (22%)</td>
</tr>
<tr>
<td>Haematuria (0.5%)</td>
<td>Eye pain (6%)</td>
</tr>
<tr>
<td>Bleeding from injection site (0.5%)</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Common haematological/biochemical parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leucopenia</td>
</tr>
<tr>
<td>Thrombocytopenia</td>
</tr>
<tr>
<td>Transaminitis</td>
</tr>
<tr>
<td>Coagulopathy—prolonged PT, APTT and reduced fibrinogen</td>
</tr>
</tbody>
</table>

Percentage figures are symptom incidence specifically for children presenting during the current Ebola virus epidemic in West Africa.
Breast feeding is contraindicated in EVD, as it is a potential mechanism of transmission of virus. The risk of transmission is from infected mother to infant as well as from infected infant to mother. It is therefore recommended that, where appropriate formula milk is available, breast feeding should cease and formula milk be provided. Where mother and infant are infected, breast feeding should continue. There have been reports of transmission of virus to infants through the breast milk of lactating Ebola survivors (personal communication), but it is unclear how long Ebola virus may persist in breast milk following a woman’s survival from the disease. In these circumstances, where testing facilities are available, we suggest breast feeding of an unaffected infant by a mother who has survived EVD should not recommence until two consecutive PCR tests at least 48 h apart have determined the breast milk to be free of virus.

Neonates are an extremely vulnerable group in the context of EVD. Unborn infants who contract the disease in utero from an infected mother are highly viraemic and often stillborn. The placenta in these cases is highly infectious and the management of pregnant women in labour, or those miscarrying, is extremely high-risk and often complicated by significant haemorrhage. Assisting with delivery should only be undertaken by those specifically trained in this area. The few infants born alive rarely live beyond a few days, there is only one report of such an infant surviving. This is a recent case born at a treatment facility in Guinea, who received treatment with ZMAPP and an experimental antiviral, GS5734. Neonates who contract Ebola during the first weeks of life from an infected parent or sibling also have very high mortality rates. In either circumstance, for the best chances of survival, treatment should be started promptly, with due consideration to the use of experimental therapies under compassionate care guidance. Testing should be performed on the basis of exposure, even in the absence of symptoms, to facilitate early intervention. With regards to poor outcomes however, due consideration should be given to the benefits of escalation of care in neonatal EVD, particularly in the case of liveborn infants to a mother infected with EVD prior to delivery.

Most pregnant women with EVD tend to miscarry or deliver their infant in the acute phase of the disease. There is evidence to suggest that women surviving Ebola with an intact pregnancy pose a significant risk to healthcare workers when they do deliver and that the products of conception and the infant may remain infected. In this circumstance the mother should be delivered at a facility specifically designed for managing pregnant women infected with EVD and the infant, whether liveborn or stillborn should be assumed to be infected. If liveborn, the infant should be monitored in a high-level isolation unit (HLIU) for a minimum of 21 days and consideration given to early testing and treatment even in the absence of symptoms.

There is some evidence for the spread of Ebola virus from individuals with mild or subclinical EVD. A recent report describes sexual transmission of EVD via semen, from a survivor who had mildly symptomatic EVD 6 months previously. A recent WHO situation report also documents a case of a stillborn infant in Sierra Leone born to a well mother with no known history of infection; the infant tested positive for Ebola virus and serological testing of the mother revealed antibodies to Ebola. These cases are relevant for the paediatrician in relation to the potential for asymptomatic infection in mothers with spread to their unborn or newborn infants through in utero transmission or possibly through breast milk. Similarity the potential for sexual transmission should be considered in

Figure 1 (A) Map of Filoviridae viral haemorrhagic fever (VHF). (B) Map of Arenaviridae viral haemorrhagic fever. (C) Map of Bunyaviridae viral haemorrhagic fever. (D) Map of Flaviviridae viral haemorrhagic fever.
adolescents with a history of EVD. It is important that comprehen- sive information regarding the use of condoms is provided to young men at discharge as well as consideration given to the monitoring of bodily fluids for viral persistence.

There are reports of post-Ebola sequelae,39 including critical illness or life-limiting complications, and of viral persistence in semen, intravascular fluid, cerebrospinal fluid (CSF), vaginal secretions and breast milk.40-42 There is no clear data as to the proportion of survivors suffering from sequelae or viral persistence in bodily fluid in the most recent epidemic; this crucial research is currently underway. The most common sequelae appear to be eye disease (usually an acute uveitis which can lead to cataracts or blindness),41 headache, chronic joint pain, chronic fatigue and sleep disturbance.42 There has been one reported case of re-emergence of the virus in CSF resulting in viral meningitis (UK national press). Most of the current reports pertain to adults, however increasingly similar sequelae in children are becoming apparent, particularly eye disease resulting in cataracts (personal communication).

Marburg haemorrhagic fever
Marburg haemorrhagic fever (MHF) is caused by Marburg murine viruses. It was first identified in 1967 when a group of cases occurred in Marburg, Germany and simultaneously in Belgrade, former Yugoslavia, linked to imported African green monkeys. Since that time there have been three significant outbreaks of Marburg (Uganda, DRC and Angola), and a handful of isolated cases or small clusters (two to three cases) in Kenya, DRC, Uganda and Zimbabwe/South Africa.44 See figure 1 for regions of the world known to be at risk from Marburg virus.

Most cases of MHF appear to be linked to work in or visits to caves or mines.3,7 The reservoir is unclear but is widely considered to be fruit bats.7 Human infection arises either through contact with bat secretions or other infected wildlife including non-human primates and forest antelopes. There is then subsequent human-to-human spread.3 The number of paediatric MHF cases has been limited, and there are few relevant publications.25

The incubation period for MHF is similar to EVD, ranging from 2 days to 21 days.3 Presentations in adults are similar to those of EVD.8,9 Management of MHF is supportive and similar principles to the management of EVD apply. There are currently no specific therapies, though therapies being trialled for EVD may be of potential benefit in MHF. Early studies showed no benefit of ribavirin.5 e 33

ARENAVIRIDAE
Lassa fever and other Arenaviridae
The Arenaviridae family incorporates seven RNA viruses known to cause haemorrhagic fever in humans. There is wide geographical distribution throughout South America and Africa, with highest endemity in West Africa. Arenaviruses are transmitted to humans through aerosolisation of excreta from rodents. Once transmitted to a human host, transmission is maintained through human-to-human and nosocomial interaction, although this appears less common than in EVD and MHF.5 The incubation period varies between species (see table 1) but overall is considered to be 5–21 days.5

Although true incidence data is lacking Lassa fever is estimated to affect approximately 300 000 people in West Africa each year with an estimated 5000 deaths.16 Lassa may present with similar clinical features to other VHFs (table 2), although in the advanced form can cause significant oedema and pleural effusions. It has less severe haemorrhagic elements when compared with EVD and MHF.10 Paediatric Lassa fever is not well described; a paper from 1987 gives mortality rates of 27% in Liberia,45 however this is likely an inflated figure as a significant proportion of milder cases will have been missed in this context. Paediatric Lassa fever may commonly present as a non-specific febrile illness, but a severe form known as ‘swollen baby syndrome’ has also been described in Liberia. This is a condition consisting of gross oedema, abdominal distension and bleeding, with a very high mortality rate.3,45 46 Ribavirin is effective in Lassa fever and if given within the first few days of symptom onset can significantly reduce mortality.47 Alongside ribavirin, supportive therapy as described for EVD above is the mainstay of treatment.

Other arenaviruses are listed in table 1 and consist of the South American Arenaviruses and Lujo virus, a recently identified virus in Zambia that caused a small cluster of nosocomial infections. There is limited paediatric data available for these viruses highlighting a significant gap in medical knowledge in this area. Large studies of the efficacy of ribavirin have not been conducted in either adult or paediatric populations, but based on its mechanism of action it is thought to be beneficial in all arenaviral infections.3,21,22,47 See figure 1 for regions of the world known to be at risk from the Arenaviridae VHF viruses.

BUNYAVIRIDAE
Bunyaviridae are vectorborne infections, generally transmitted by arthropods, including ticks and mosquitoes, or through contact with rodents.

Crimean-Congo haemorrhagic fever
Crimean-Congo haemorrhagic fever virus is an RNA virus in the Bunyaviridae family. It has a widespread geographical distribution causing outbreaks of Crimean-Congo haemorrhagic fever (CCHF) throughout Africa, eastern Europe, the Middle East, central and western Asia (see figure 1 for regions known to be at risk). It has become increasingly common in recent years in Turkey, South-West Russia and Ukraine.48 Children are most commonly infected through tick bites, but thereafter there is a risk of nosocomial spread within healthcare settings. It has an incubation period of 1–13 days3 and presents in a similar fashion to other VHFs (table 2), although with a higher preponderance for haemorrhagic symptoms. Children who make up approximately 20% of all cases, present in a similar fashion to adults, but appear to have a milder course of disease with better survival rates (>90%).3,11–13 Table 1 further identifies some of the more disease-specific clinical features.

Treatment is currently supportive, although there is ongoing discussion on the benefits of the WHO recommendation to use ribavirin.13,14,40,50 A case example is found in box 3, highlighting the clinical features of CCHF and providing a comparison to the clinical case of EVD; further specific details of CCHF are identified in table 1.

FLAVIVIRIDAE
The Flaviviridae family of RNA viruses includes the Yellow Fever and Dengue viruses, which can present with haemorrhagic symptoms. These will not be discussed as there is no clear demonstration of nosocomial or person-to-person spread, although there have been reports of transmission of Dengue virus through needlestick injuries.15,16 Lesser-known VHF viruses in this family are the Omsk, Kyasanur and Alkhurma haemorrhagic fever viruses, which are thought to spread from bites of infected ticks, although other methods of transmission have been postulated (see table 1). Outbreaks of these diseases have been small, but
Box 2  Case study—Ebola virus disease (EVD)

▸ A 4-year-old African girl presents with her mother to the emergency department of a children’s hospital with 2 days of fever, having returned from visiting family in the Democratic Republic of Congo. Seven days before, they attended a funeral of a family friend who died unexpectedly. The mother denies any involvement in the burial preparation of the body.

▸ The child developed diarrhoea and vomiting approximately 24 h ago, and is increasingly weak and floppy, complaining of abdominal pain and headache. On examination she appears weak and lethargic, with tachycardia, sunken eyes and reduced skin turgor. Abdominal examination reveals generalised tenderness. Blood testing detects a potassium of 3.3 mmol/L and a urea of 7.5 mmol/L. She has a normal full blood count and C reactive protein. A malaria film is negative.

▸ She is admitted to a cubicle where basic barrier nursing measures are applied. She deteriorates overnight, remaining pyrexial up to 40.5°C. The following morning her diarrhoea is noted to have occasional streaks of fresh red blood. Repeat blood tests reveal a platelet count of 80×10⁹/L, WBC 18×10⁹/L, potassium 3.1 mmol/L, urea 8.5 mmol/L, creatine 105 μmol/L, ALT 450 IU and CRP 30 mg/L. Because of the travel history, and presence of bleeding with low platelets, concerns are raised over the possibility of a viral haemorrhagic fever and advice is sought from the regional paediatric infectious disease team, and the national Rare and Imported Fever Service. Ebola virus is confirmed by PCR at the national reference laboratory.

▸ She is transferred to a high level isolation unit (HLIU) but deteriorates, developing multiorgan failure. On day 6 she arrests and resuscitation attempts are unsuccessful. Her family is monitored for signs of the disease, and her mother and 10 year-old brother develop a fever and test positive for EVD.

▸ The clinical team are monitored for 21 days following exposure. On day 11, a nurse who cared for her prior to her HLIU admission, and without full personal protective equipment, develops a fever and tests positive for EVD.

Box 3  Case study—Crimean-Congo haemorrhagic fever (CCHF)

▸ A 7-year-old boy returned from a holiday in Turkey 7 days ago. He presents to the children’s assessment unit at a district general hospital with a fever of 39°C for 24 h, muscle pain and headache. He is vomiting, with streaks of blood. His mother reports that he played in fields and woods while in Turkey.

▸ Clinical assessment reveals an acutely unwell, pale child with tachycardia. He develops melaena and epistaxis. Blood results reveal a leucopenia of 1.9×10⁹/L WBC, platelets of 6×10⁹/L, ALT of 300 IU, prothrombin time of 17 s and activated partial thromboplastin time of 78 s. He is treated with fresh frozen plasma, a platelet transfusion and vitamin K, following which his bleeding resolves. An infection screen at this stage reveals a positive PCR result for CCHF virus. He is transferred to a high-level isolation unit.

▸ Over the course of the next 48 h he improves gradually. Repeat blood tests on day 7 are negative for CCHF on PCR. He makes a full recovery. The rest of his family remain well. The medical and nursing staff at the district general hospital are monitored for 14 days but none shows signs of developing CCHF.

nosocomial spread has been suggested although not demonstrated in Kyasanur forest disease (KFD). Laboratory transmission has occurred in Omsk HF and KFD where transmission through the handling of infected monkey carcasses has also occurred. There are few documented paediatric cases of Alkhurma and KFD, but approximately a third of cases of Omsk HF occur in the paediatric population, where there is a biphasic disease pattern with a high incidence (41%) of meningitic/encephalitic symptoms in the second phase. Treatment is supportive with no evidence for ribavirin. See figure 1 for regions of the world known to be at risk from Flaviviridae VHF and table 1 for clinical details of each disease.

SAFE MANAGEMENT OF PAEDIATRIC PATIENTS WITH VHF

The fundamental principle of initial management is an early and careful risk assessment, made in conjunction with regional and national paediatric infectious diseases and public health teams, together with proportionate patient isolation and use of PPE to reduce risks of onward transmission, pending the outcome of VHF testing. The approach, including the challenges of isolating and safely transporting the child, is discussed in more detail by Herberg JA et al.

Patients with confirmed hazard group 4 VHF must be cared for in units with appropriate isolation facilities, by a designated, trained team. The features of such a unit, the appropriate PPE for healthcare workers and the management of such patients are described in the document ‘Management of Hazard Group 4 viral haemorrhagic fevers and similar human infectious diseases of high consequence’ produced by the ACDE.

Provision of intensive care including ventilation and haemofiltration to patients with VHF may increase the risks of onward transmission to healthcare workers, but several returning healthcare workers with EVD have made a good recovery after receiving intensive care. There are no reports of children with VHF receiving intensive care and weighing of risks and benefits must be made on a case-by-case basis. Current UK Department of Health guidance suggests intensive care procedures should only be undertaken where there is clear benefit to the patient, at an HLIU where protection of healthcare workers is maximised.

SUMMARY

VHFs include varied, unrelated viruses with the potential to cause haemorrhagic symptoms as part of the disease process. Not all patients with VHF will manifest haemorrhagic symptoms and early presentations of disease may be very non-specific. Paediatric presentations may be even more generic when compared with presentations of other infectious diseases and are currently poorly described in the literature, indicating a significant need for informed research in this area. The importance of taking a detailed, specific history to identify the possibility of such a pathogen to the attending clinician must be emphasised due to the high risk of nosocomial spread. Early prompt treatment by a specialist team in an HLIU improves patient outcome and protects healthcare workers.
Acknowledgements The authors thank Dr Andrew Simpson for his advice and review of the paper.

Funding NEM receives funding from the Wellcome Trust Institutional Strategic Support Fund and the Institute of Global Health Innovation at Imperial College London; JAH is supported by NIHR Imperial College BRC and EU fp7 funding.

Competing interests None declared.

Provenance and peer review Commissioned; externally peer reviewed.

REFERENCES

Viral haemorrhagic fever in children

Nathalie E MacDermott, Surjo De and Jethro A Herberg

Arch Dis Child 2016 101: 461-468 originally published online January 19, 2016
doi: 10.1136/archdischild-2014-307861

Updated information and services can be found at:
http://adc.bmj.com/content/101/5/461

These include:

References
This article cites 47 articles, 2 of which you can access for free at:
http://adc.bmj.com/content/101/5/461#BIBL

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections
Articles on similar topics can be found in the following collections
ADC Global child health (48)
Editor's choice (147)

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/